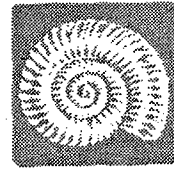


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IKU

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REPORT TITLE/ TITTEL			
HYDROCARBON CHARACTERISATION OF WELL 31/5-2.			
CLIENT/ OPPDRAGSGIVER			
Saga Petroleum a.s.			
RESPONSIBLE SCIENTIST/ PROSJEKTANSVARLIG			
Susanne Betts			
AUTHORS/ FORFATTERE			
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SUMMARY/ SAMMENDRAG

One oil and one gas sample from 31/5-2 were analysed.

The oil (B-8116) is a mature biodegraded paraffinic oil.

The gas (B-8971) associated with the oil is 80% methane and shows depletion of n-alkanes of carbon number above C₄ (relative to branched and cyclic compounds of similar molecular weight).

KEY WORDS/ STIKKORD

31/5-2

TROLL

Hydrocarbon Characterisation

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INTRODUCTION

IKU was supplied with one oil and one gas sample from this well, the two samples together make up a representative reservoir fluid.

The following information on collection of the oil and gas samples was supplied by GECO.

Flash of reservoir fluid to stock tank conditions

Flash conditions	:	250 barg, 67.6 ⁰ C to atmosphere and 15 ⁰ C.
Gas oil ratio	:	60.5 sm ³ /m ³ .
Bo at 250 barg	:	1.158 m ³ /m ³ .
Bo at bubble point	:	1.170 m ³ /m ³ .
Density of oil at 15 ⁰ C	:	894.4 kg/m ³ .
Molecular weight of oil	:	251.
Standard conditions	:	for gas volumes = 15 ⁰ C and 1 atm. for oil volumes = 15 ⁰ C and atmospheric pressure.

Saga supplied information on reservoir temperature (67.6⁰C) and pressure 156 barg.

The following analyses were carried out on the oil sample:

- Measurement of API gravity of the oil.
- C₂-C₈ hydrocarbon characterisation.
- Measurement of the fraction boiling under 210⁰C.
- Fractionation of the fraction boiling over 210⁰C by MPLC (medium pressure liquid chromatography).
- Molecular sieving of the saturated fraction into normal and branched/cyclic alkanes.
- Gas chromatography of the saturated/branched and cyclic/aromatic fractions.
- GC-MS of saturated fraction (m/z 191, 217 and 218).
- GC-MS of aromatic fraction (m/z 231 and 253).
- δ¹³C isotope analysis of saturated and aromatic hydrocarbon fractions.

EXPERIMENTAL PROCEDURES

C₂-C₈ analysis

C₄-C₁₀ analysis was carried out on a HP 5880 A gas chromatograph equipped with a 50m x 0.2mm (I.D.) fused silica column coated with OV-101. Helium was used as carrier gas at 1ml/min. The inlet split ratio was 1:60. The temperature program was 35^oC (5 min) - 8^oC/min. - 200^oC (2 min.) which gave good resolution from C₃ upwards. Quantitation was carried out using the same standard gas as for C₁-C₅+ analysis.

Total organic carbon

Bulk samples were crushed in a mortar. Aliquots of the samples were then weighed into Leco crucibles and treated three times with hot 10% HCl to remove carbonate, and washed 4 times with distilled water to remove traces of HCl. The crucibles were then placed on a hot plate and dried for 24 hours. The total organic carbon (TOC) content of the dried samples was determined using a Leco CR12 carbon analyser.

Evaporation of the light components in fluid samples

Prior to chromatographic separation of oil/condensate samples, the fractions boiling below 210^oC were removed by heating the samples to constant weight at 210^oC is obtained. The heating is performed at atmospheric pressure.

The fraction of light components is determined as the weight difference between the original sample and the amount that is left after the heating.

Chromatographic separation

The fraction of the oil boiling above 210^oC was separated into saturated fraction, aromatic fraction and non hydrocarbon fraction using a MPLC system with hexane as eluant (Radke et al., Anal. Chem., 1980). The various fractions were evaporated on a Buchi Rotavapor and transferred to glass vials and dried in stream of nitrogen.

Molecular sieve adsorption

The sample containing 2mg of n-alkanes was dissolved in 35ml of cyclohexane and 1gs of Molecular Sieve pellets (5A) which had been activated at 300°C in 24 hours, were added. This mixture was then refluxed for about 24 hours. While the solution was still hot, the sieve pellets were removed from the solution by filtering. The solvent was then removed on a Buchi Rotavapor. GC analysis were performed on the samples, using the same conditions as for the other GC analysis.

The normal alkanes were recovered from the Molecular Sieve pellets by destruction of the pellets with hydrofluoric acid. The solution was extracted with boric acid and hexane, and the solvent was then removed on a Buchi Rotavapor. GC analysis were performed on the samples, using the same conditions as for the other GC analysis.

Gas chromatographic analysis

The C₂-C₈ hydrocarbons of the oil were determined on a Carlo Erba Fractovap GC. The column used was a 30m fused silica capillary column coated with SE-54. The temperature program applied was 50°C (2min.) to 180°C at 4°C/min.

The saturated, the branched/cyclic and the aromatic hydrocarbon fractions were each diluted with n-hexane and analysed on a HP 5730A. The GC is equipped with a 15m DB-1 fused silica column and hydrogen (ca. 2.5 ml/min.) is used as carrier gas. Injections are performed in split mode (split ratio 1:10). The temperature program applied is 80°C (2 min.) to 280°C at 4°C/min.

The data processing for all the GC analyses was performed on a VG Multichrom lab data system.

Gas chromatography - mass spectrometry (GC-MS)

GC-MS analyses were performed on a VG Micromass 70-70H GC-MS-DS system. The Varian Series 3700 GC was fitted with a fused silica OV-1 capillary column (30m x 0.3mm i.d.). Helium (0.7kg/cm²) was used as carrier gas and the injections were performed in split mode (1.5µl, split ratio 1:15). The GC oven was programmed from 70°C to 280°C at 4°C/min. after an ini-

tial isothermal period of 2 minutes.

The saturated hydrocarbons were analysed in multiple ion mode (MID) at a scan cycle time of approximately 2 secs. Full data collection was applied for the aromatic hydrocarbons at a scan time of 1 sec/decade. The mass spectrometer operated at 70eV electron energy and an ion source temperature of 200°C. Data acquisition was done by VG data systems.

Peak identification was performed applying knowledge of elution patterns in certain mass chromatograms. Calculation of peak ratios was done from peak height in the appropriate mass chromatograms.

$\delta^{13}\text{C}$ isotope analysis

The $\delta^{13}\text{C}$ isotope analysis was performed by mass spectrometry at Institute for Energy Technology (IFE) in Oslo according to their method. Their reference value for the standard NBS-22 is -29.8.

Analyses carried out on the gas sample

Gas analysis

$\text{C}_1\text{-C}_{10}$ analysis was carried out on an HP 5880 gas chromatograph equipped with a 50m x 0.2mm (I.D.) column fused silica column coated with OV 101. Helium was used as a carrier gas at 1ml/min. The inlet split ratio was 1:50. The temperature program was -10°C for 2mins., 10°C/min to 160°C, 160°C for 5 mins.

Quantitation was carried out using a standard gas containing methane, ethane, propane, n-butane, n-pentane and n-hexane. In addition a natural gas standard obtained from Norsk Hydro was used.

RESULTS

API gravity

The specific gravity of B-8116 at 60^oF = 0.8934
API^o gravity = 26.8^o

This is a medium gravity oil. The gravity is probably higher now than it was originally due to biodegradation and consequent loss of lighter hydrocarbons.

Fraction boiling below 210^oC (see table 1)

Oil composition (see tables 2 and 3)

A biodegraded paraffinic oil.

C₂-C₈ hydrocarbons

The distribution of C₂-C₈ hydrocarbons shows a similar trend to that seen in the nC₁₅+ fraction. The n-alkanes have been depleted by biodegradation. Table 4 and figure 1 show the relative weights and percentage weights of C₂-C₈ compounds present in the sample. The light hydrocarbon ratios used by Thompson, 1979 to estimate type of source and maturity are probably not valid for this sample because it is biodegraded. In addition to this it has not been possible to determine some of the necessary compounds, e.g. heptane.

Gas analysis

The results have been tabulated (table 5). A similar trend of depletion of n-alkanes is also seen in the gas sample (above C₄). The iC₄/nC₄ ratio is approximately 6.5 and iC₅/nC₅ is 4.8. A similar predominance of branched pentanes over n-hexane, and branched hexanes over n-heptane is seen.

Isotopic analyses

The results (table 7) are rather unusual as it is normally the saturated fraction that is the more depleted of the two. The difference is fairly

large and hard to account for. The samples were analysed twice to check but with the same result.

Gas chromatography results

Saturated fraction

The low abundance of lower molecular weight alkanes (below C_{16}) and predominance of isoprenoid alkanes suggests that the oil is biodegraded. The total alkane range is C_{13} to C_{38} with nC_{16} to nC_{30} being present in the greatest abundance. The pristane/ nC_{17} ratio is high, 2.15 but this probably indicates biodegradation rather than low maturity. The abundance of higher i.e. over C_{20} alkanes is noteworthy. It is possible that the C_{30} plus alkanes are derived from the biodegrading microorganisms themselves. Another possibility is the presence of a high molecular weight additive although this seems unlikely and the absence of higher molecular weight compounds in the aromatic fraction chromatogram indicates that this is not the case. This range of alkanes is also seen in some of the immature source rock extracts from 31/5-2, e.g. from 1976m (B-226, p.68). Whether the abundance of high molecular weight alkanes persists to higher maturities in source rocks outside the area of the well is not known.

Branched and cyclic alkanes

The chromatogram is dominated by isoprenoid alkanes which are ubiquitous in extracts and oils and therefore give little specific information on source. The pristane/phytane ratio is fairly high 1.9 and is within the range to the ratio from the 31/5-2 rock extracts below 2000m (1.4-3.4).

Aromatic fraction

The chromatogram is dominated by methyl and dimethyl naphthalenes with relatively little higher molecular weight material. The distribution of the methyl naphthalenes (i.e. with 2 methyl naphthalene dominant) and the absence of high molecular weight material indicates that the oil is mature.

Molecular ratios from terpane and sterane mass chromatograms applied as maturity and source characteristic parameters

Geochemical fossils or biological marker components are characteristic of the type of organic matter present at the time the sediments were

deposited. The biological isomers of these components undergo changes due to increased maturity in particular, but also to a certain degree caused by migration and weathering processes.

Source characteristic parameters

In the m/z 191 mass chromatograms, representing terpanes, the hopanes and moretanes are the major components in most extracts and oils. Of the hopanes the C₂₇ and C₂₉-C₃₅ homologs are ubiquitous, while the C₂₈ bisnorhopane is believed to be typical of certain types of source rocks. This is also the case for the component, probably gammacerane, sometimes seen to coelute with the 22S isomer of the C₃₁ 17 α (H)-hopanes (H). In the sterane mass chromatograms, m/z 217 and m/z 218, the molecular weight distribution of the C₂₇-C₂₉ regular steranes is believed to be representative of the original input of organic matter. The highest molecular weight compounds, the C₂₉ steranes, represent organic matter of terrestrial origin, while the lower molecular weight analogs originate from more marine type environments.

Maturity dependant parameters

The biological isomers of the hopanes, the 17 β (H), 21 β (H)-hopanes, undergo structural changes during the maturation process. The isomerisation reactions are thought to be produced via the 17 β (H), 21 α (H)-hopanes (moretanes) to the most stable 17 α (H), 21 β (H)-hopanes. At equilibrium 100% of the 17 α (H)-hopanes are seen. The ratio $\alpha\beta/\alpha\beta+\beta\alpha$ is used to describe this reaction. In the extended hopanes (\geq C₃₁), the thermally stable S configurations at C-22 become increasingly more abundant as compared to the biological preferred R configurations at increased maturity level. The equilibrium ratio is approximately 60% of the 22S configuration. Another ratio that is known to change with maturity is the Tm/Ts (Seifert et al., 1978) of the C₂₇ hopanes. The maturable 18 α (H)-trisnorhopane (Tm) is reduced in intensity relative to the more stable 17 α (H)-trisorhopane (Ts), causing the Tm/Ts to decrease at increased maturity. This ratio is also believed to be source dependant, and this should be born in mind when applying the ratio for maturity comparison. The amount of tricyclic terpanes is also to a certain extent seen to be maturity dependant.

Two isomerisation reactions taking place in the steranes are most commonly applied for maturity assignments from the m/z 217 mass chromatograms. The biologically preferred 14 α (H), 17 α (H)-isomers of the regular steranes is transformed to the thermally stable 14 β (H), 17 β (H)-steranes, the % $\beta\beta$ approaching 75% at equilibrium. An equilibrium concentration of 50% is seen of the stable S configuration at C-20 as opposed to the 100% of the biological 20R epimer (Mackenzie et al., 1980). The abundance of rearranged steranes increased with increasingly maturity.

One of the reactions taking place at an early stage of diagenesis is the aromatisation of steranes, leading to the formation of mono- and tri-aromatic analogs. This process is measured as the abundance of tri-aromatic relative to mono-aromatic compounds (% tri/tri + mono) in the m/z 231 and 253 mass chromatograms, respectively. In addition the degree of side chain cracking, as %C₂₀/C_{26, 27} and %C₂₁/C_{28,29} respectively, is applied. These cracking processes are also taking place during early diagenesis, and are used for maturity assignment together with the previously mentioned ratios.

Migration and weathering

The effect on the geochemical fossils of migration and weathering, is less apparent than the maturity induced changes. Migration is believed to cause an increase in the relative amounts of rearranged and 14 β (H), 17 β (H) regular steranes (Seifert and Moldowan, 1978, 1981). Severe biological alteration leads to the formation of desmethyl-hopanes (Seifert and Moldowan, 1979).

GC-MS analysis of saturated steranes and terpanes

The oil sample was analysed for the relative distribution of steranes (m/z 217, 218) and terpanes (m/z 191). Mass chromatograms and tabulated data are presented in Figures 6 and 7 and Tables 8 and 9, respectively.

The oil is seen to contain biomarkers of high maturity, all the isomerisation reactions having reached equilibrium. A certain content of bisnor-hopane (Z in m/z 191) is seen in the sample. This does not, however, necessarily mean that the source rock for this oil contains any bisnor-hopane, since it is known that the relative abundance of this compound is also dependant on maturity. The relative molecular weight distribu-

tion of regular steranes, indicates a high proportion of terrestrial input from the high abundance of C₂₉ steranes.

GC-MS analysis of aromatic hydrocarbons

Total ion and mass chromatograms representing aromatic hydrocarbons are presented in Figure 8, while maturity ratios from aromatic steranes are presented in Tables 8 and 9.

This oil sample contains a high proportion of low molecular weight naphthalenes compared to phenanthrenes. Front end biased distribution of alkylated mono-aromatic hydrocarbons is seen, and the relatively low abundance of aromatic steranes suggests mature hydrocarbons.

CONCLUSION

From the analyses carried out the sample provided appears to be a mature biodegraded paraffinic oil.

The associated gas sample is largely methane (80%) and is noticeably depleted in C_4+ n-alkanes also probably the effect of biodegradation. Some of the methane may be bacterial in origin but as isotopic study of the gas was not requested the relative proportions of biogenic v.s. thermogenic methane cannot be established.

Table 1. Fraction boiling below 210°C.

Oil 31/5-2

<u>Starting wt. (g)</u>	<u>after evaporation (g)</u>	<u>wt loss (mg)</u>
2.0801	1.8713	84.0
% boiling under 210°C	=	4%



Percentage composition of the main fractions in the oil sample

I	:	:	Sat	:	Aro	:	HC	:	SAT	:	Non HC	:	HC	I
I	IKU-No	:	DEPTH	:		:		:		:		:		I
I	:	:	EOM	:	EOM	:	EOM	:	Aro	:	EOM	:	Non HC	I
I	:	(m)	:	:	:	:	:	:	:	:	:	:	:	I
I														I
I	B 8116		44.3		22.6		66.9		196.2		33.1		202.6	I
I	Oljeproeve													I

DATE : 1 - 11 - 84.

Table 4. C₂-C₈ hydrocarbons, μg and %.

B-8116

31/5-2 C₂ to C₈ hydrocarbons

$\rho = 0.8934$

	<u>μg</u>	<u>%</u>
C ₂		
C ₃		
MC ₃	0.48	0.18
nC ₄	0.30	0.11
MC ₄	0.88	0.33
NC ₅	0.32	0.12
CyC ₅ + 2.3DMC ₄	1.08	0.40
2MC ₅	1.16	0.43
3MC ₅	0.88	0.33
nC ₆	2.72	1.01
MCyC ₅	3.94	1.47
benzene	-	
CyC ₆	6.16	2.30
2MC ₆	-	
2.3DMC ₅	0.73	0.27
3MC ₆	1.26	0.47
DMCyC ₅ (1.3, 1.2)	3.23	1.20
nC ₇	-	
MCyC ₆	12.45	4.65
toluene	0.44	0.16
2MC ₇	0.40	0.14
3MC ₇	0.79	0.29
DMCyC ₆ (1.2)	2.47	0.92
nC ₈	1.55	0.58
M/P-xylene	4.01	1.50
O-xylene	-	-

FIGURE 1

Gas chromatogram, C₂-C₈ hydrocarbons

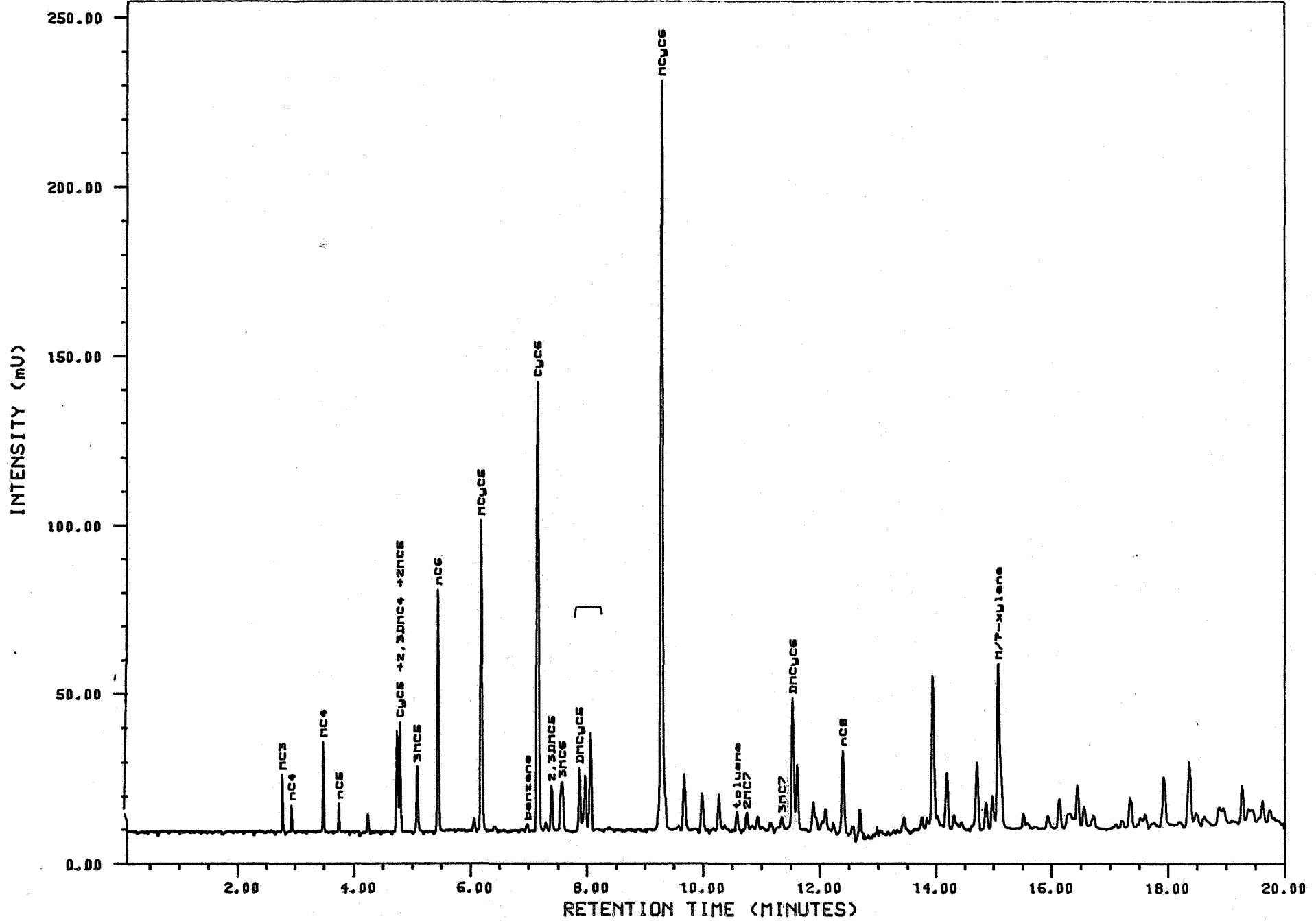


Table 5. Results of gas analysis 31/5-2

B-8971

Percentage composition

C ₁	80.7	
C ₂	8.1	
C ₃	1.5	
iC ₄	1.5	} iC ₄ /nC ₄ = 6.5
nC ₄	0.23	
iC ₅	0.22	} iC ₅ /nC ₅ = 4.8
nC ₅	0.046	
2.2DMeC ₄	0.024	
2.3DMeC ₄	0.049	
CyC ₅	0.039	
2-MeC ₅	0.077	
3-MeC ₅	0.041	
nC ₆	0.009	
MeCyC ₅	0.10	
2.4-DMeC ₅	0.01	
CyC ₆	0.10	
nC ₇	0.0002	
MeCyC ₆	0.069	

Table 6. Data from gas analysis.

CV# STOP RUN

[HP] 5899A MANUAL INJECTION @ 11:31 NOV 22, 1984
AREA %

RT	AREA	TYPE	WIDTH	HEIGHT	BASELINE	AREA %
0.00				BASELINE @ START RUN = 69.85		
0.00				THRESHOLD @ START RUN = 3		
0.00				PEAK WIDTH @ START RUN = 0.04		
3.14	123777.00	SV	0.03	65140.60	69.93	70.600
3.20	24908.00	VB	0.03	14330.80	70.23	14.207
3.74	7062.13	SB	0.03	4493.49	70.23	4.316
4.54	9971.45	SV	0.03	5279.93	70.02	5.448
5.19	1901.21	SB	0.03	912.77	70.40	0.656
5.48	151.70	SB	0.031	77.77	70.47	0.087
7.06	1922.87	SV	0.03	909.20	70.13	1.097
7.82	397.66	SB	0.031	201.75	70.36	0.227
8.36	216.95	SB	0.033	101.88	70.28	0.124
9.69	633.15	SV	0.033	207.81	70.23	0.248
9.76	349.36	VV	0.035	155.35	70.37	0.199
9.90	606.91	VB	0.03	324.56	70.63	0.302
10.31	369.90	SB	0.033	173.41	70.39	0.211
10.81	81.62	SB	0.033	39.11	70.21	0.047
11.51	40.29	SV	0.031	20.67	70.18	0.023
11.89	895.39	VV	0.035	398.77	70.22	0.511
11.68	39.60	VV	-----	36.45	70.25	0.051
11.83	61.91	VB	0.043	22.60	70.31	0.035
12.41	20.46	SV	0.034	9.34	70.23	0.012
12.53	969.15	VV	0.038	481.20	70.26	0.553
12.72	15.70	VV	-----	7.50	70.30	0.009
12.79	84.66	VV	0.035	38.11	70.31	0.048
12.89	75.13	VV	0.034	34.55	70.32	0.043
12.96	68.53	VB	0.04	29.77	70.34	0.039
13.17	83.46	SV	0.034	38.83	70.24	0.048
13.25	91.91	VV	-----	39.97	70.31	0.052
13.33	140.22	VB	0.04	53.80	70.33	0.080
14.23	610.41	SB	0.040	239.62	70.29	0.353
14.59	34.41	VB	0.037	14.65	70.37	0.020
14.71	22.84	SB	0.036	9.84	70.17	0.013
14.90	17.43	SV	0.036	7.58	70.13	0.010
15.67	34.24	VV	-----	14.40	70.17	0.020
15.72	17.63	VB	-----	7.46	70.17	0.010
16.19	15.92	SV	0.037	6.65	70.19	0.009
17.02	12.44	VB	0.039	4.96	70.33	0.007

TOTAL AREA = 175322.00
MULTIPLIER = 1

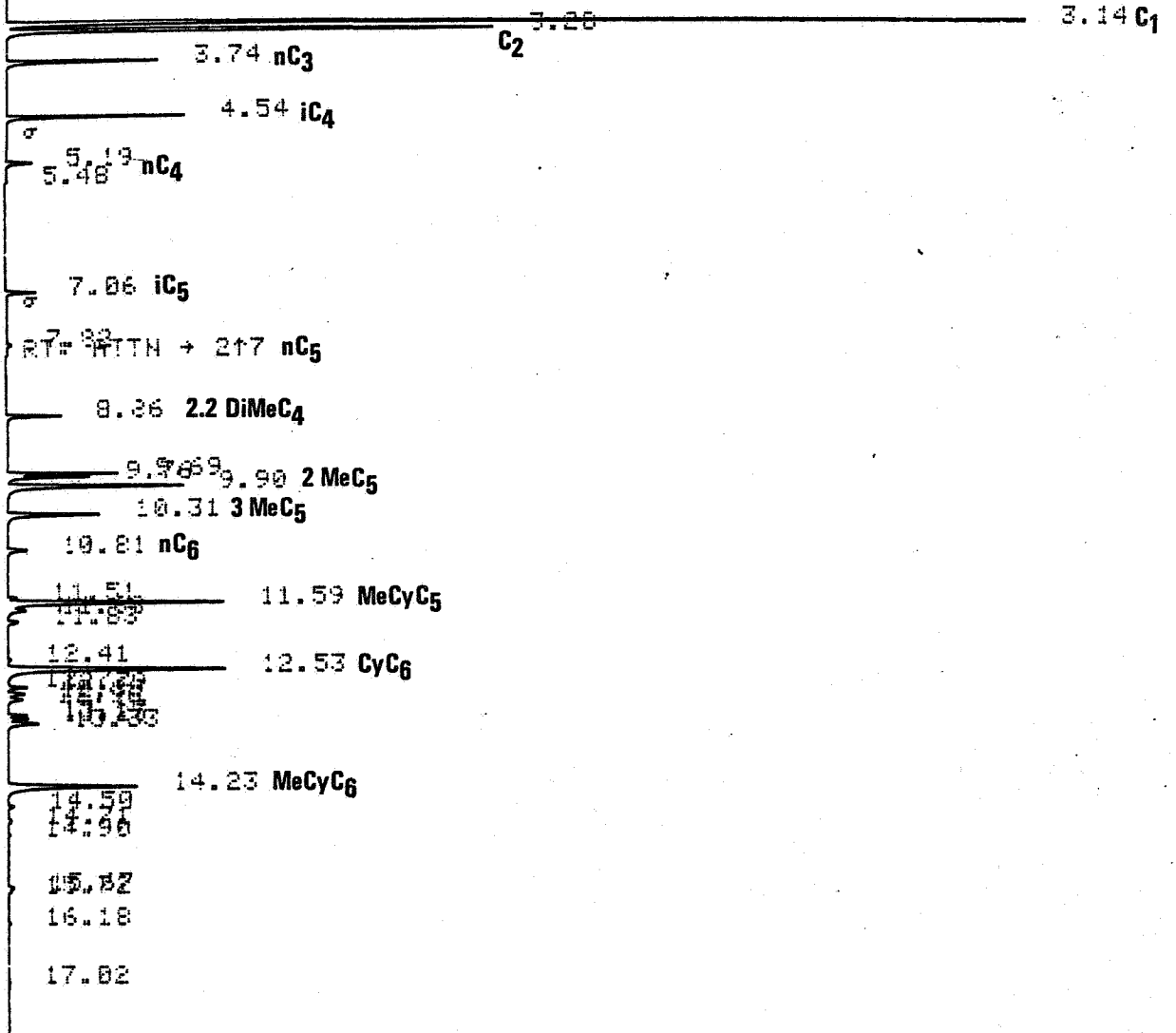
FIGURE 2

Gas chromatogram of gas sample

RT: VALVE 2 → ON

OVEN TEMP : FINAL TIME → 30.00 MIN

OV: START PRGM RATE 1



OV: START FINAL TIME 1

Gas chromatogram (C₁ - C₁₀)
31/5 - 2 gas sample B 8971

FIGURE 3

Saturated fraction gas chromatograms

- a - nC₁₇
- b - pristane
- c - phytane
- nC_x - n-alkane of that
carbon number

reated at 12:37 on 31/Oct/84

DATA PLOT-CHANNEL 3
Data Scale Plot

Box 1 of 1

Analysis : 750B8116S Sample f: 1 Injection f: 1
Sample Name : B8116, SAT, 31/5-2, GH Maximum signal {%} : 5.179

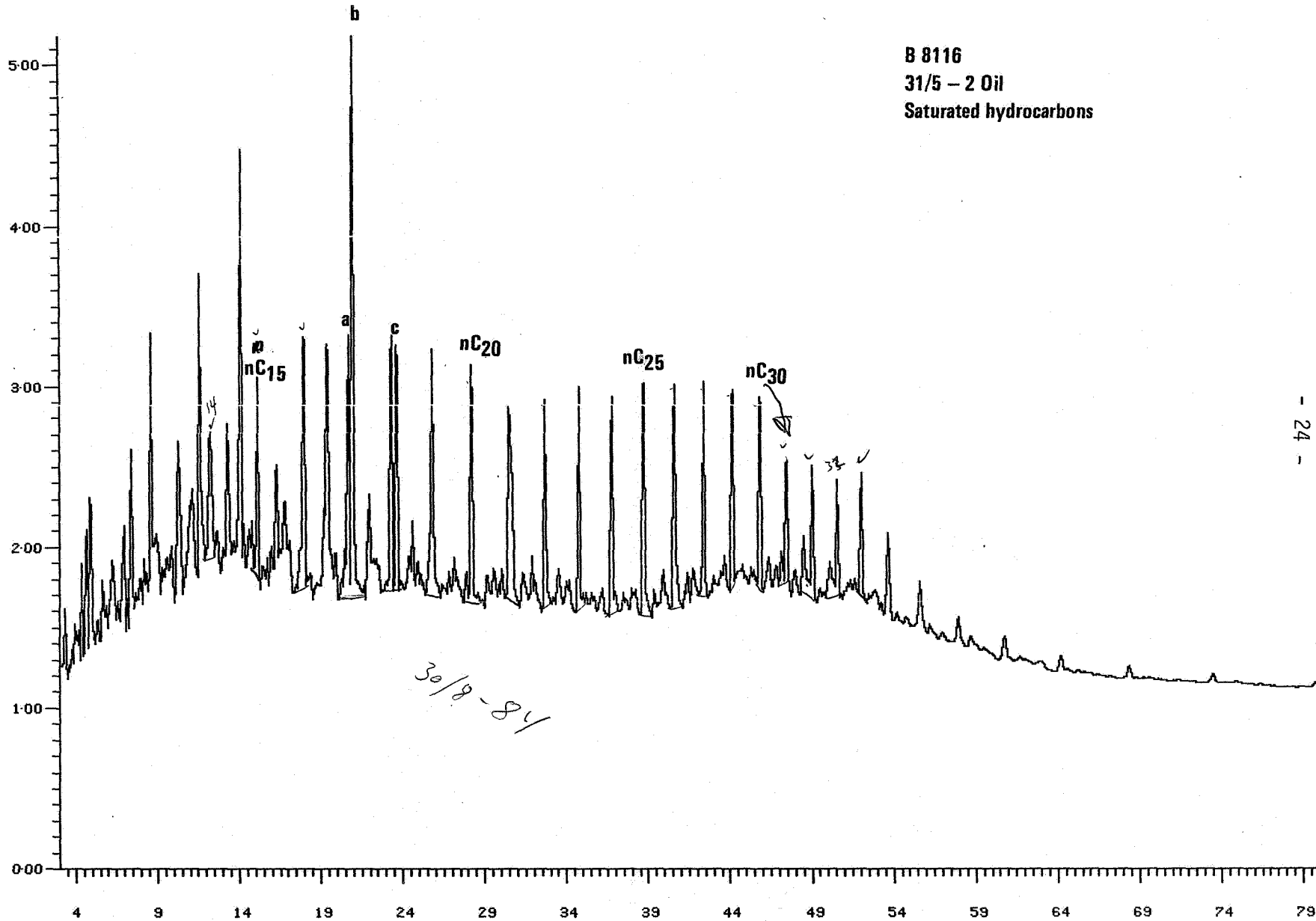


FIGURE 4

Branched and cyclic alkanes gas chromatogram

Pr - pristane
Ph - phytane
iso 16 Isoprenoid alkanes with 16 and 18
iso 18 carbon atoms respectively

reated at 17:56 on 31/Oct/84

DATA PLOT-CHANNEL 3

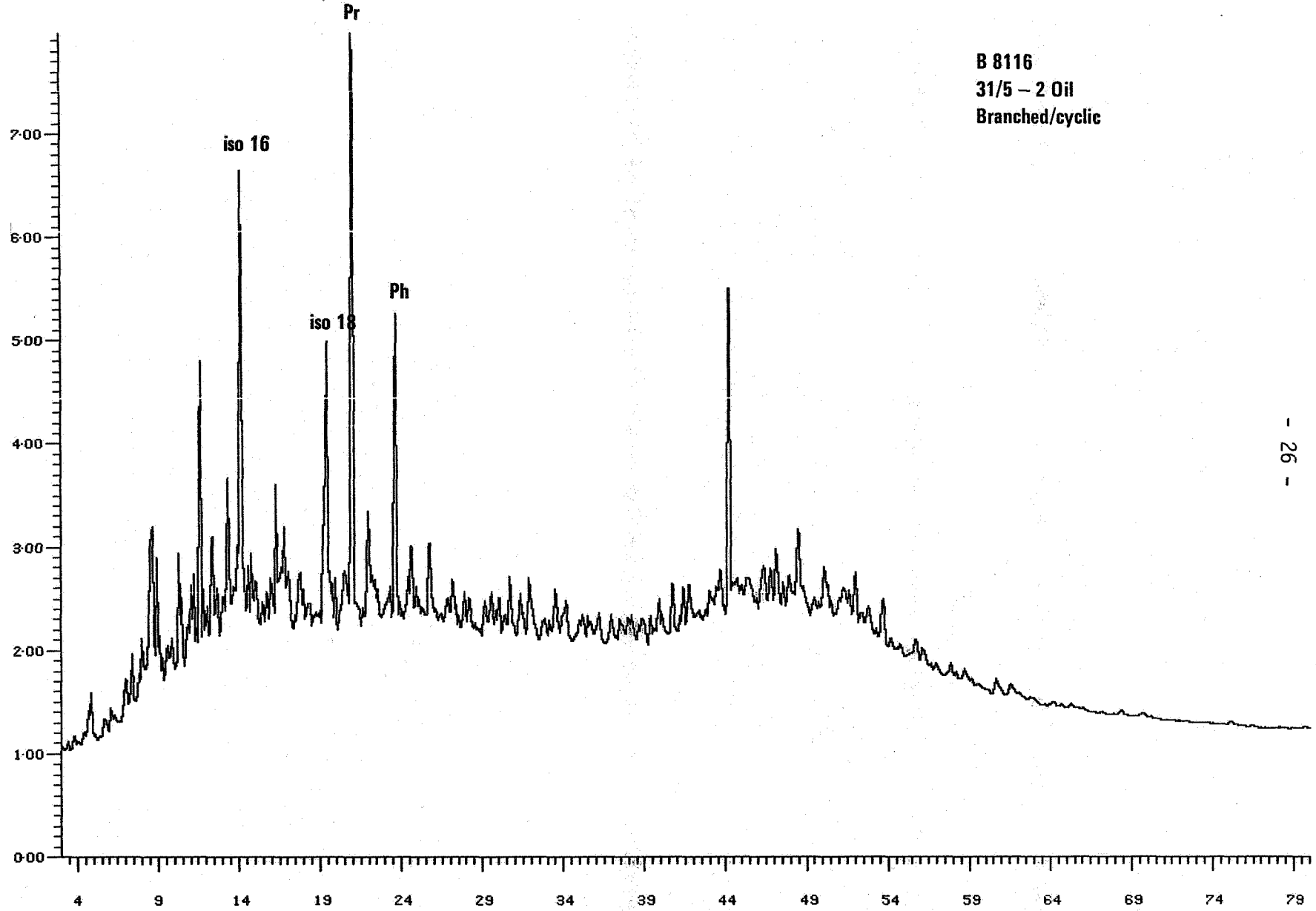
Data Scale Plot

Box 1 of 1

Analysis: 750B8116B Sample f: 1 Injection f: 1

Sample Name: BB116, B/C, 31/5-2, GH

Maximum signal <X> : 7.972



B 8116
31/5 - 2 Oil
Branched/cyclic

FIGURE 5

Aromatic fraction gas chromatogram

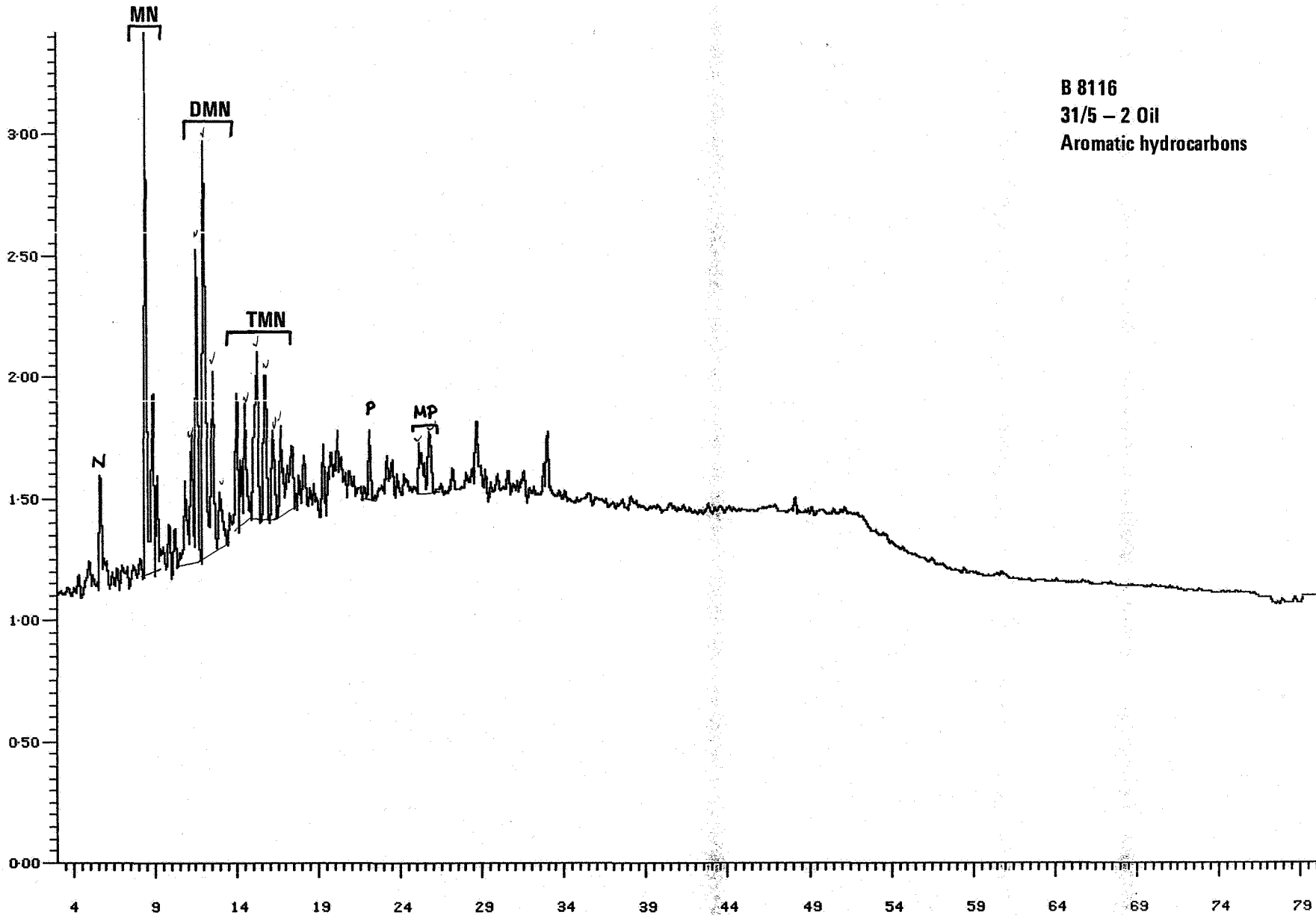
- N - naphthalene
- MN - methyl naphthalenes
- DMN - dimethyl naphthalenes
- TMN - trimethyl naphthalenes

reated at 12:41 on 31/Oct/84

DATA PLOT-CHANNEL 2

Data Scale Plot Box 1 of 1

Analysis : 750B8116A Sample #: 1 Injection #: 1
Sample Name : B8116, ARD, 31/5-2, GH Maximum signal (%): 3.419



B 8116
31/5 - 2 Oil
Aromatic hydrocarbons

Table 7. Results of $\delta^{13}\text{C}$ analyses on saturated and aromatic fractions of 31/5-2 oil (B-8116).

Sat. ($^{13}\delta\text{C}$ ‰)	Arom. ($^{13}\delta\text{C}$ ‰)
-28.8	-31.1



Table 8.

Molecular ratios calculated from terpane
and sterane mass chromatograms.

Maturity ratios.

IKU No.	DEPTH (m)	1) $\alpha\beta/\alpha\beta + \beta\alpha$	2) %22S	3) % $\beta\beta$	4) %20S
B8116	0	0.89	60.0	73.9	54.5

1) E/E+F in m/z 191.

2) % distribution between first and second
elution isomers of doublet J (m/z 191)

3) $2(r+s)/(q+t+2(r+s))$ in m/z 217.

4) $q/q+t$ in m/z 217.



Table 9.

Molecular ratios calculated from terpane
and sterane mass chromatograms.
Source characteristic and maturity ratios.

IKU No.	DEPTH (m)	1) Q/E	2) Tm/Ts	3) X/E	4) a/a+j	5) Z/E
B8116	0	0.09	0.84	0.11	0.70	0.24

- 1) Relative abundance of tricyclic terpanes(Q/E in m/z 191).
- 2) B/A in m/z 191.
- 3) Relative abundance of unknown(X/E in m/z 191).
- 4) Relative abundance of C27 rearranged steranes(a/a+j).
- 5) Relative abundance of bisnorhopane(Z/E in m/z 191).

Figure 6.

Mass chromatograms representing terpanes (m/z 191)

A	T _s , 18α(H)-trisorneohopane	C ₂₇ H ₄₆	(III)
B	T _m , 17α(H)-trisnorhopane	C ₂₇ H ₄₆	(I, R=H)
C	17α(H)-norhopane	C ₂₉ H ₅₀	(I, R=C ₂ H ₅)
D	17β(H)-normoretane	C ₂₉ H ₅₀	(II, R=C ₂ H ₅)
E	17α(H)-hopane	C ₃₀ H ₅₂	(I, R=C ₃ H ₇)
F	17β(H)-moretane	C ₃₀ H ₅₂	(II, R=C ₃ H ₇)
G	17α(H)-homohopane (22S)	C ₃₁ H ₅₄	(I, R=C ₄ H ₉)
H	17α(H)-homohopane (22R)	C ₃₁ H ₅₄	(I, R=C ₄ H ₉)
	+ unknown triterpane (gammacerane?)		
I	17β(H)-homomoretane	C ₃₁ H ₅₄	(II, R=C ₄ H ₉)
J	17α(H)-bishomohopane (22S,22R)	C ₃₂ H ₅₆	(I, R=C ₅ H ₁₁)
K	17α(H)-trishomohopane (22S,22R)	C ₃₃ H ₅₈	(I, R=C ₆ H ₁₃)
L	17α(H)-tetrakishomohopane (22S,22R)	C ₃₄ H ₆₀	(I, R=C ₇ H ₁₅)
M	17α(H)-pentakishomohopane (22S,22R)	C ₃₅ H ₆₂	(I, R=C ₈ H ₁₇)
Z	bisnorhopane	C ₂₈ H ₄₈	
X	unknown triterpane	C ₃₀ H ₅₂	
P	tricyclic terpene	C ₂₃ H ₄₂	(IV, R=C ₄ H ₉)
Q	tricyclic terpene	C ₂₄ H ₄₄	(IV, R=C ₅ H ₁₁)
R	tricyclic terpene (17R,17S)	C ₂₅ H ₄₆	(IV, R=C ₆ H ₁₃)
S	tetracyclic terpene	C ₂₄ H ₄₂	(V)
T	tricyclic terpene (17R,17S)	C ₂₆ H ₄₈	(IV, R=C ₇ H ₁₅)

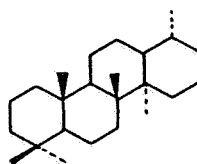
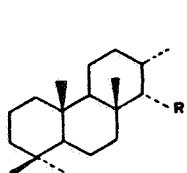
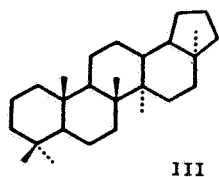
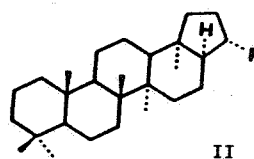
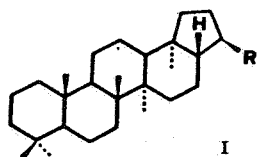
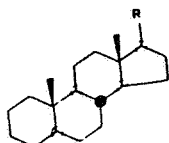
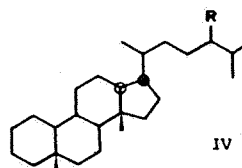
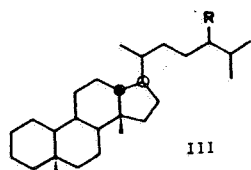
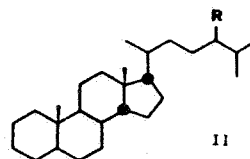
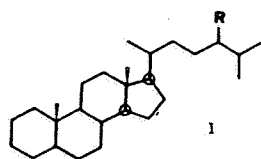


Figure 7.

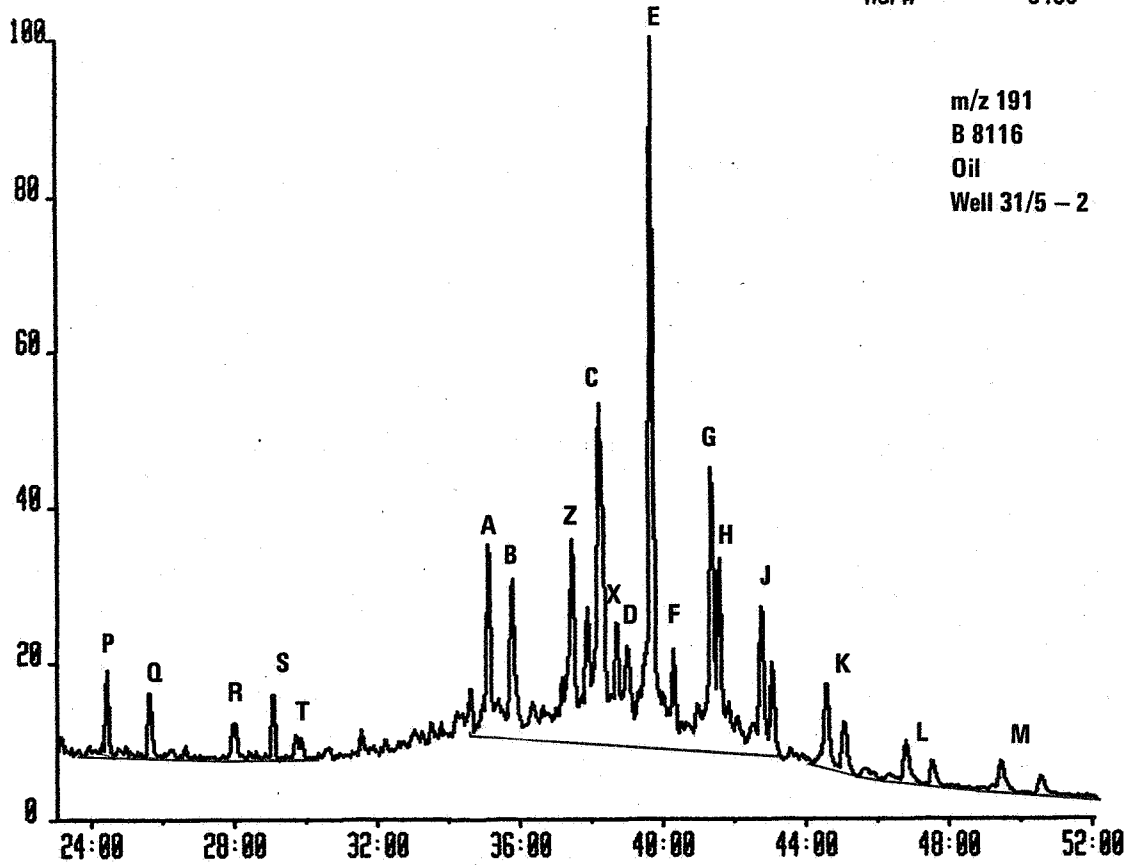
Mass chromatograms representing steranes (m/z 217 and 218)

a	13 β (H),17 α (H)-diasterane (20S)	C ₂₇ H ₄₈	(III,R=H)
b	13 β (H),17 α (H)-diasterane (20R)	C ₂₇ H ₄₈	(III,R=H)
c	13 α (H),17 β (H)-diasterane (20S)	C ₂₇ H ₄₈	(IV,R=H)
d	13 α (H),17 β (H)-diasterane (20R)	C ₂₇ H ₄₈	(IV,R=H)
e	13 β (H),17 α (H)-diasterane (20S)	C ₂₈ H ₅₀	(III,R=CH ₃)
f	13 β (H),17 α (H)-diasterane (20R)	C ₂₈ H ₅₀	(III,R=CH ₃)
g	13 α (H),17 β (H)-diasterane (20S)	C ₂₈ H ₅₀	(IV,R=CH ₃)
	+ 14 α (H),17 α (H)-sterane (20S)	C ₂₇ H ₄₈	(I,R=H)
h	13 β (H),17 α (H)-diasterane (20S)	C ₂₉ H ₅₂	(III,R=C ₂ H ₅)
	+ 14 β (H),17 β (H)-sterane (20R)	C ₂₇ H ₄₈	(II,R=H)
i	14 β (H),17 β (H)-sterane (20S)	C ₂₇ H ₄₈	(II,R=H)
	+ 13 α (H),17 β (H)-diasterane (20R)	C ₂₈ H ₅₀	(IV,R=CH ₃)
j	14 α (H),17 α (H)-sterane (20R)	C ₂₇ H ₄₈	(I,R=H)
k	13 β (H),17 α (H)-diasterane (20R)	C ₂₉ H ₅₂	(III,R=C ₂ H ₅)
l	13 α (H),17 β (H)-diasterane (20S)	C ₂₉ H ₅₂	(III,R=C ₂ H ₅)
m	14 α (H),17 α (H)-sterane (20S)	C ₂₈ H ₅₀	(I,R=CH ₃)
n	13 α (H),17 β (H)-diasterane (20R)	C ₂₉ H ₅₂	(III,R=C ₂ H ₅)
	+ 14 β (H),17 β (H)-sterane (20R)	C ₂₈ H ₅₀	(II,R=CH ₃)
o	14 β (H),17 β (H)-sterane (20S)	C ₂₈ H ₅₀	(II,R=CH ₃)
p	14 α (H),17 α (H)-sterane (20R)	C ₂₈ H ₅₀	(I,R=CH ₃)
q	14 α (H),17 α (H)-sterane (20S)	C ₂₉ H ₅₂	(I,R=C ₂ H ₅)
r	14 β (H),17 β (H)-sterane (20R)	C ₂₉ H ₅₂	(II,R=C ₂ H ₅)
	+ unknown sterane		
s	14 β (H),17 β (H)-sterane (20S)	C ₂₉ H ₅₂	(II,R=C ₂ H ₅)
t	14 β (H),17 β (H)-sterane (20R)	C ₂₉ H ₅₂	(I,R=C ₂ H ₅)
u	5 α (H)-sterane	C ₂₁ H ₃₆	(V,R=C ₂ H ₅)
v	5 α (H)-sterane	C ₂₂ H ₃₈	(IV,R=C ₃ H ₇)



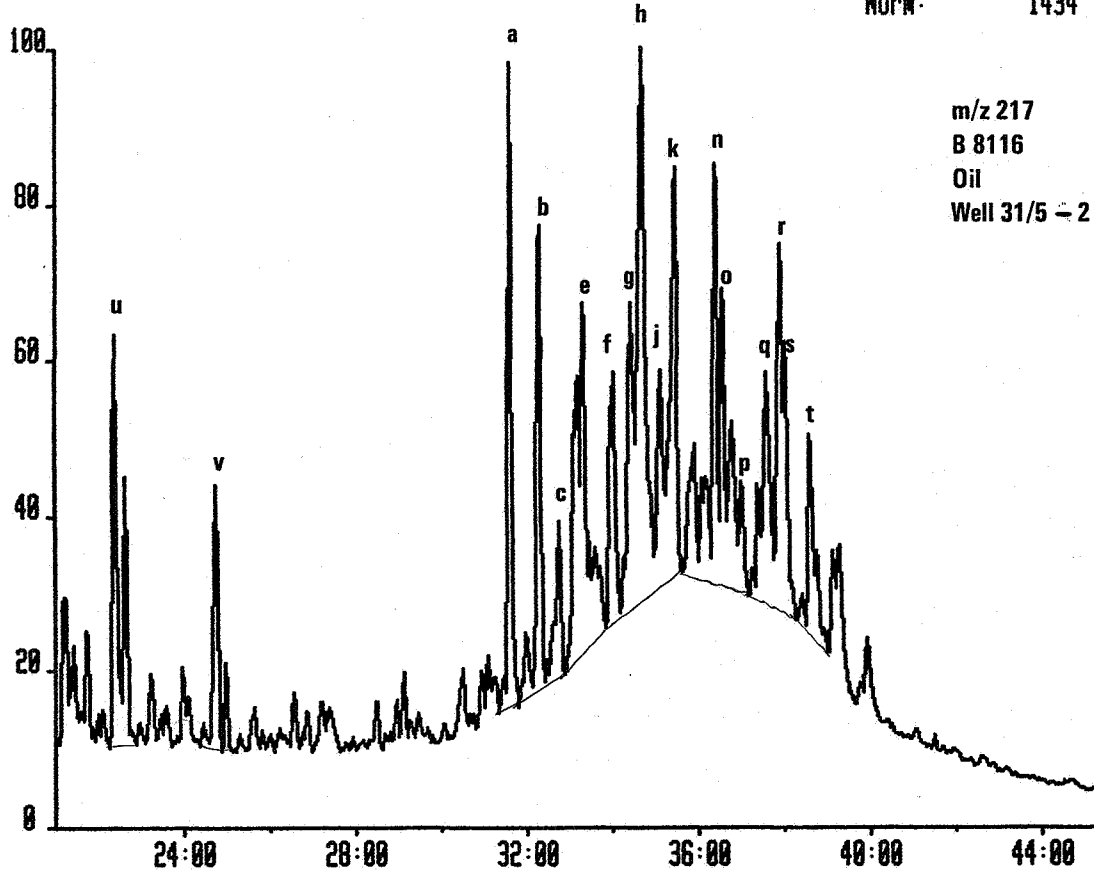
00116SAT 191.1000 61 11 S1

Norm: 5435



08116SAT 217.1000 G1 I1 S1

Norm: 1434



B0116SAT 210.1000 G1 I1 S1

Norm: 1164

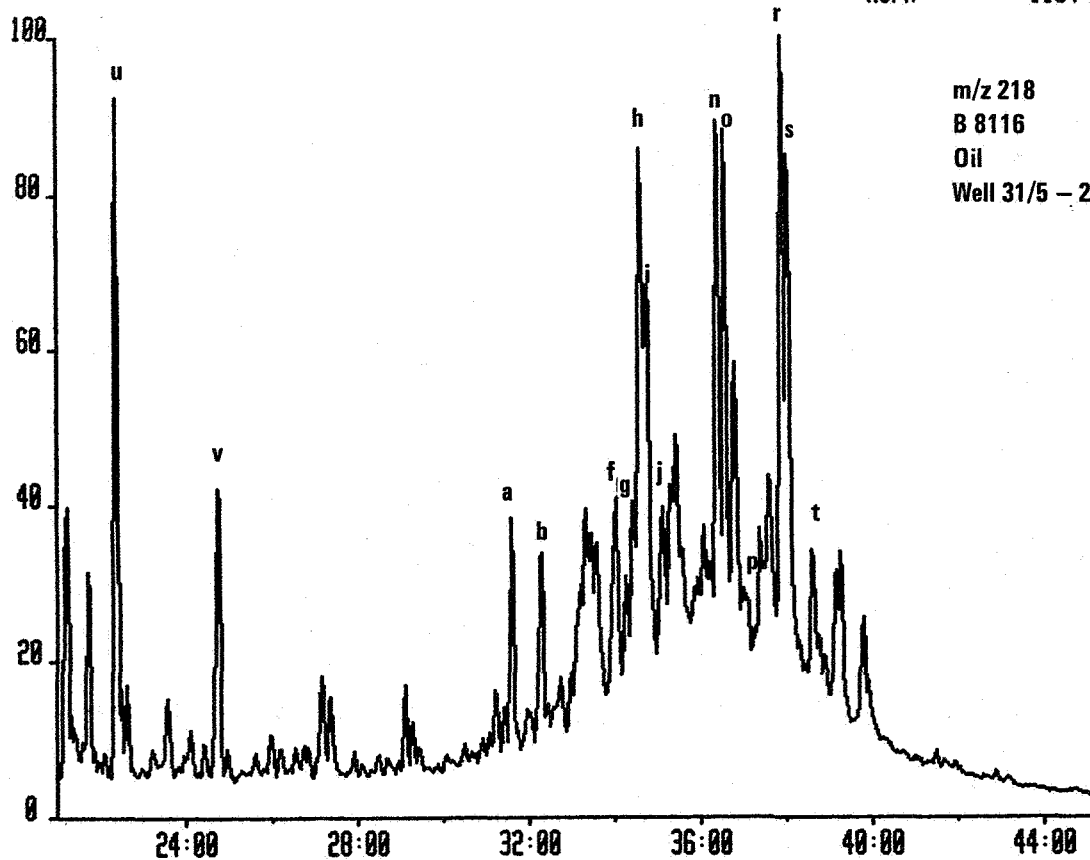
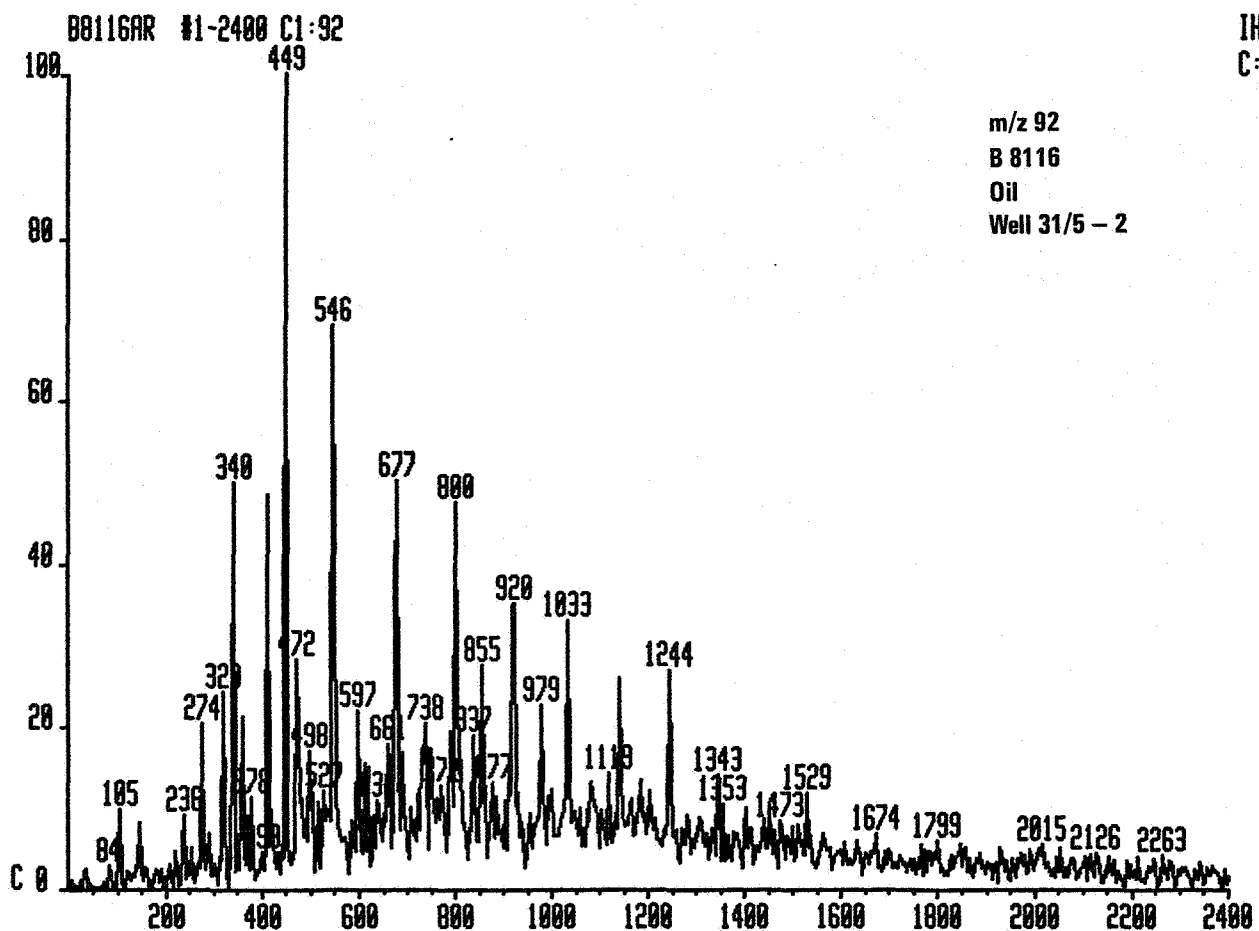
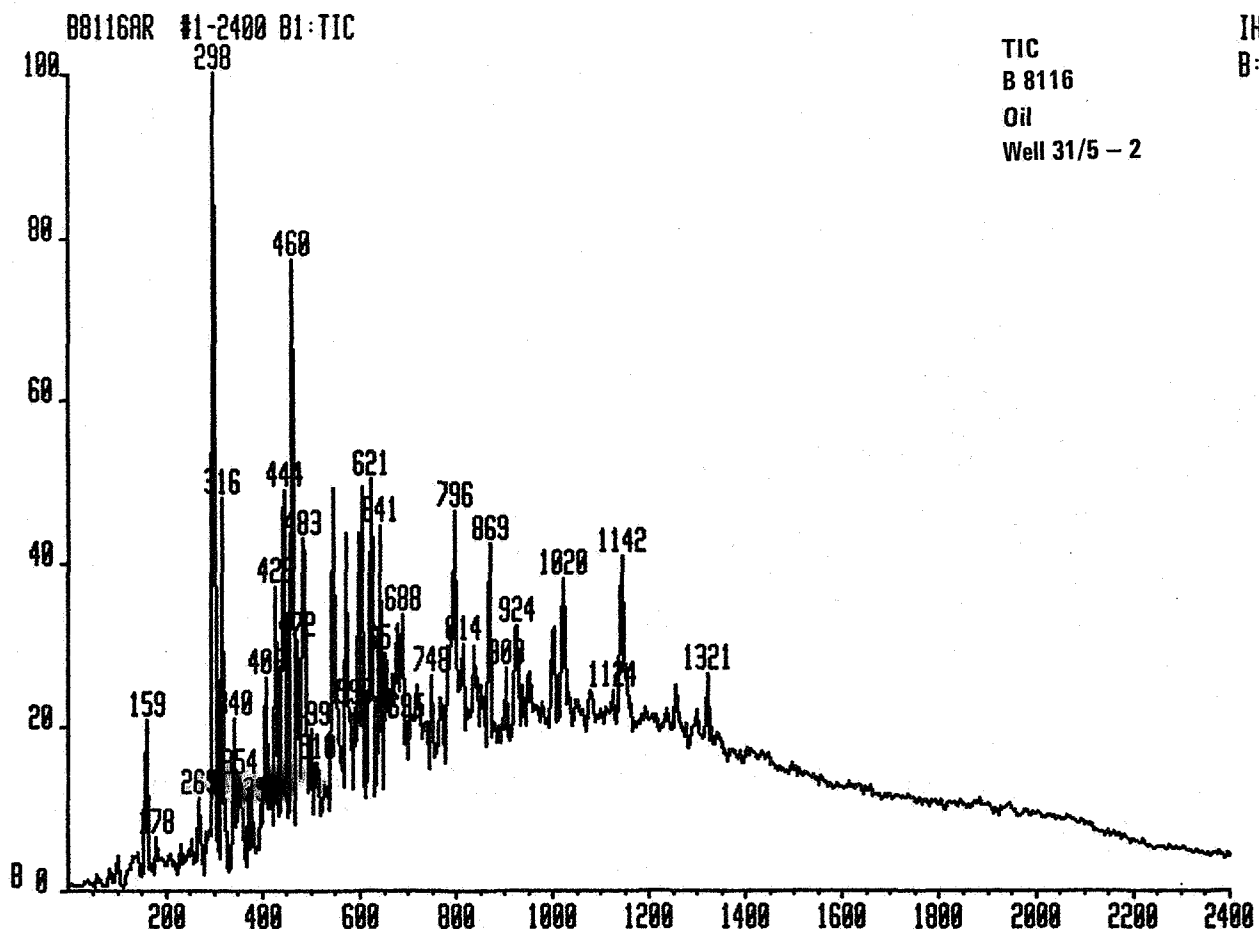


FIGURE 8

Mass chromatograms of aromatic hydrocarbons

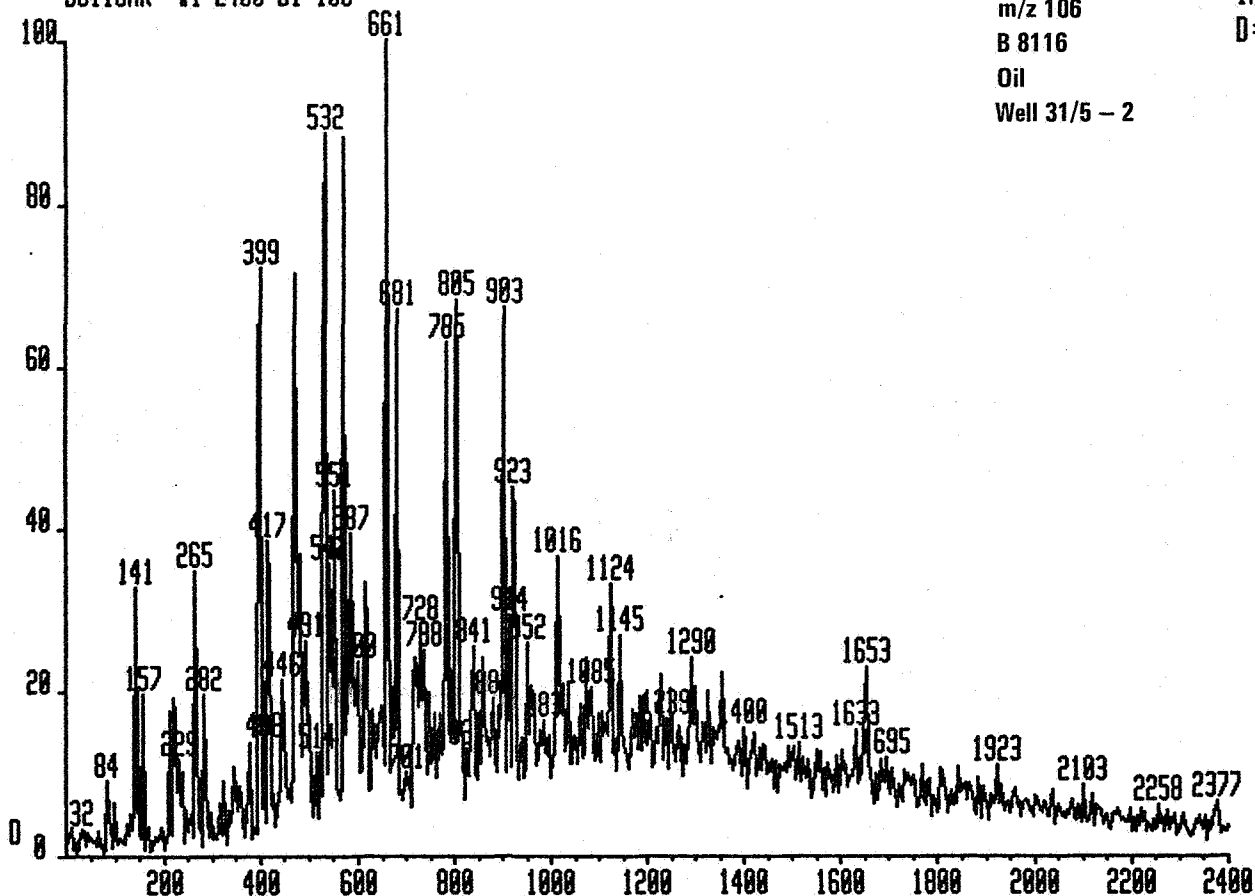
TIC	- total ion chromatograms
m/z 92,106	- monoaromatic hydrocarbons
m/z 142,156,170	- alkylated naphthalenes
m/z 178,192,206	- alkylated phenanthrenes
m/z 184,198,212	- alkylated dibenzothiophenes
m/z 231	- triaromatic steranes
m/z 253	- monoaromatic steranes
m/z 166,180	- fluorens
m/z 202	- pyrene, fluoranthen

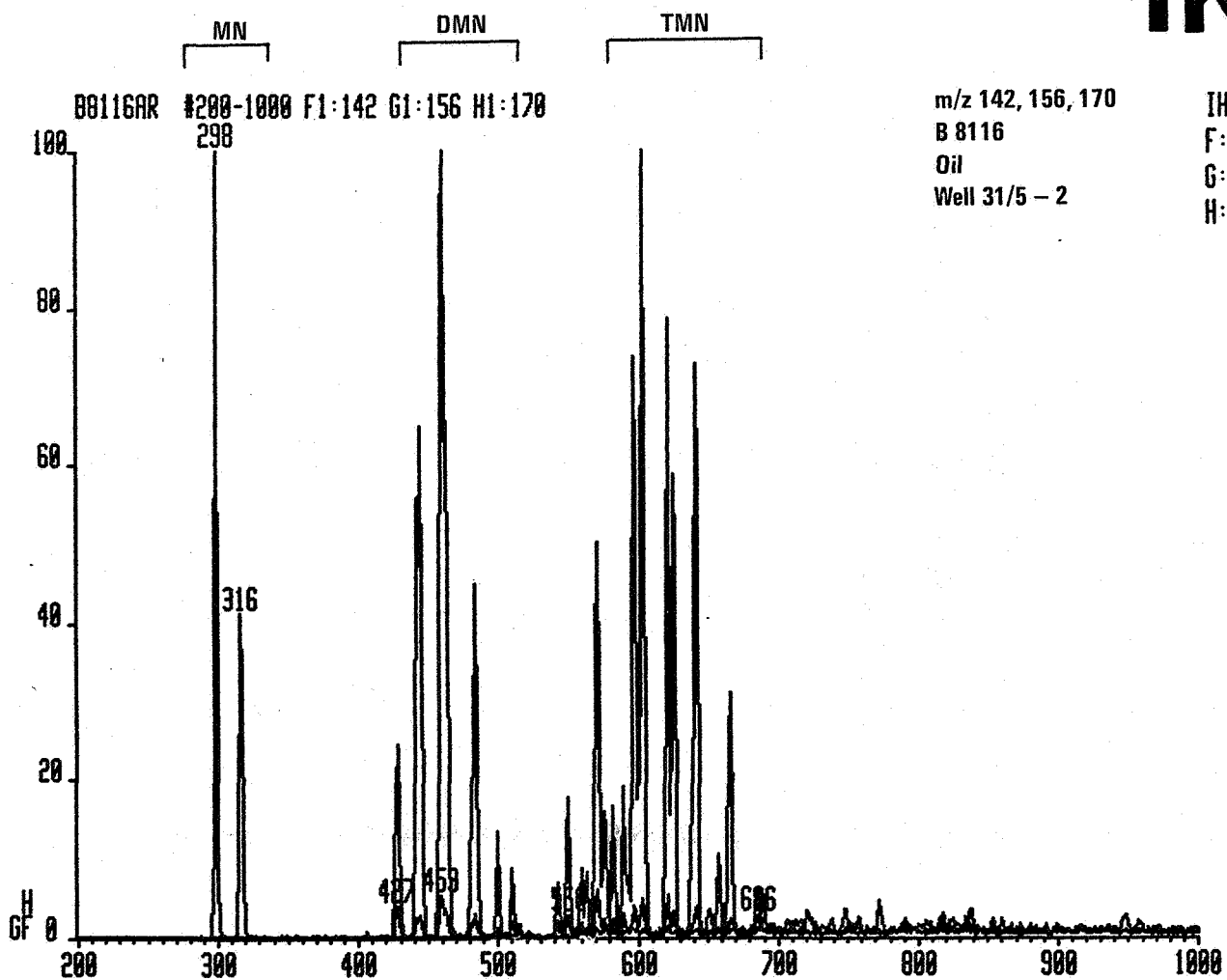


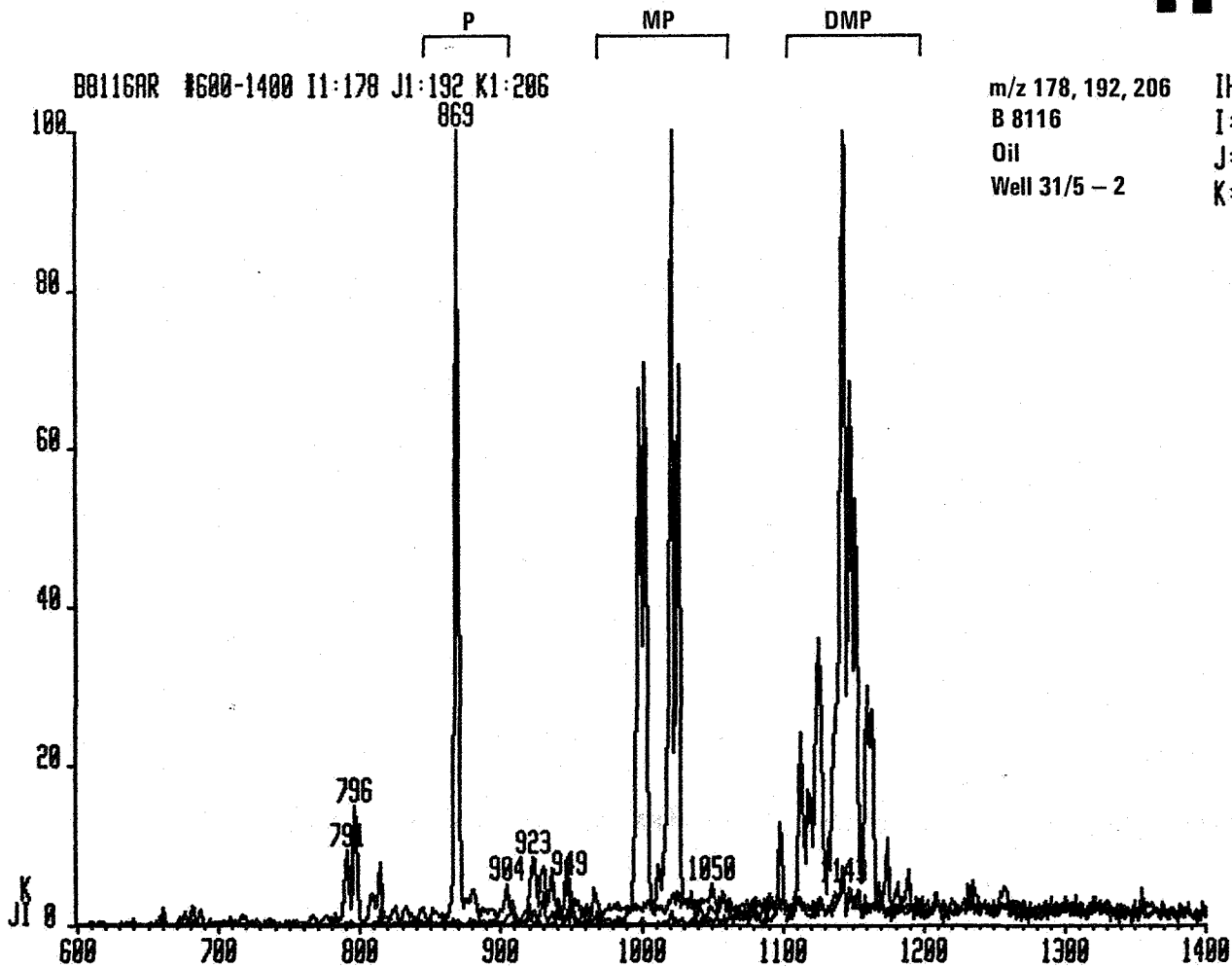
BB116AR #1-2400 01:106

m/z 106
B 8116
Oil
Well 31/5 - 2

IHP
D: 3395900

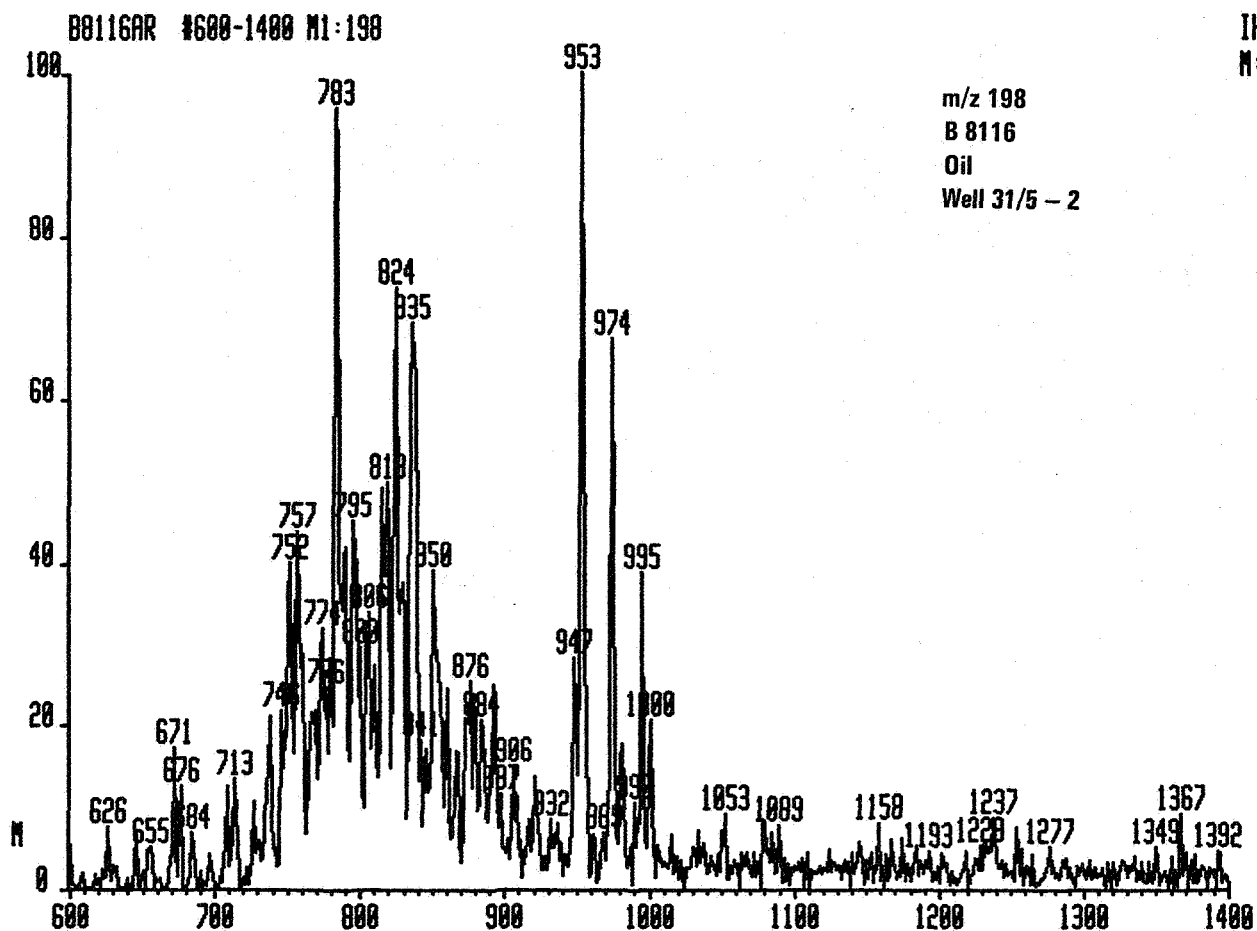
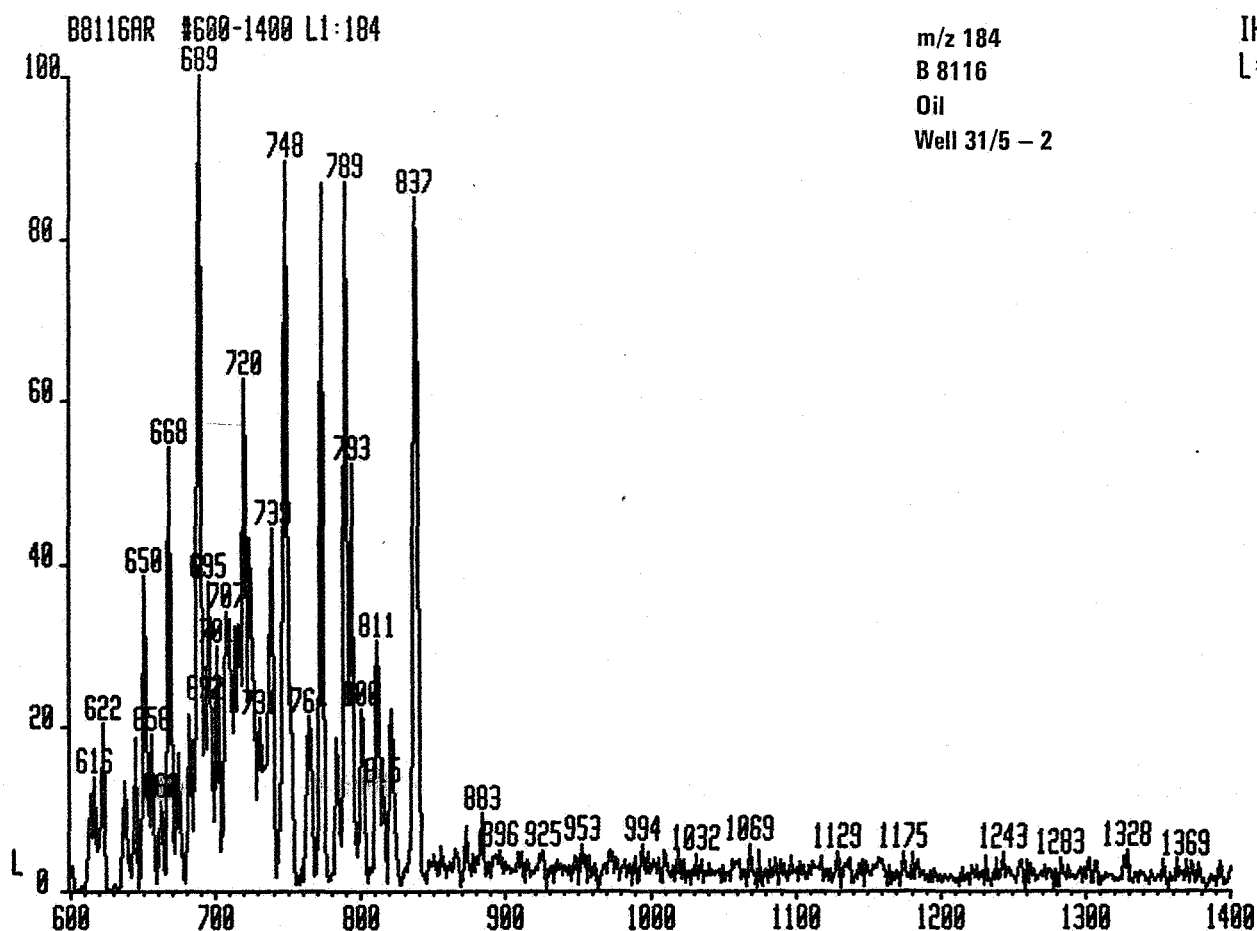






m/z 178, 192, 206
B 8116
Oil
Well 31/5 - 2

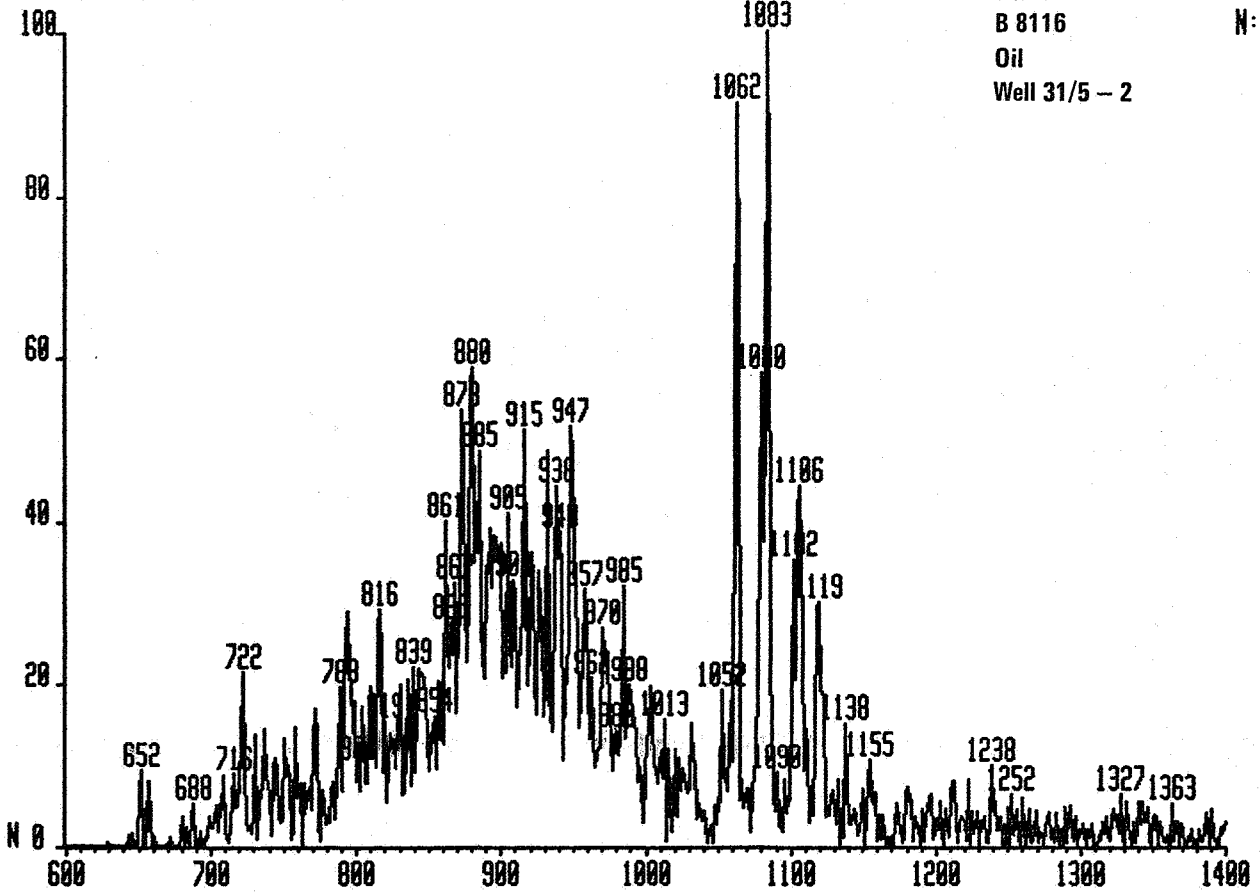
IHP
I: 1597500
J: 962000
K: 655340

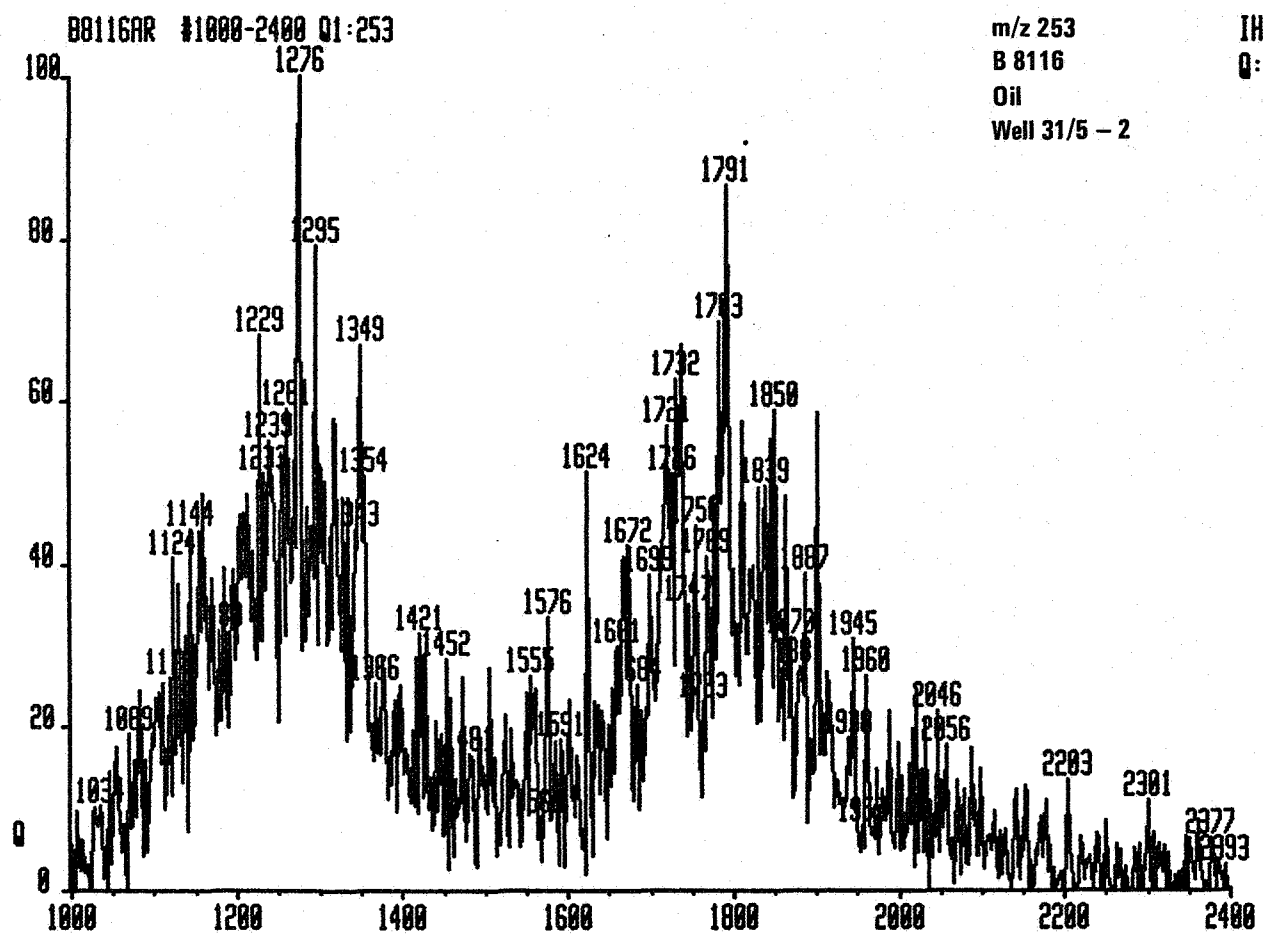
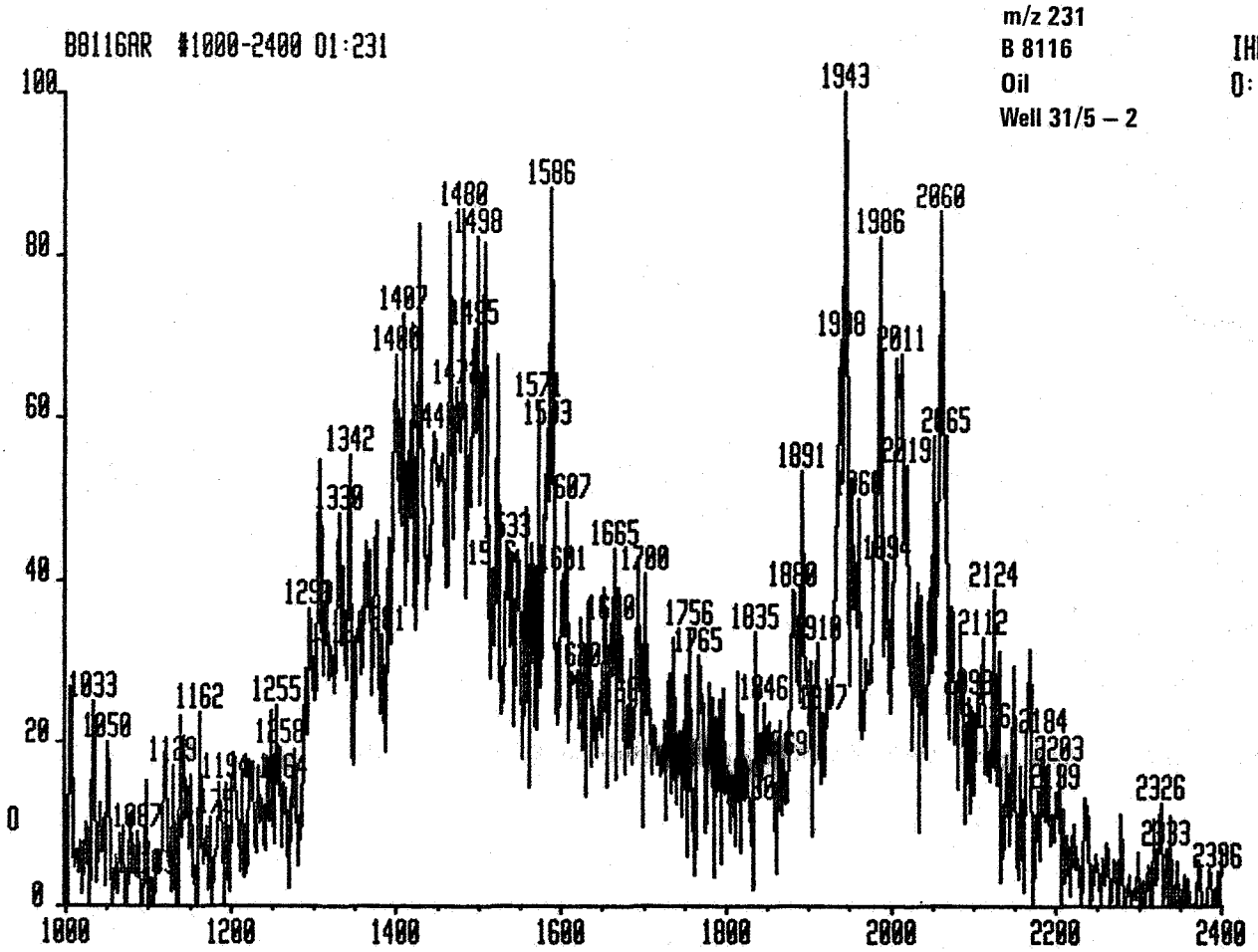


88116AR #600-1400 N1:212

m/z 212
B 8116
Oil
Well 31/5 - 2

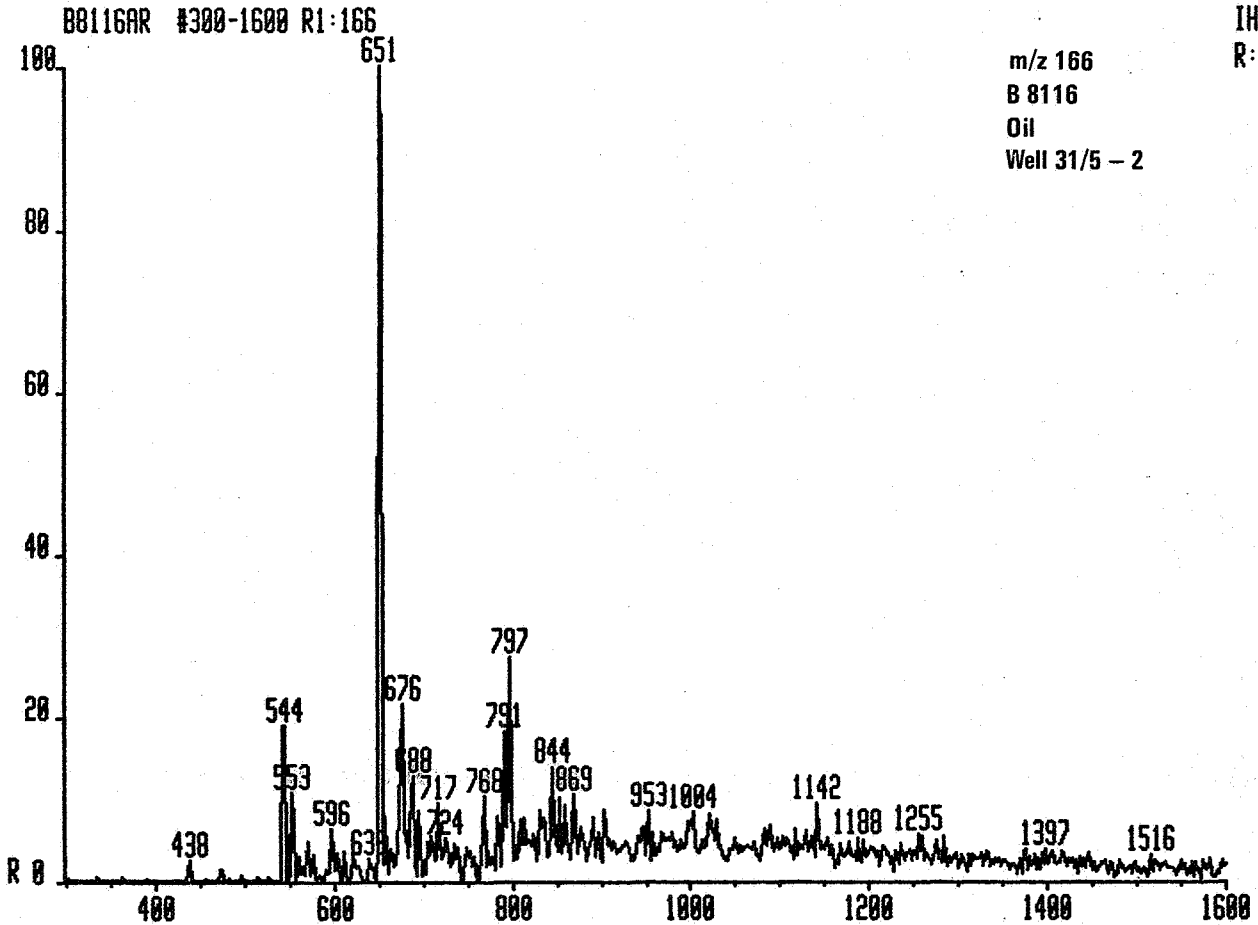
IHP
N: 149250





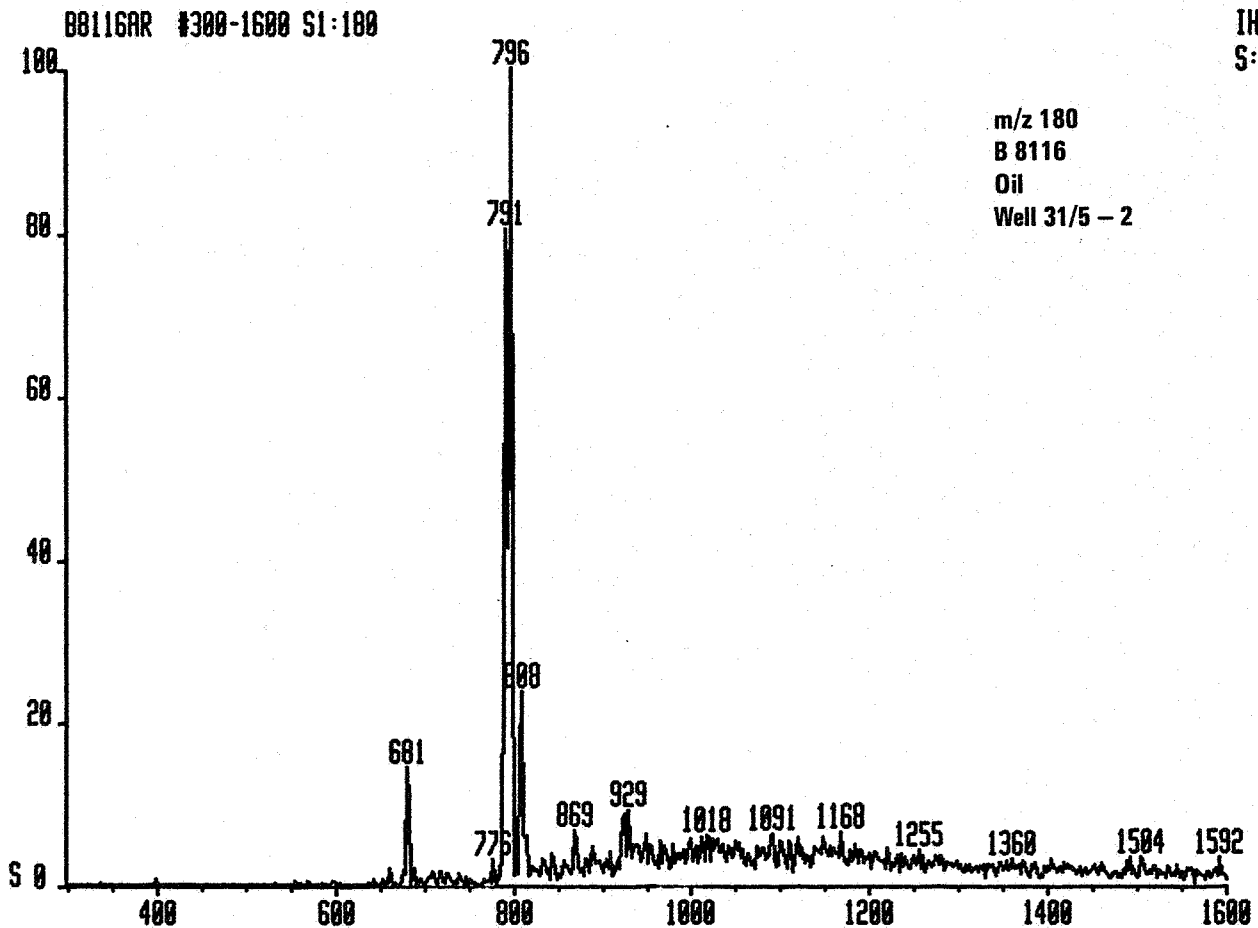
IHP
R: 5878300

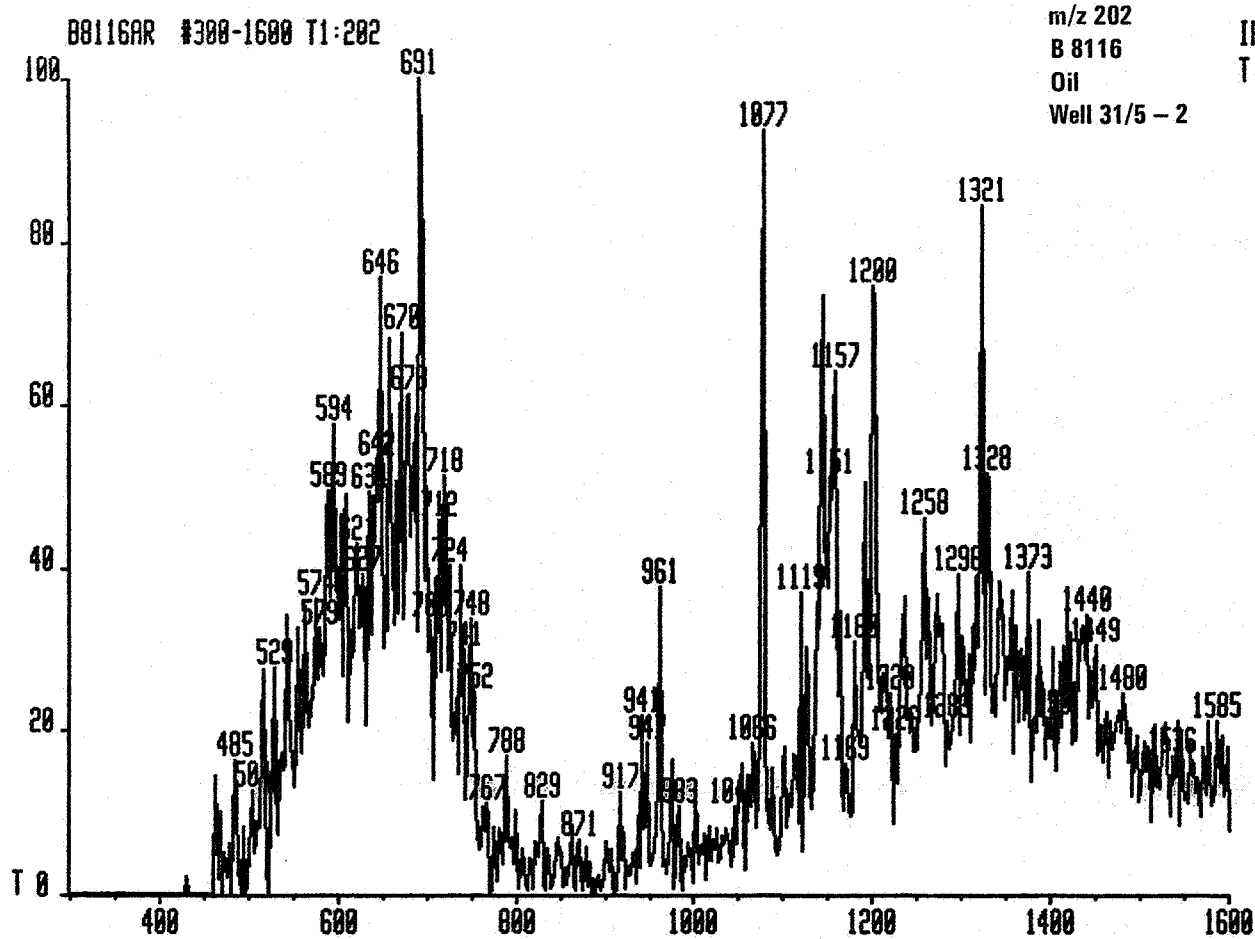
m/z 166
B 8116
Oil
Well 31/5 - 2



IHP
S: 6553400

m/z 180
B 8116
Oil
Well 31/5 - 2





**IKU**

Visual Kerogen Analysis

TABLE NO.: 9.
WELL NO.: 31/5-2

Sample	Depth (m)	Composition of residue	Particle size	Preservation palynomorphs	Thermal maturation index	Remarks
B-580	2052	C,W,WR!,Cut,S,P/Am,Cy Algal	F-M	good	1/1+,2-,2/2,2+	Abundant pyritic, coaly fragments and sapropelised woody fragments <u>Botryococcus</u> . Variable colouring.
B-583	2079	W,Cut,C,S,P/Am	F-M-L	good	1/1+, 2/2+	Abundant structured woody material and variably coloured spores.
B-587	2115	Cut,P,S,W,WR!/Am	F-M-L	good	1/1+, 2-/2, 2	Screening enriches particularly woody structured material <u>Chasmatosporites</u> .
B-591	2151	Cut,W,P,S,WR!/Am,Cy	F-M-L	good	1/1+	<u>Tyttodiscus</u> , <u>Tasmanites</u> , <u>Nannoceratopsis gracilis</u> .

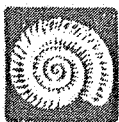
ABBREVIATIONS

Am Amorphous
He Herbaceous
Cut Cuticles

Cy Cysts, algae
P Pollen grains
S Spores

W Woody material
C Coal
R! Reworked

F Fine
M Medium
L Large

**IKU**

Visual Kerogen Analysis

TABLE NO.: 9.
WELL NO.: 31/5-2

Sample	Depth (m)	Composition of residue	Particle size	Preservation palynomorphs	Thermal maturation index	Remarks
B-595	2187	C,W,Cut,P,S/Am,Cy	F-M-L	good	1/1+	<u>Callialasporites</u> present in the ass. described above.
B-598	2214	C,Cut,P,S,W/Am		good	1+/2-, 2-	Pyritic residue, coaly fragments. Large very well preserved cuticles.

ABBREVIATIONS

Am Amorphous
He Herbaceous
Cut Cuticles

Cy Cysts, algae
P Pollen grains
S Spores

W Woody material
C Coal
R! Reworked

F Fine
M Medium
L Large

FIGURE 1

Saturated hydrocarbon gas chromatograms

- a = nC₁₇
- b = pristane
- c = phytane
- nC₁₅ etc. = n-alkane of that number
- X = contaminants
- Sq = squalane

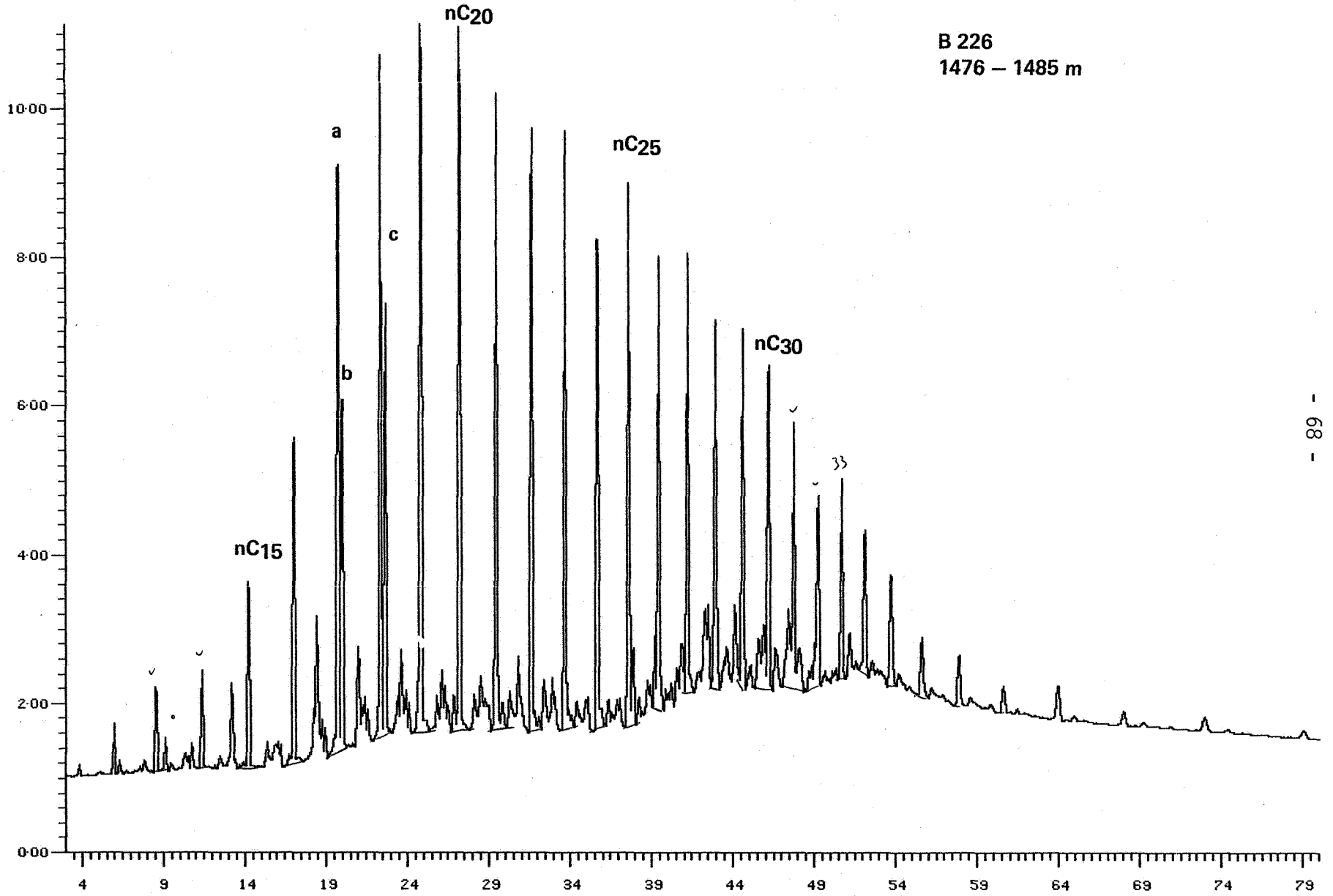
Created at 14:39 on 15/Feb/84

RAW DATA PLOT - CHANNEL 3

Data Scale Plot Box 1 of 1

Analysis : 198B226S1 Sample #: 1 Injection #: 1

Sample Name : B-226,SAT,31/5-1,AD Maximum signal (%): 11.132



B 226
1476 - 1485 m

Created at 14:43 on 15/Feb/84

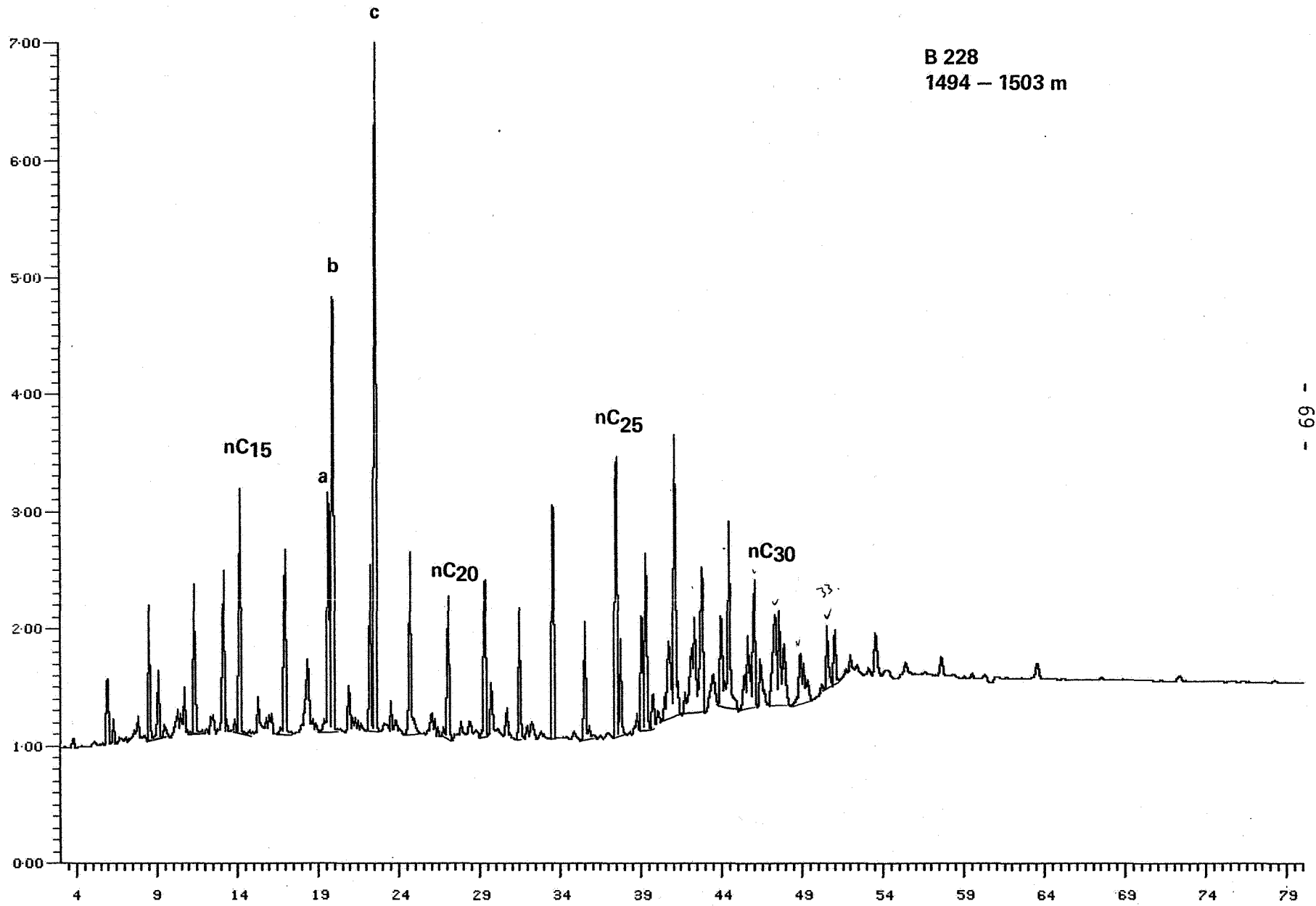
RAW DATA PLOT-CHANNEL 3

Data Scale Plot Box 1 of 1

Analysis : 198B22851 Sample #: 1 Injection #: 1

Sample Name : B-228, SAT, 31/5-1, AD

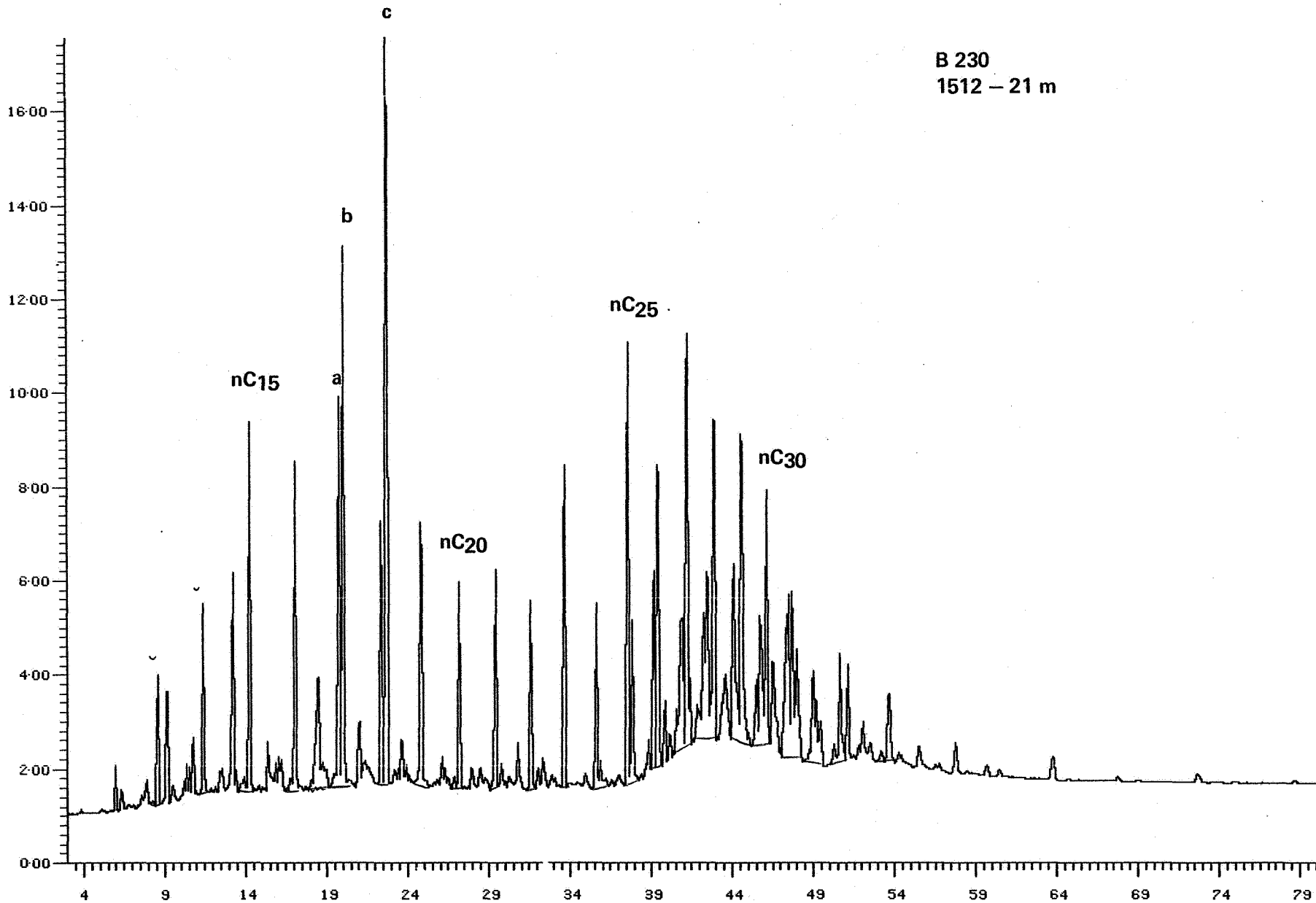
Maximum signal (%): 7.003



Created at 14:46 on 15/Feb/84

RAW DATA PLOT-CHANNEL 3
Data Scale Plot Box 1 of 1

Analysis : 198B230S1 Sample f: 1 Injection f: 1
Sample Name : B-230,SAT,31/5-1,AD Maximum signal [%] : 17.549



B 230
1512 - 21 m

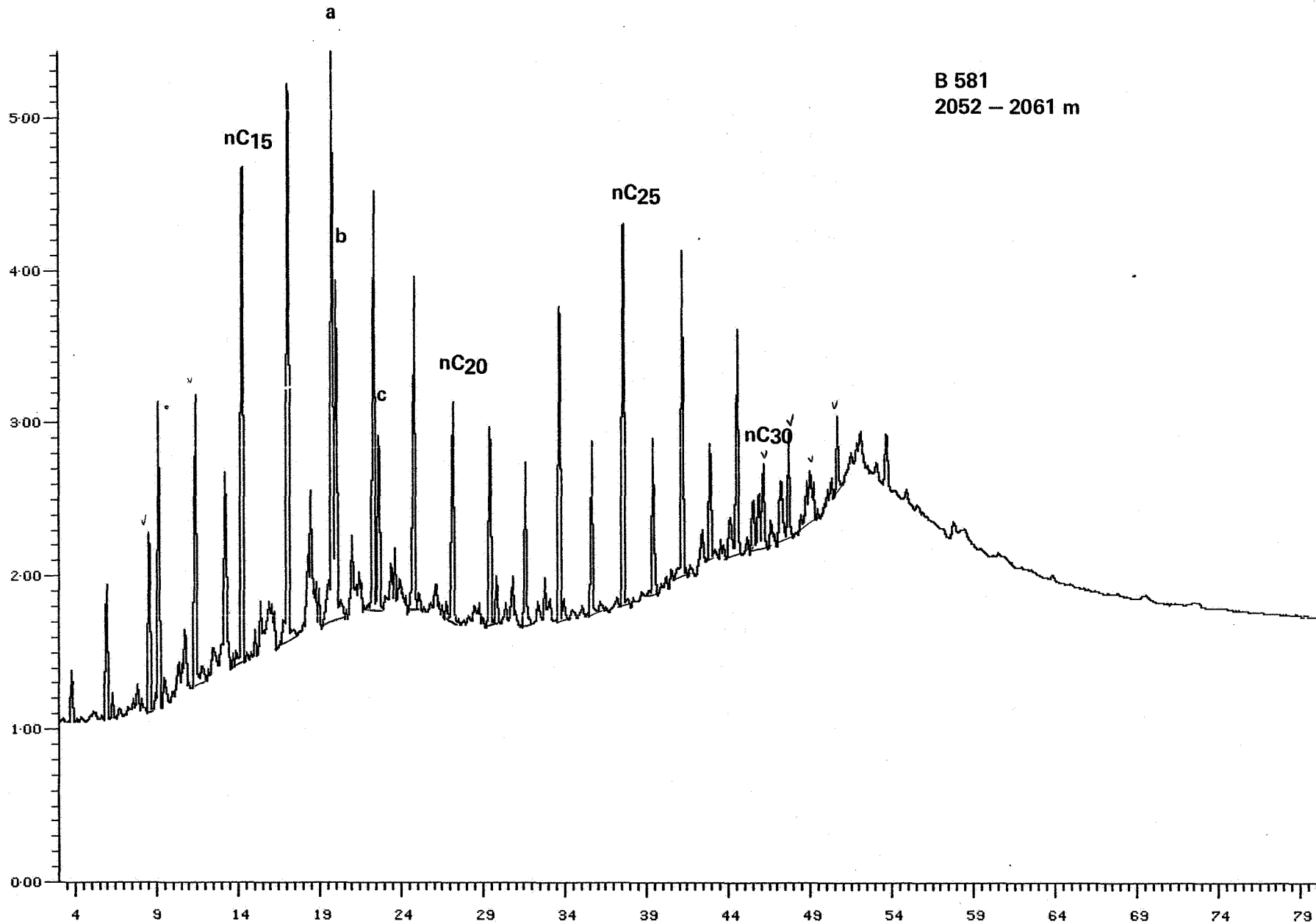
Created at 14:34 on 15/Feb/84

RAW DATA PLOT-CHANNEL 3
Data Scale Plot

Box 1 of 1

Analysis : 198B581S1 Sample f: 1 Injection f: 1
Sample Name : B-581,SAT,31/5-1,AD

Maximum signal <%> : 5.429



B 581
2052 - 2061 m

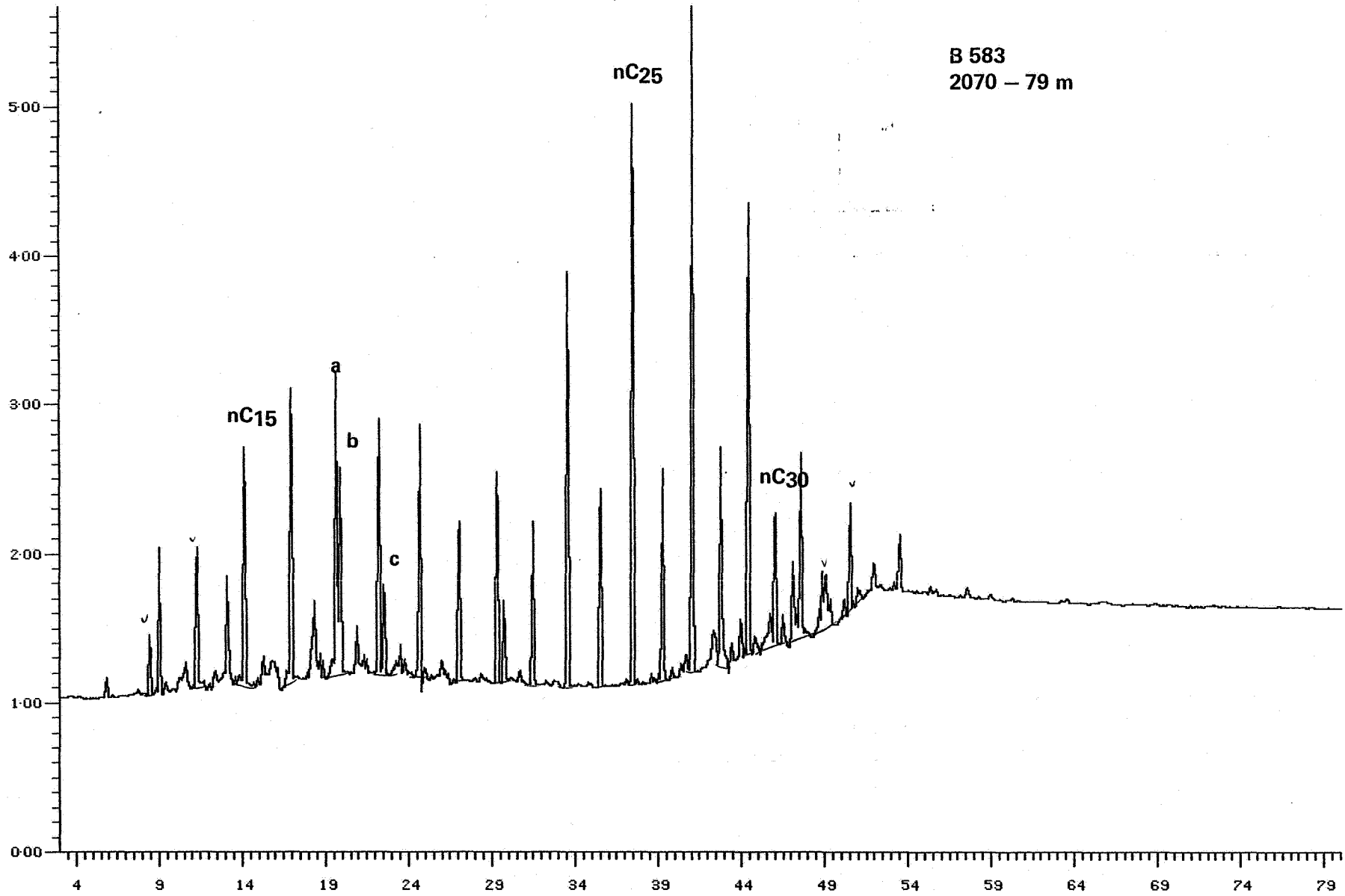
Created at 14:51 on 15/Feb/84

RAW DATA PLOT-CHANNEL 3

Data Scale Plot

Box 1 of 1

Analysis : 198B583S1 Sample f: 1 Injection f: 1
Sample Name : B-583, SAT, 31/5-1, AD Maximum signal (%): 5.669



Created at 14:36 on 15/Feb/84

RAW DATA PLOT-CHANNEL 3

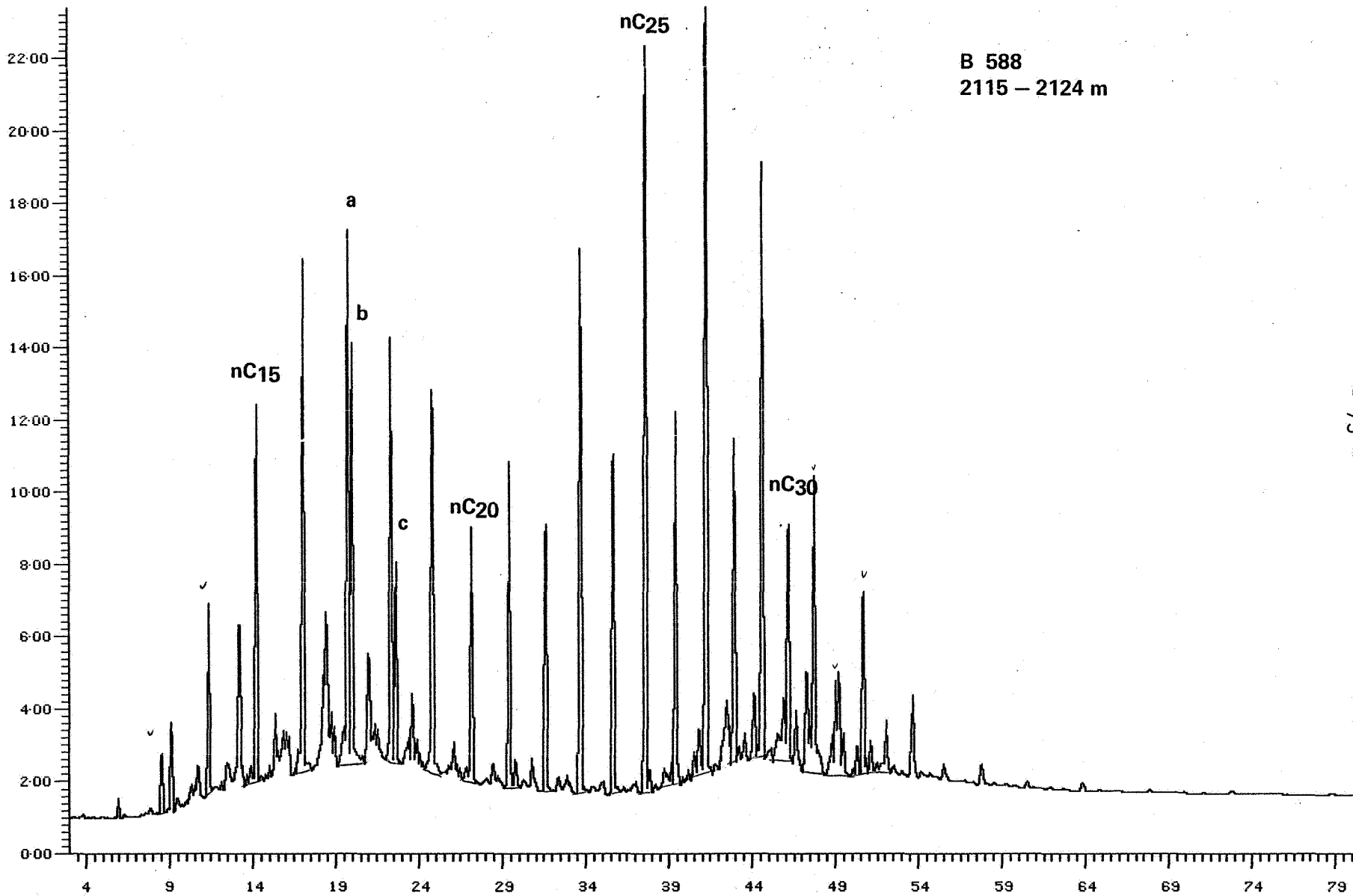
Data Scale Plot

Box 1 of 1

Analysis : 198B588S1 Sample f: 1 Injection f: 1

Sample Name : B-588,SAT,31/5-1,AD

Maximum signal (%): 23.392



B 588
2115 - 2124 m

Created at 14:48 on 15/Feb/84

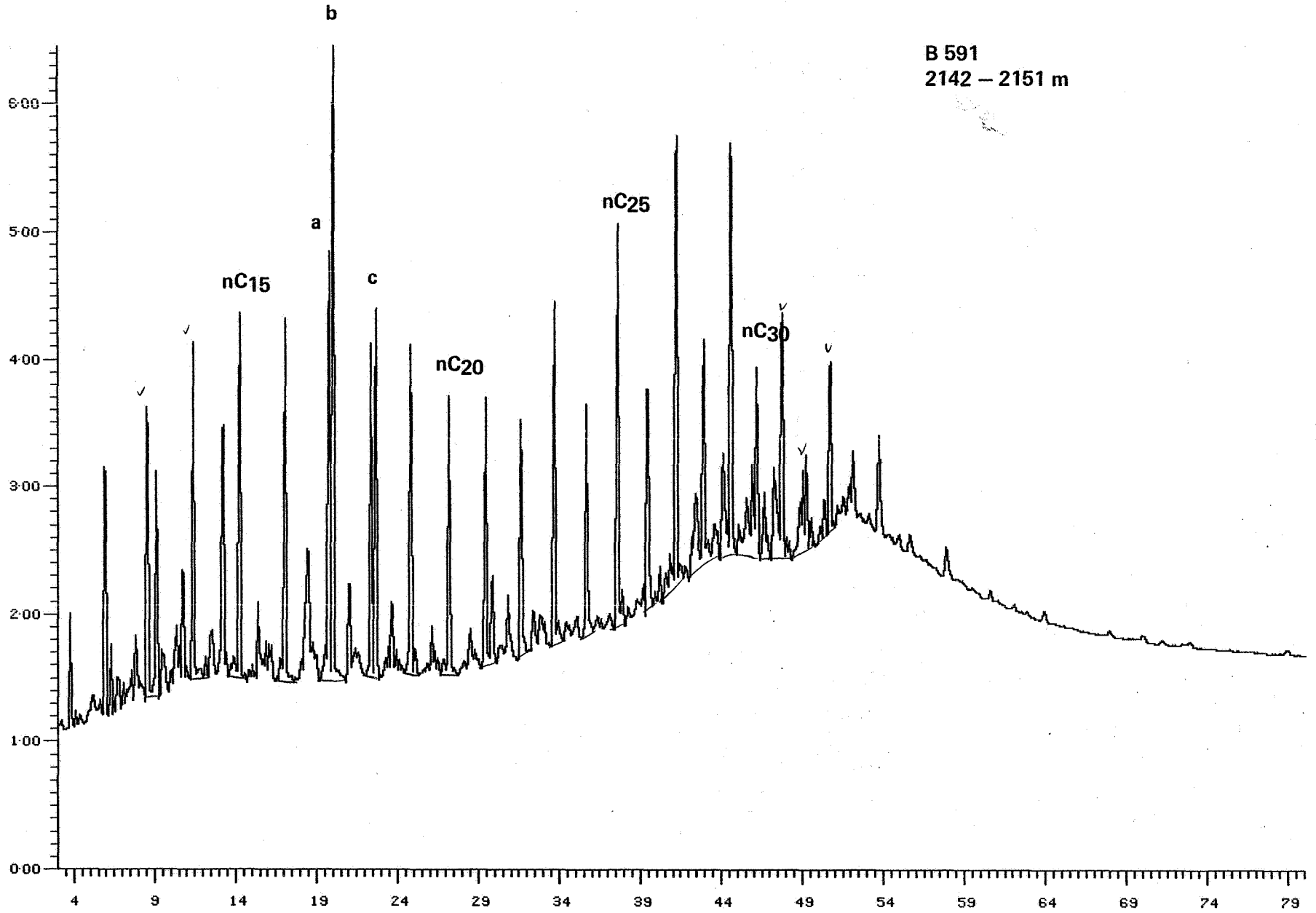
RAW DATA PLOT-CHANNEL 3

Data Scale Plot Box 1 of 1

Analysis : 198B591S1 Sample f: 1 Injection f: 1

Sample Name : B-591, SAT, 31/5-1, AD

Maximum signal <%> : 6.450



Created at 13:37 on 24/Feb/84

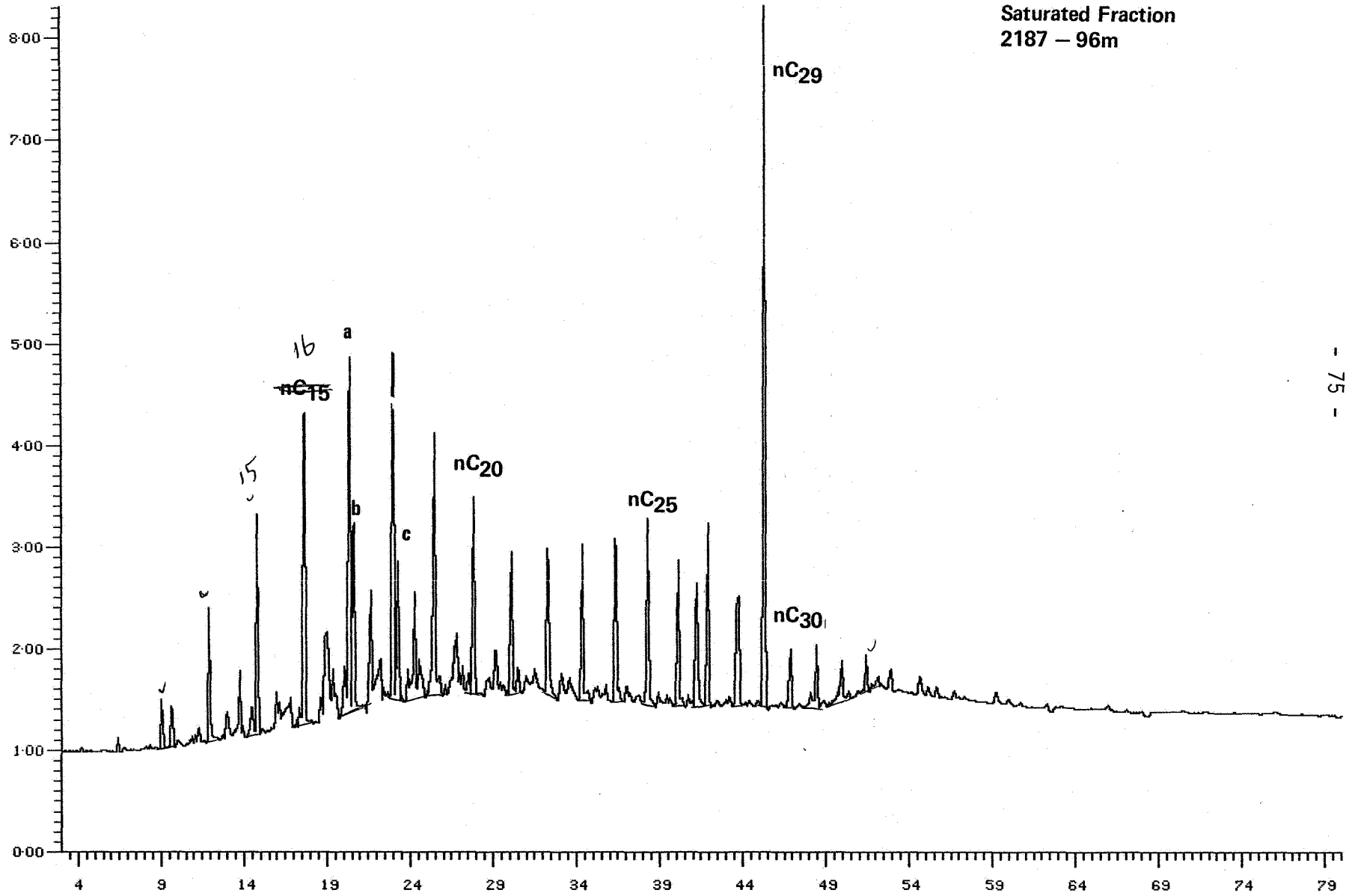
RAW DATA PLOT-CHANNEL 3

Data Scale Plot Box 1 of 1

Analysis : 198B596S1 Sample f: 1 Injection f: 1

Sample Name : B-596,SAT,31/5-1,AD

Maximum signal (%): 8.315



B 596
Saturated Fraction
2187 - 96m

Created at 08:40 on 15/Feb/84

RAW DATA PLOT-CHANNEL 3
Data Scale Plot

Box 1 of 1

Analysis : 198B598S1 Sample f: 1 Injection f: 1

Sample Name : B-598,SAT,31/5-1,AD

Maximum signal [%] : 3.605

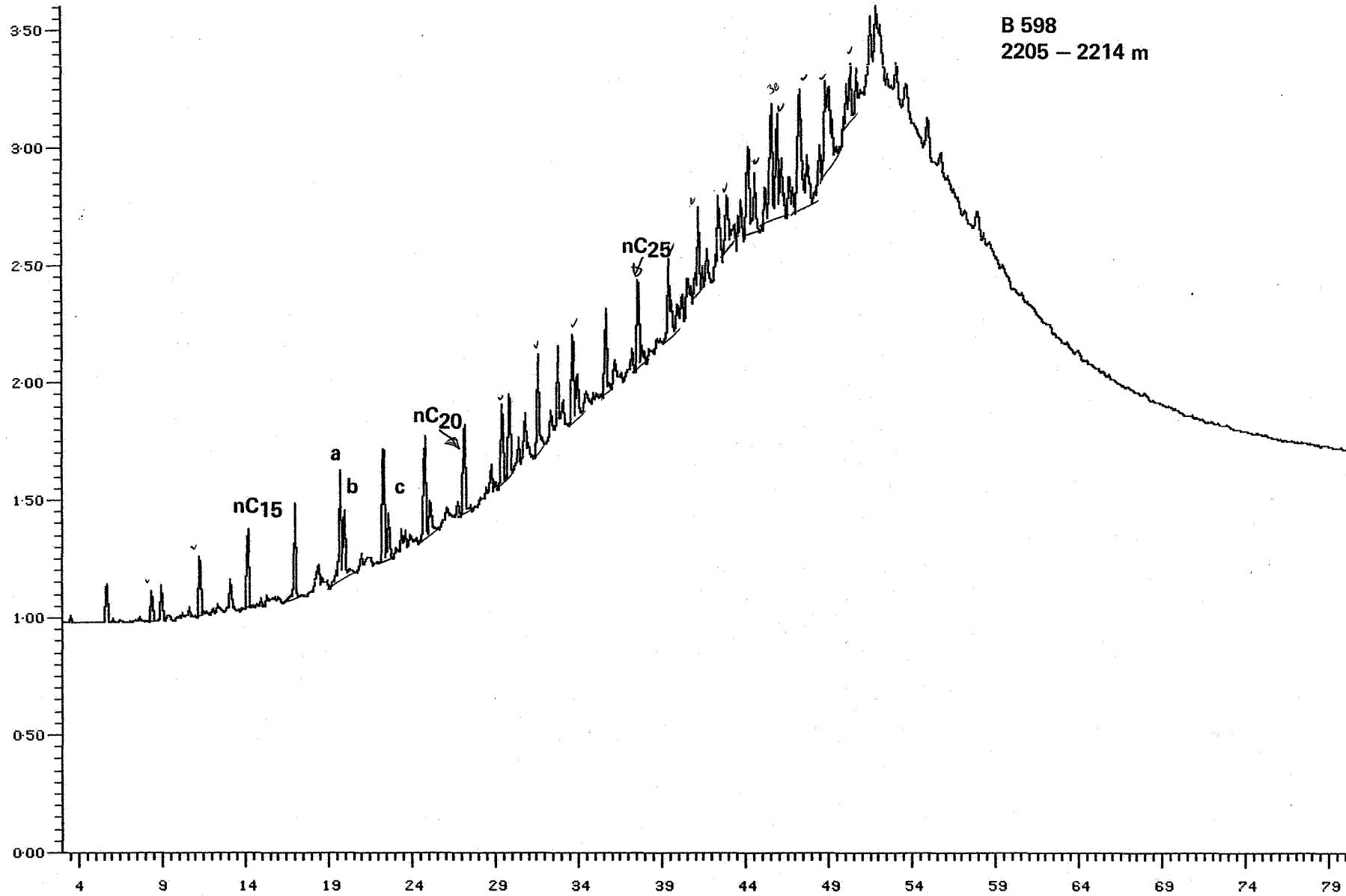


FIGURE 2

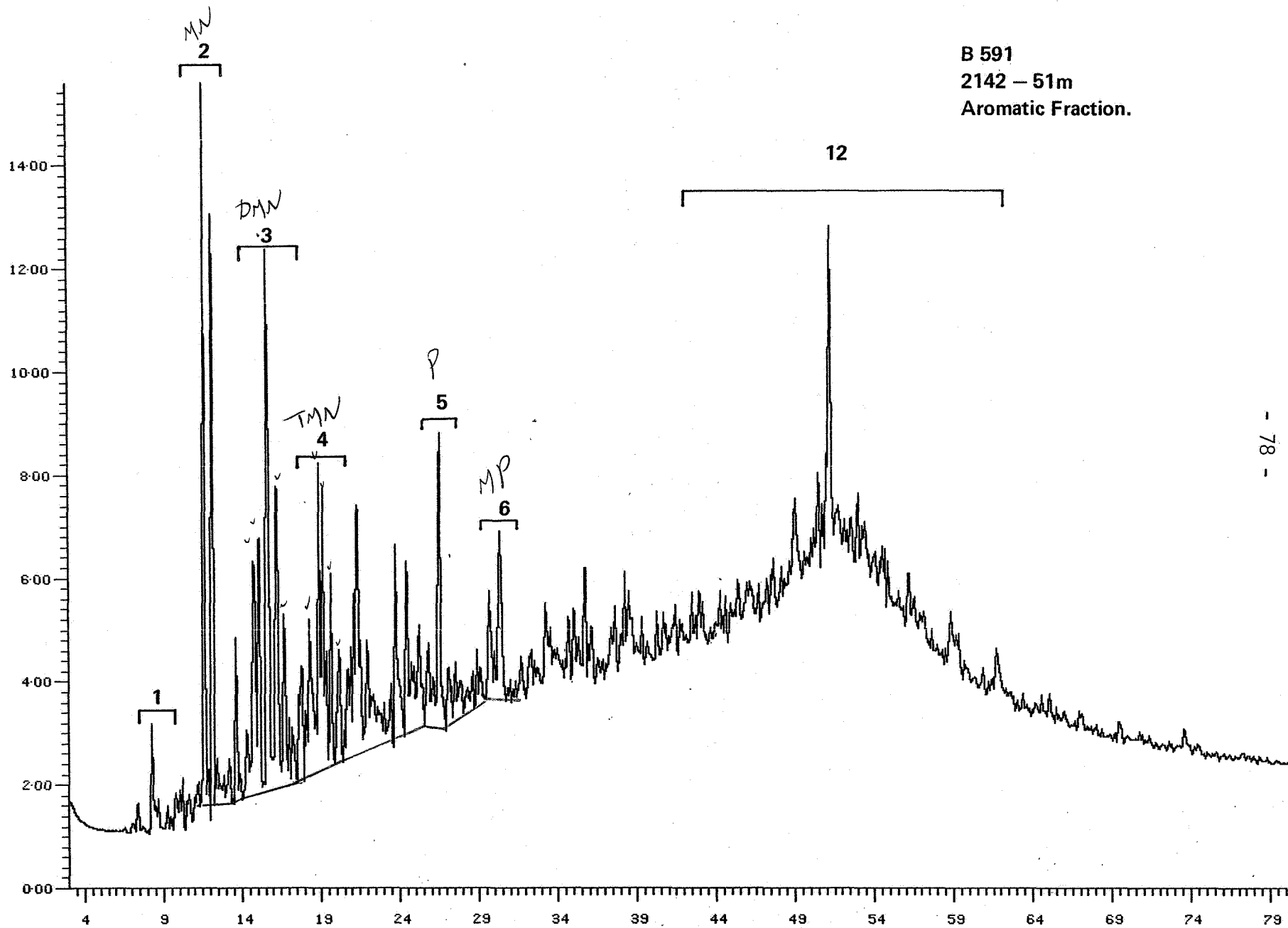
Aromatic hydrocarbon gas chromatograms

- 1 = naphthalene
- 2 = methyl naphthalenes
- 3 = dimethyl naphthalenes
- 4 = trimethyl naphthalenes
- 5 = phenanthrene
- 6 = methyl phenanthrenes

Created at 08:44 on 20/Feb/84

RAW DATA PLOT-CHANNEL 4
Data Scale Plot Box 1 of 1

Analysis : 198B591A1 Sample f: 1 Injection f: 1
Sample Name : B-591,ARO,31/5-1,AD Maximum signal (%): 15.595



B 591
2142 - 51m
Aromatic Fraction.

Created at 08:52 on 20/Feb/84

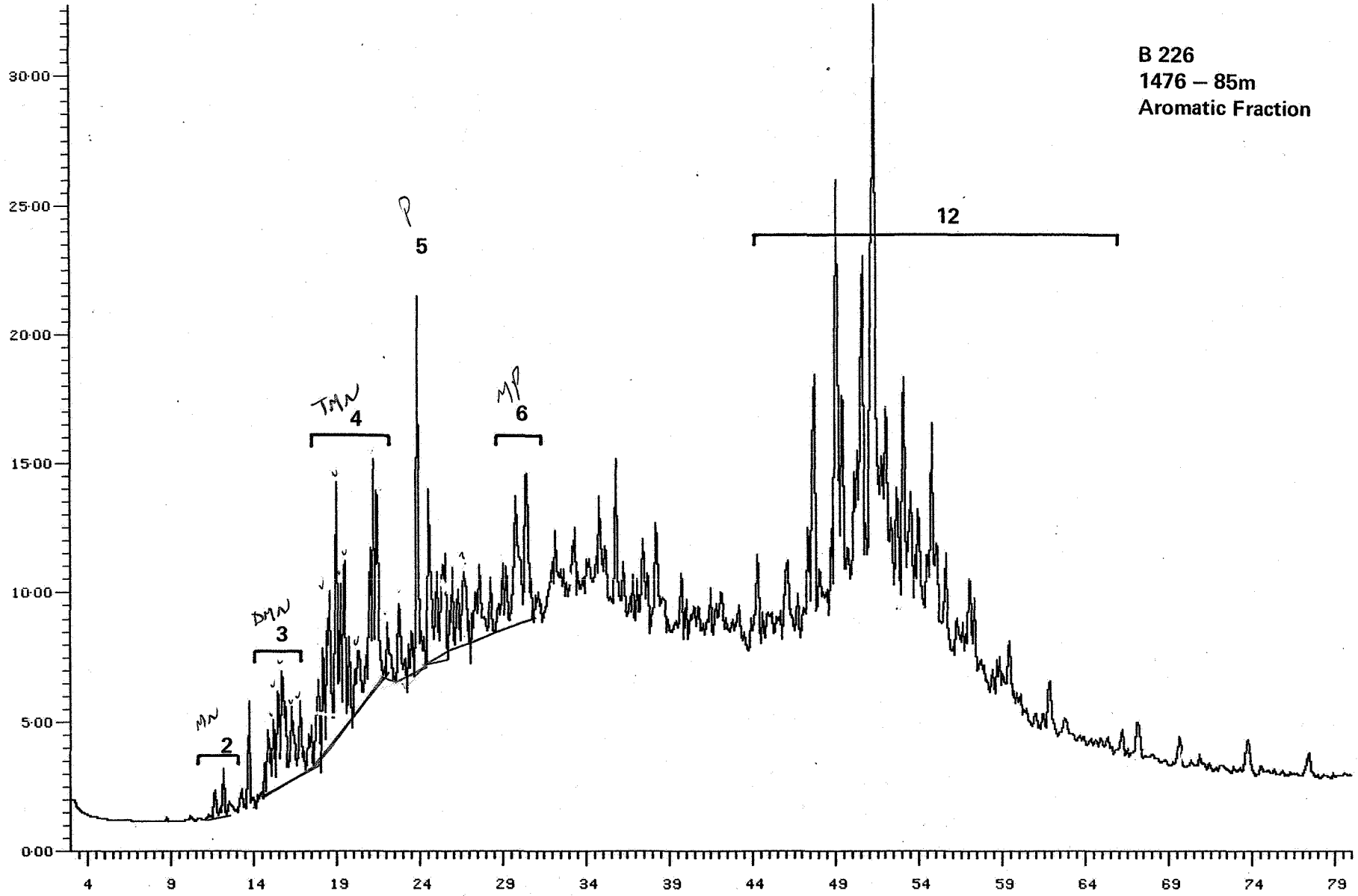
RAW DATA PLOT-CHANNEL 4

Data Scale Plot Box 1 of 1

Analysis : 198B226A1 Sample f: 1 Injection f: 1

Sample Name : B-226, ARD, 31/5-1, AD

Maximum signal (%): 32.735



B 226
1476 - 85m
Aromatic Fraction

Created at 13:47 on 17/Feb/84

RAW DATA PLOT-CHANNEL 4

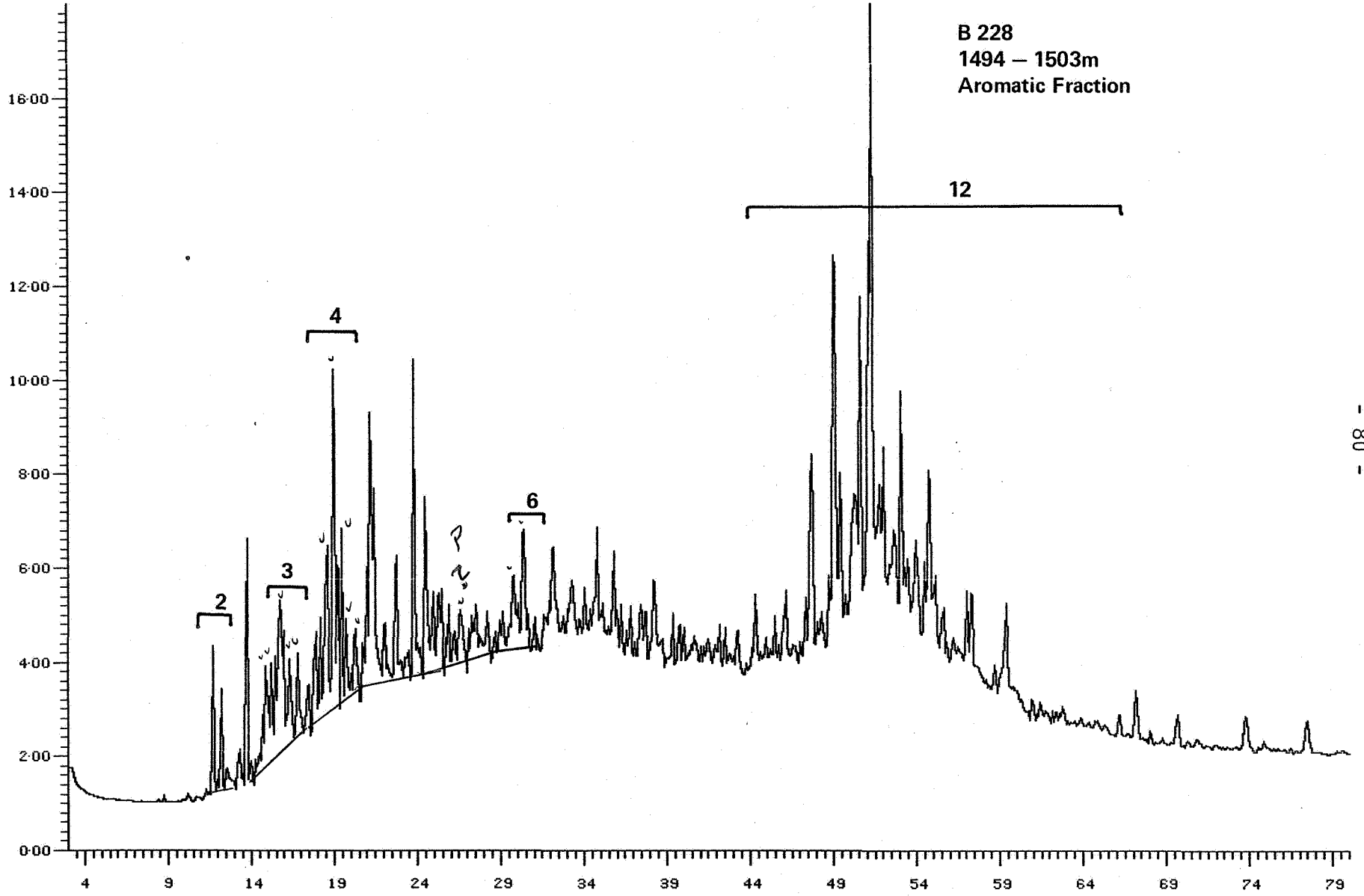
Data Scale Plot

Box 1 of 1

Analysis : 198B228A1 Sample f: 1 Injection f: 1

Sample Name : B-228, ARO, 31/5-1, AD

Maximum signal < % > : 17.999



Created at 11:20 on 17/Feb/84

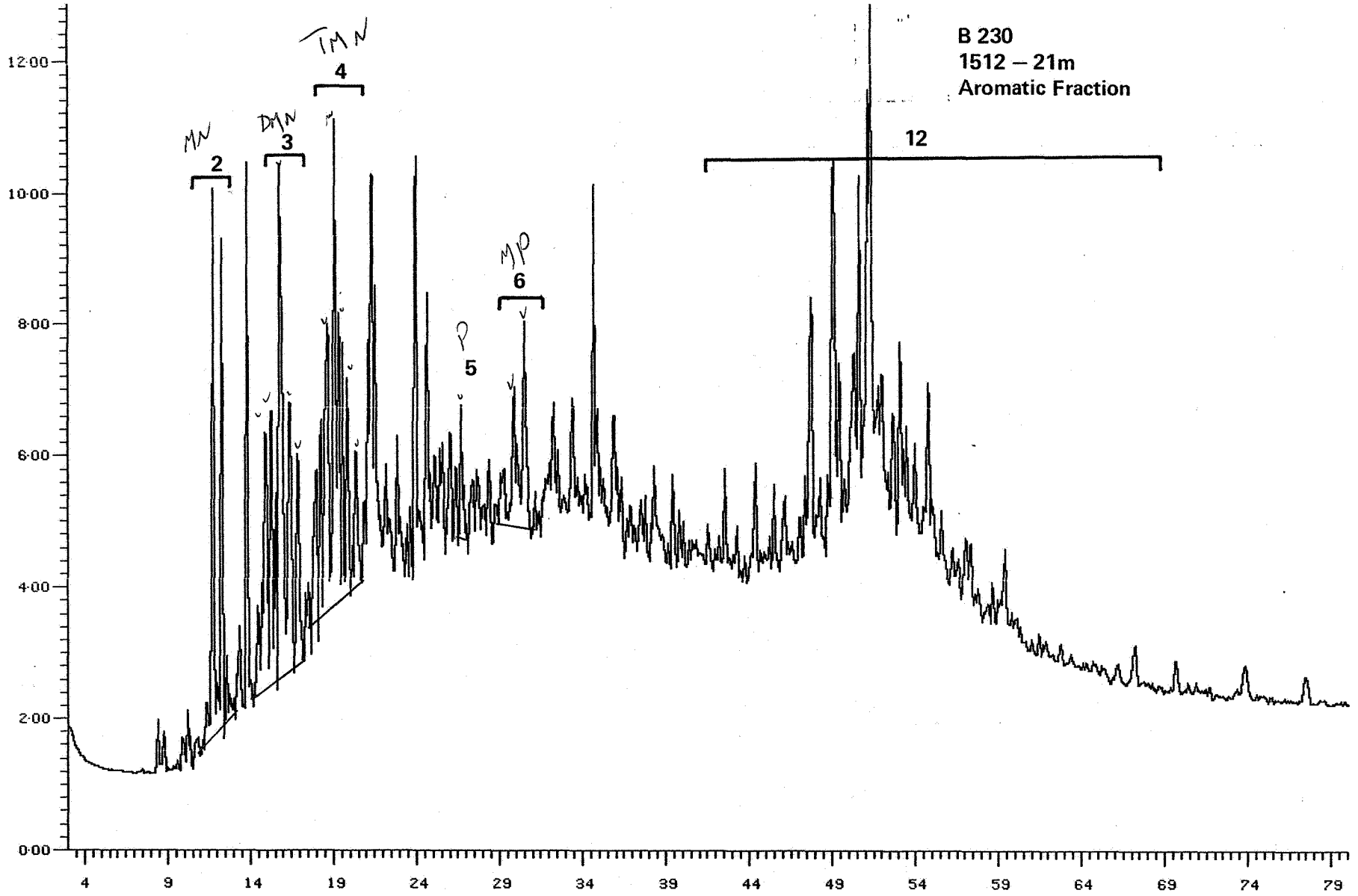
RAW DATA PLOT-CHANNEL 4

Data Scale Plot Box 1 of 1

Analysis : 198B230R1 Sample f: 1 Injection f: 1

Sample Name : B-230,ARO,31/5-1,AD

Maximum signal {>} : 12.862



Created at 15:47 on 16/Feb/84

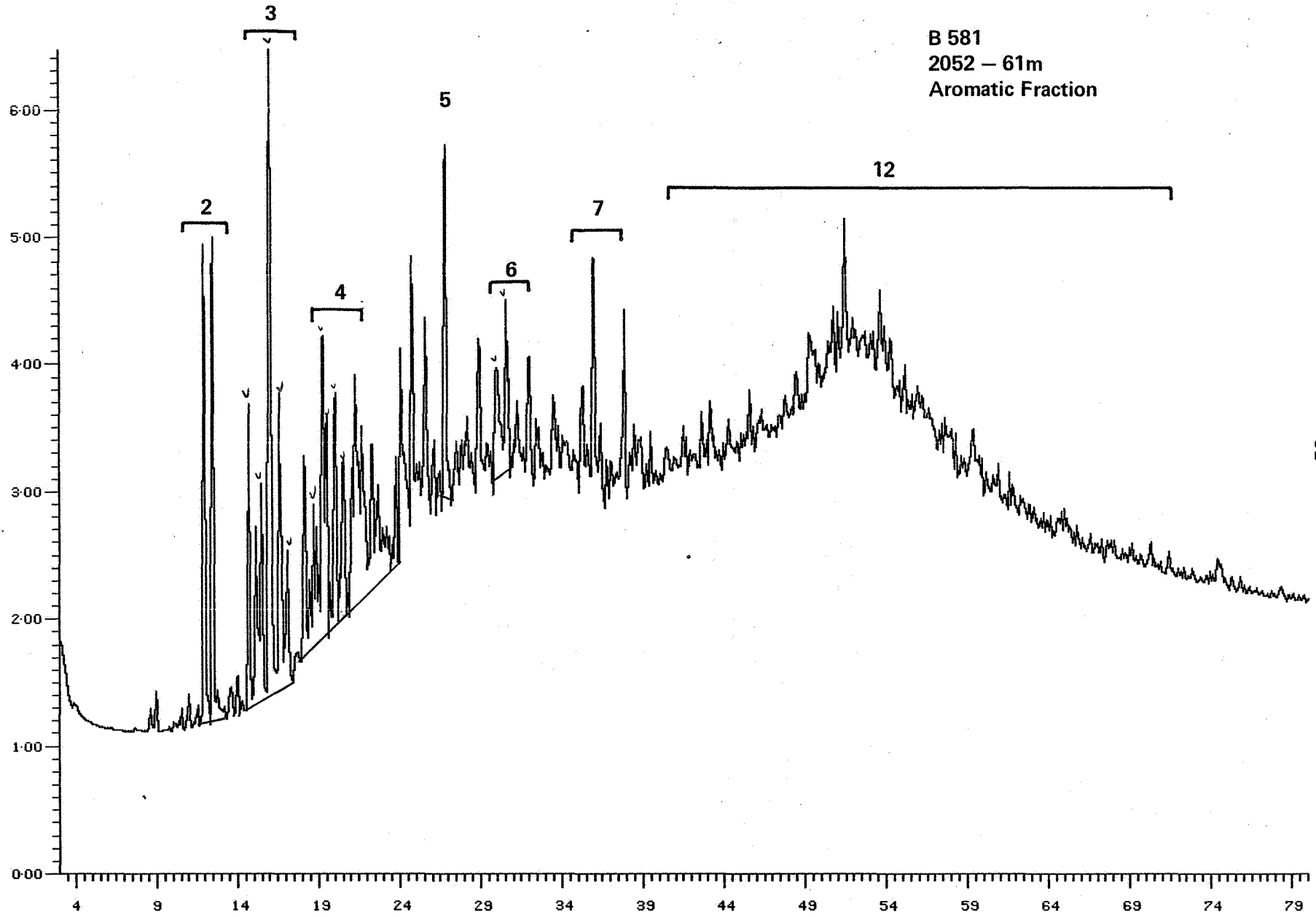
RAW DATA PLOT-CHANNEL 4

Data Scale Plot Box 1 of 1

Analysis : 198B581A1 Sample f: 1 Injection f: 1

Sample Name : B-581,ARO,31/5-1,AD

Maximum signal {>} : 6.467



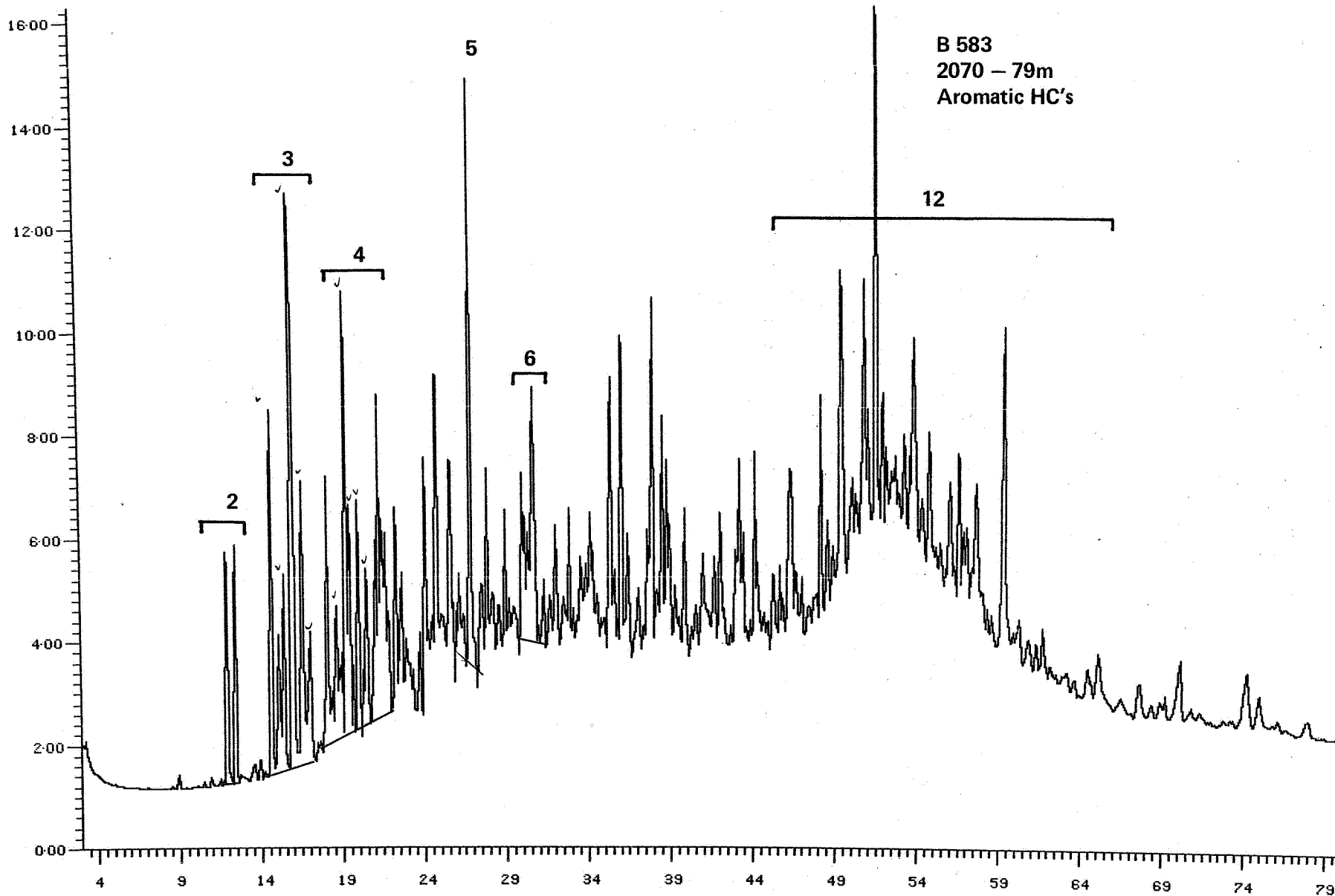
B 581
2052 - 61m
Aromatic Fraction

Created at 07:56 on 17/Feb/84

RAW DATA PLOT-CHANNEL 4
Data Scale Plot

Box 1 of 1

Analysis : 198B583A1 Sample f: 1 Injection f: 1
Sample Name : B-583,ARO,31/5-1,AD Maximum signal (%): 16.313



Created at 15:48 on 17/Feb/84

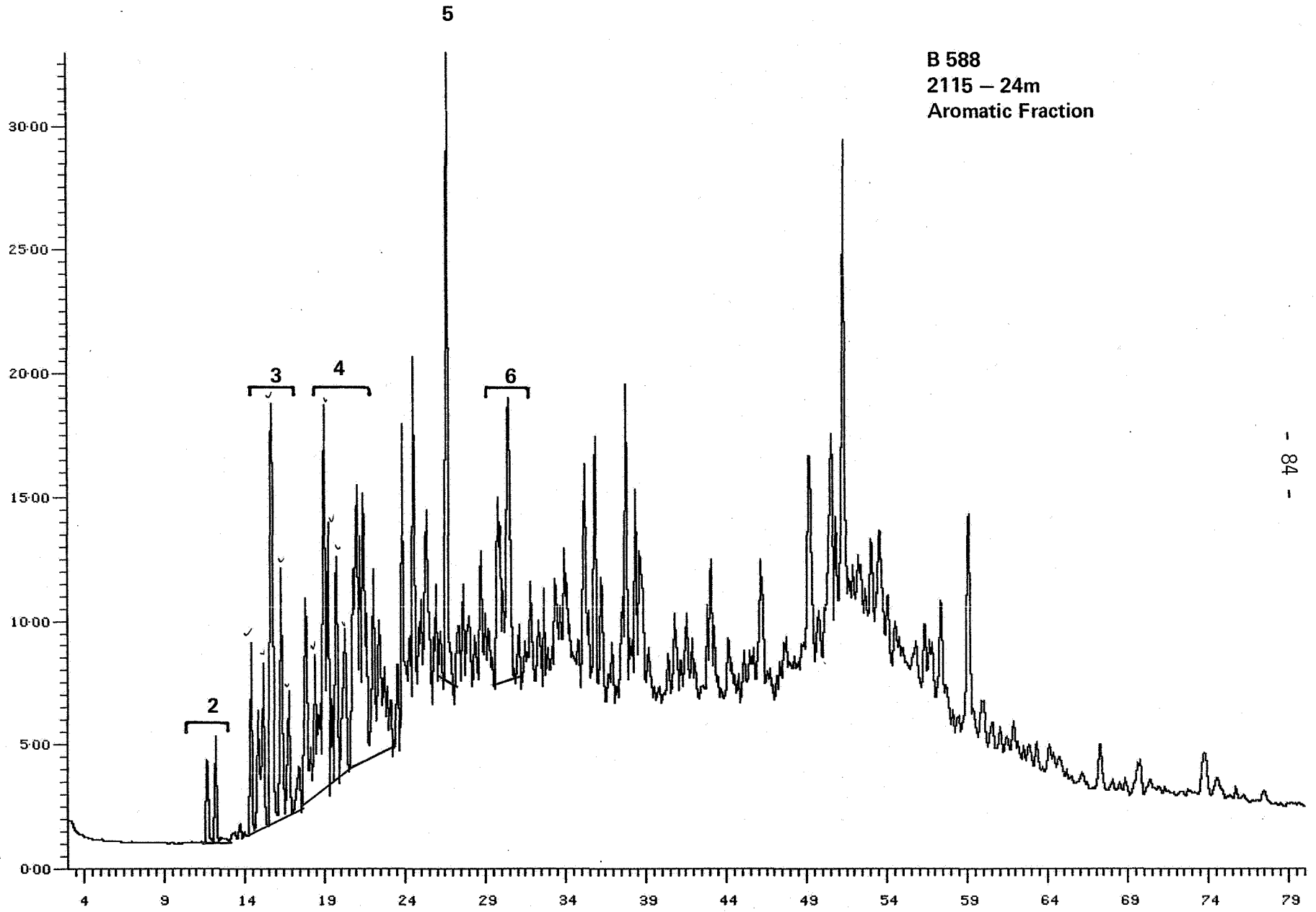
RAW DATA PLOT-CHANNEL 4

Data Scale Plot Box 1 of 1

Analysis :198B588A1 Sample f: 1 Injection f: 1

Sample Name :B-588,ARO,31/5-1,AD

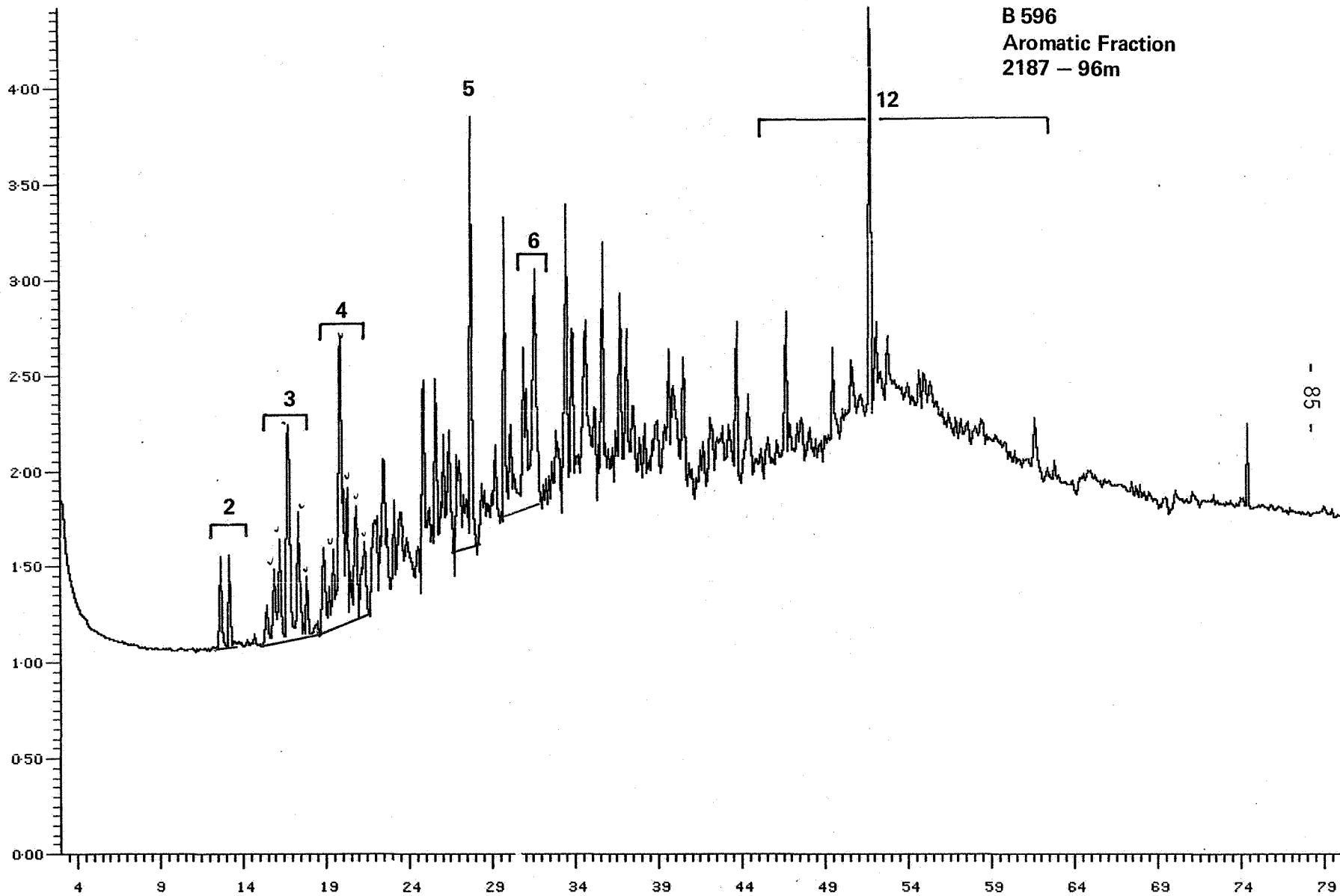
Maximum signal (%):32.930



Created at 13:34 on 24/Feb/84

RAW DATA PLOT-CHANNEL 4
Data Scale Plot Box 1 of 1

Analysis : 198B596R1 Sample f: 1 Injection f: 1
Sample Name : B-596, AR0, 31/5-1, AD Maximum signal (%): 4.420



B 596
Aromatic Fraction
2187 - 96m

Created at 08:56 on 20/Feb/84

RAW DATA PLOT-CHANNEL 4

Data Scale Plot Box 1 of 1

Analysis : 198B598A1 Sample f: 1 Injection f: 1

Sample Name : B-598,ARO,31/2-1,AD

Maximum signal <%> : 6.496

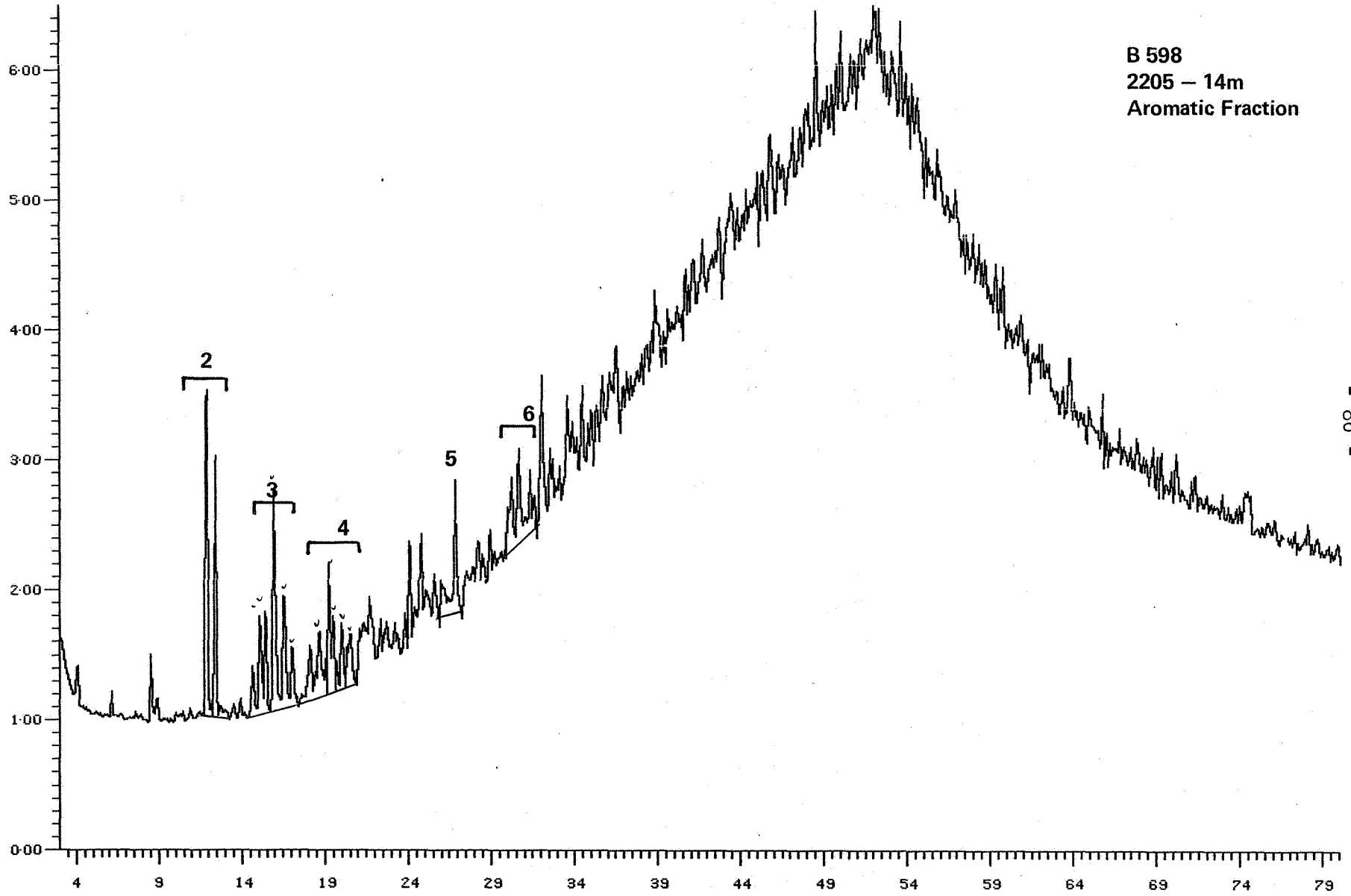
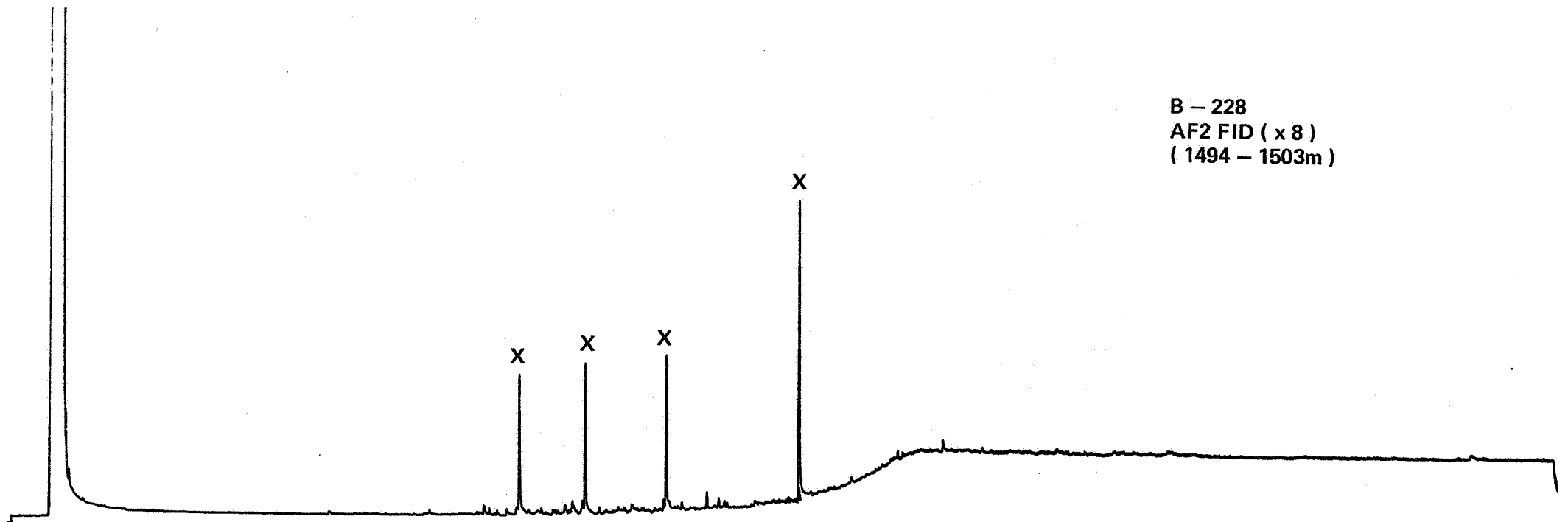


FIGURE 3

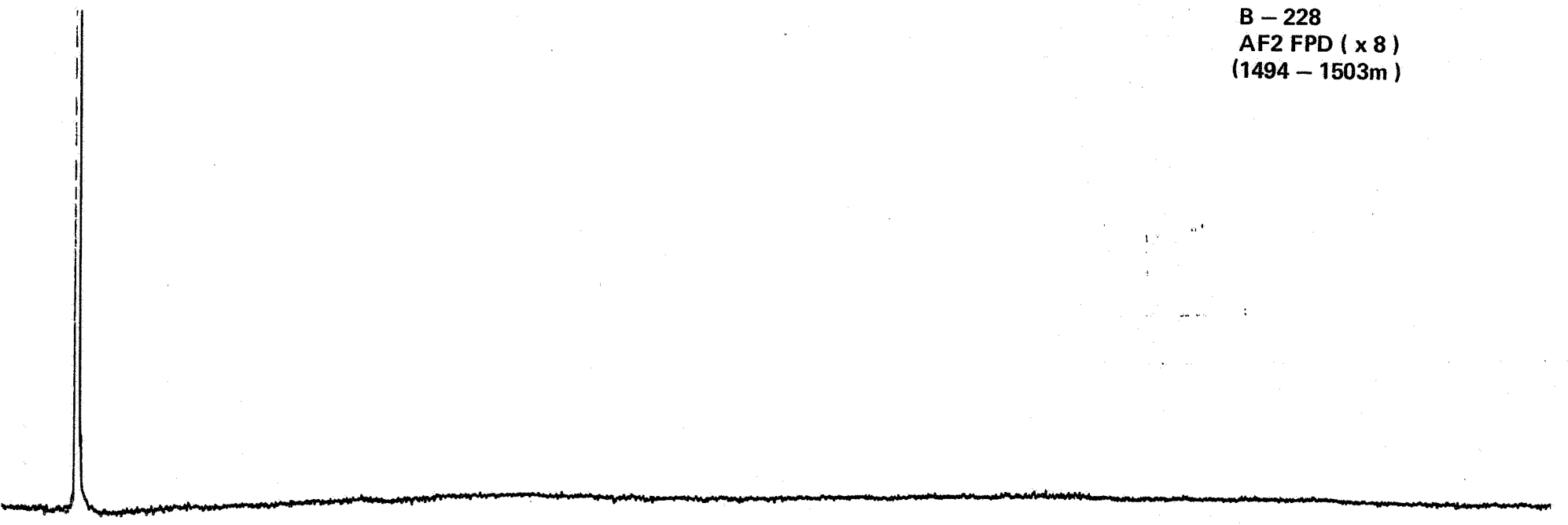
AF2 hydrocarbon chromatograms (FID and FPD)

- x = contaminant
- P = phenanthrene
- MP = methyl phenanthrene
- DMP = dimethyl phenanthrene
- DBT = dibenzothiophene
- MDBT = methyldibenzothiophene
- DMDBT = dimethyldibenzothiophene

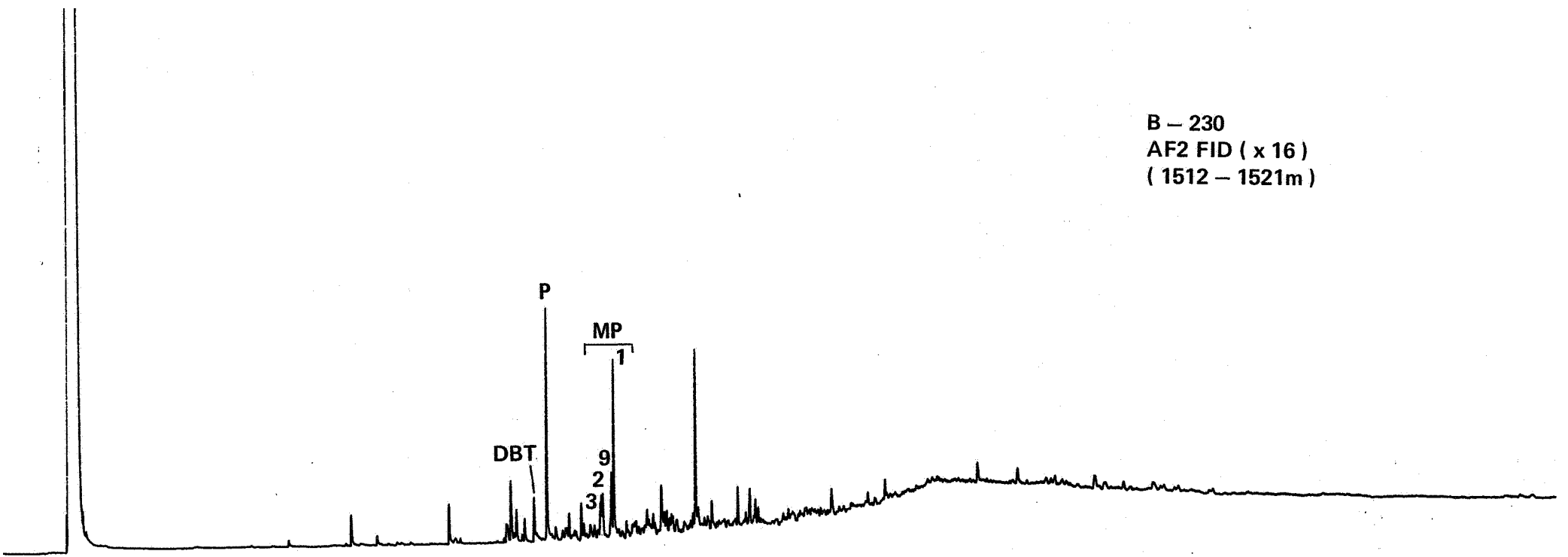
B - 228
AF2 FID (x 8)
(1494 - 1503m)



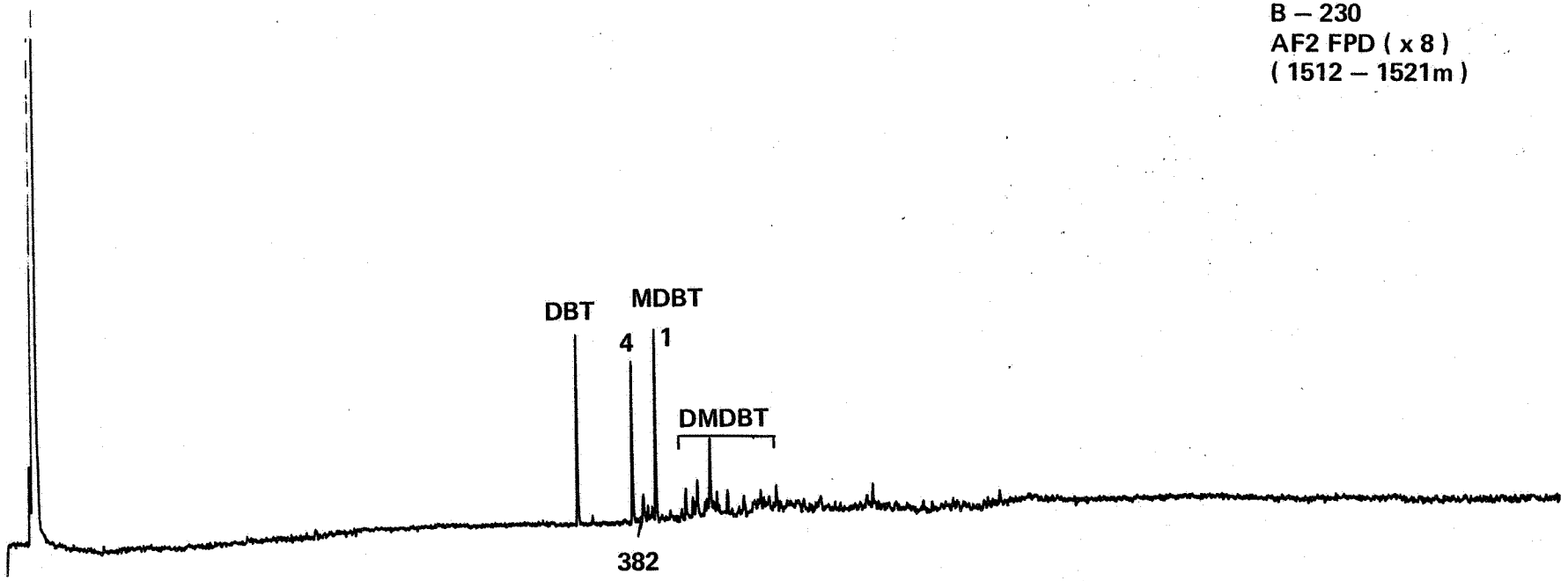
B - 228
AF2 FPD (x 8)
(1494 - 1503m)

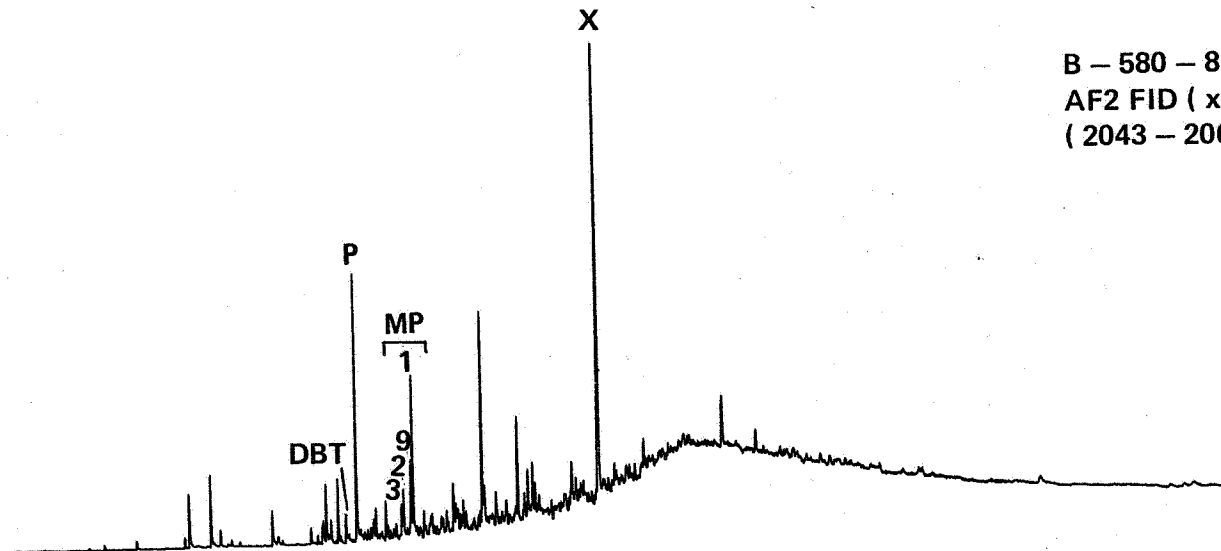


B - 230
AF2 FID (x 16)
(1512 - 1521m)

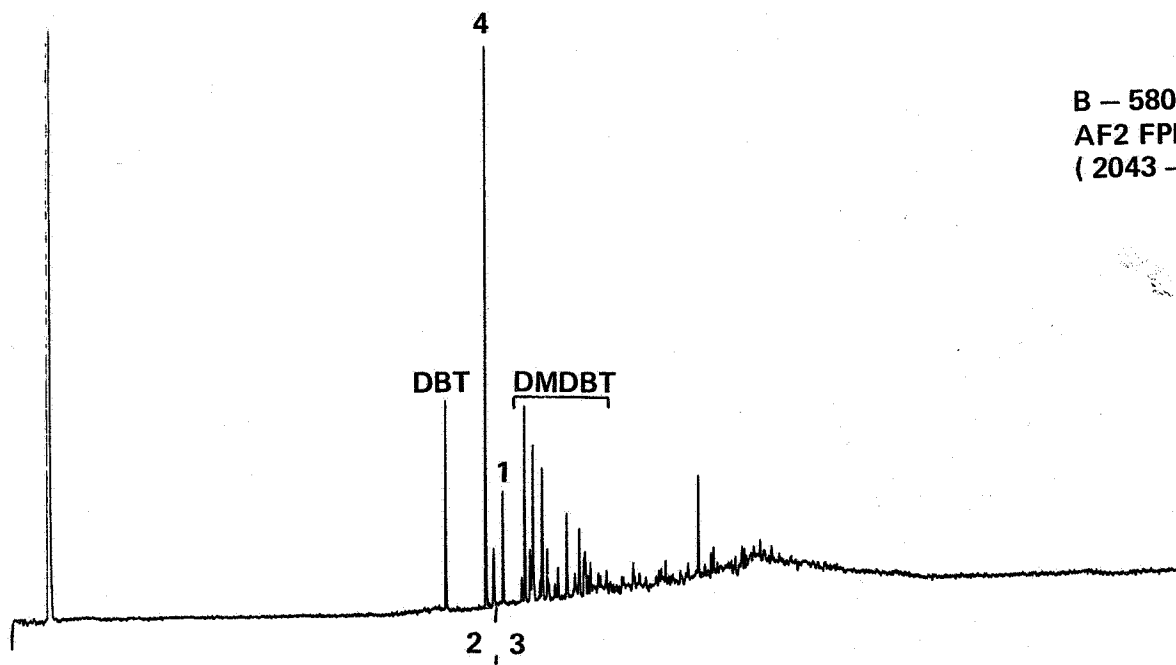


B - 230
AF2 FPD (x 8)
(1512 - 1521m)



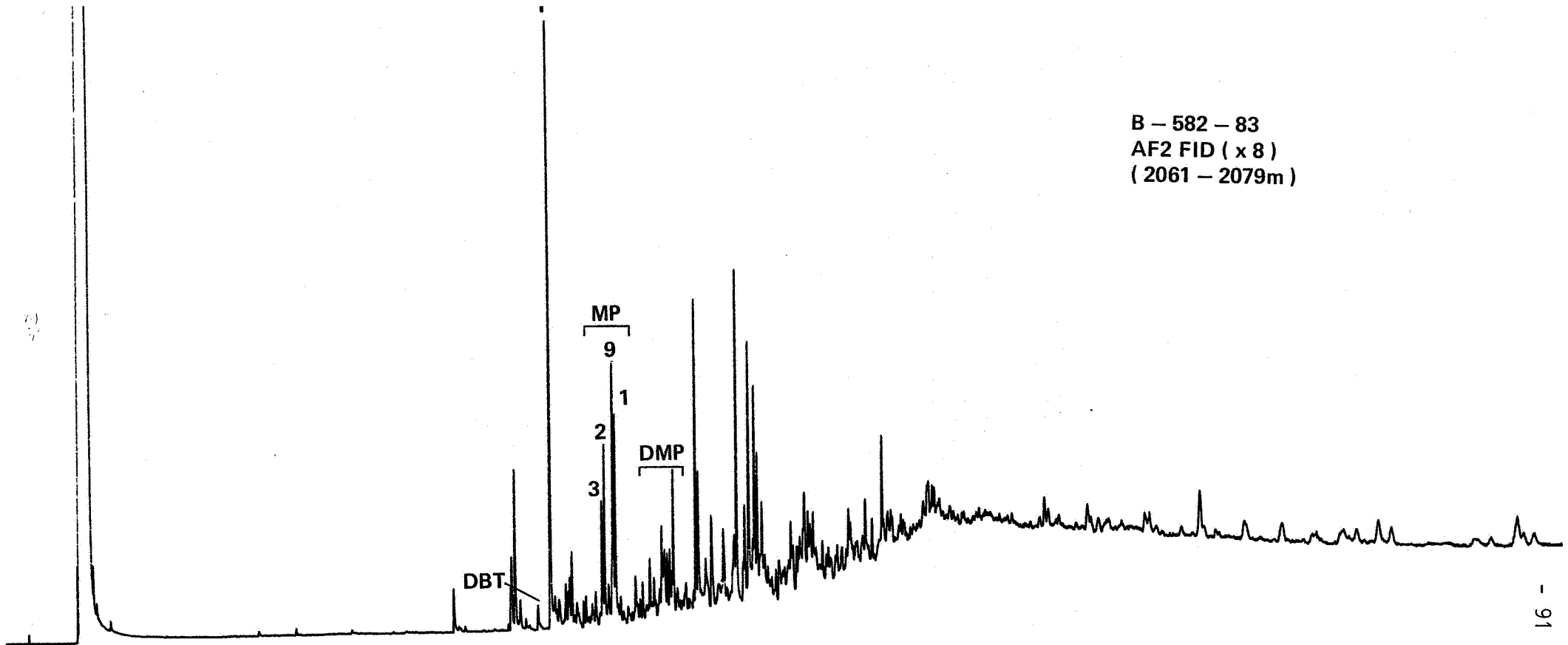


B - 580 - 81
AF2 FID (x 16)
(2043 - 2061m)

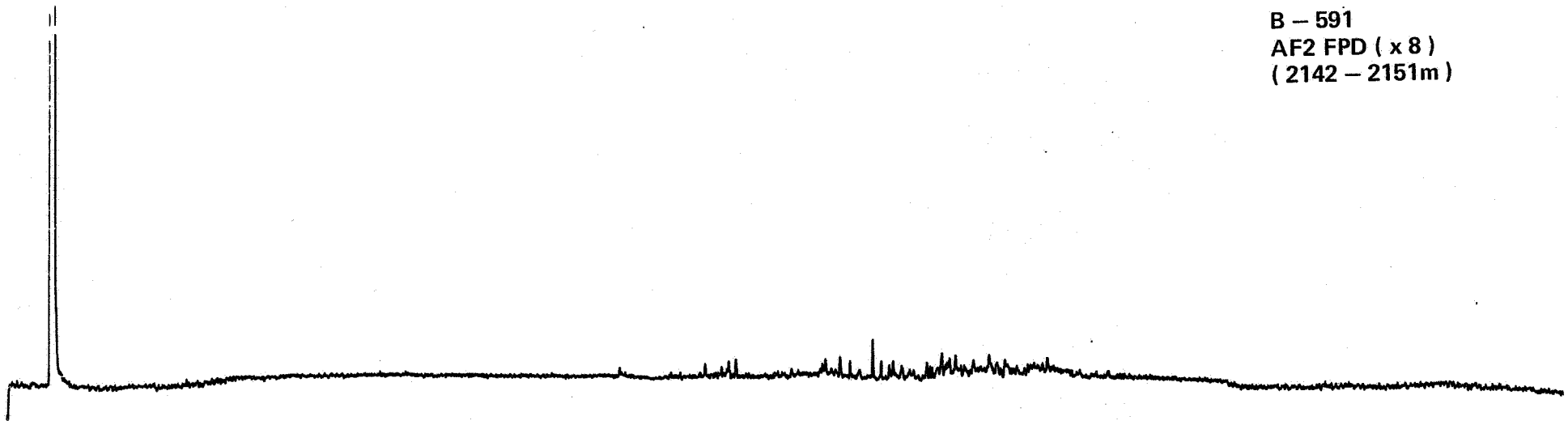
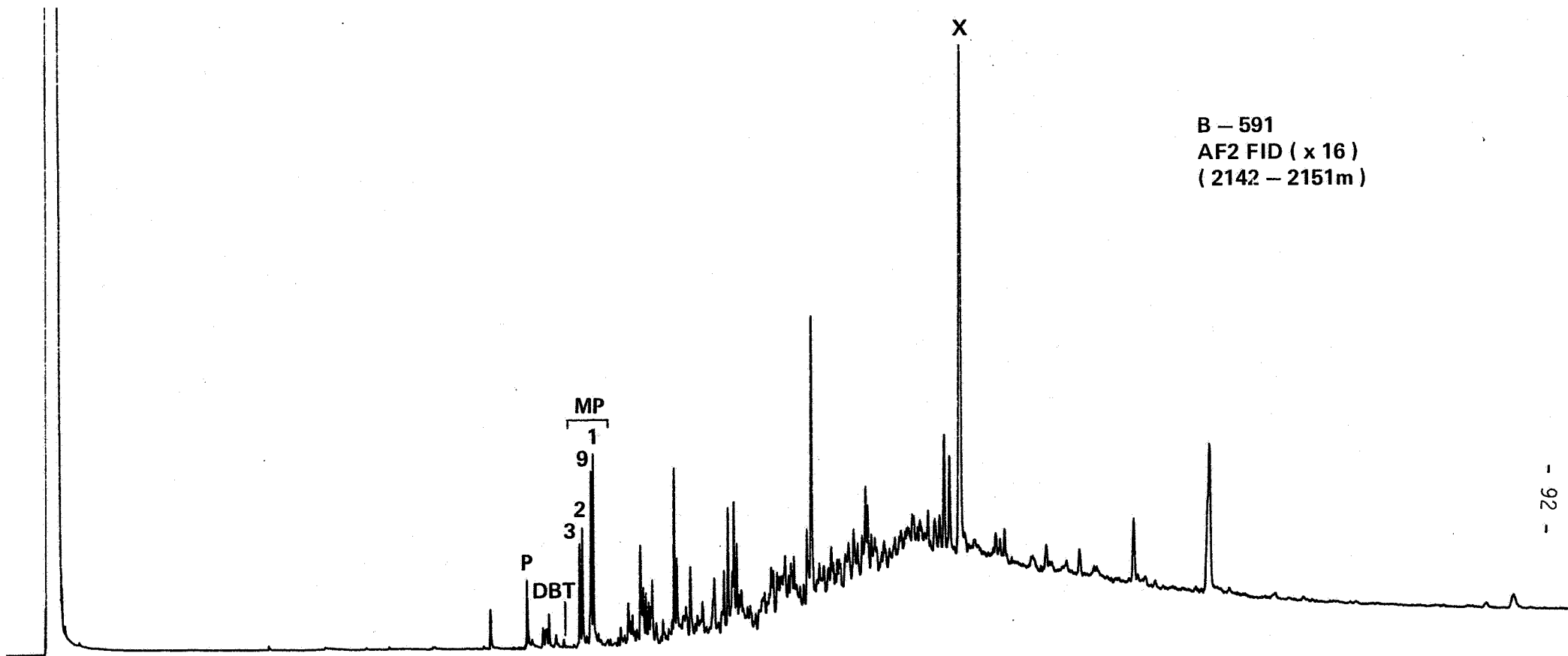


B - 580 - 81
AF2 FPD (x 8)
(2043 - 2061m)

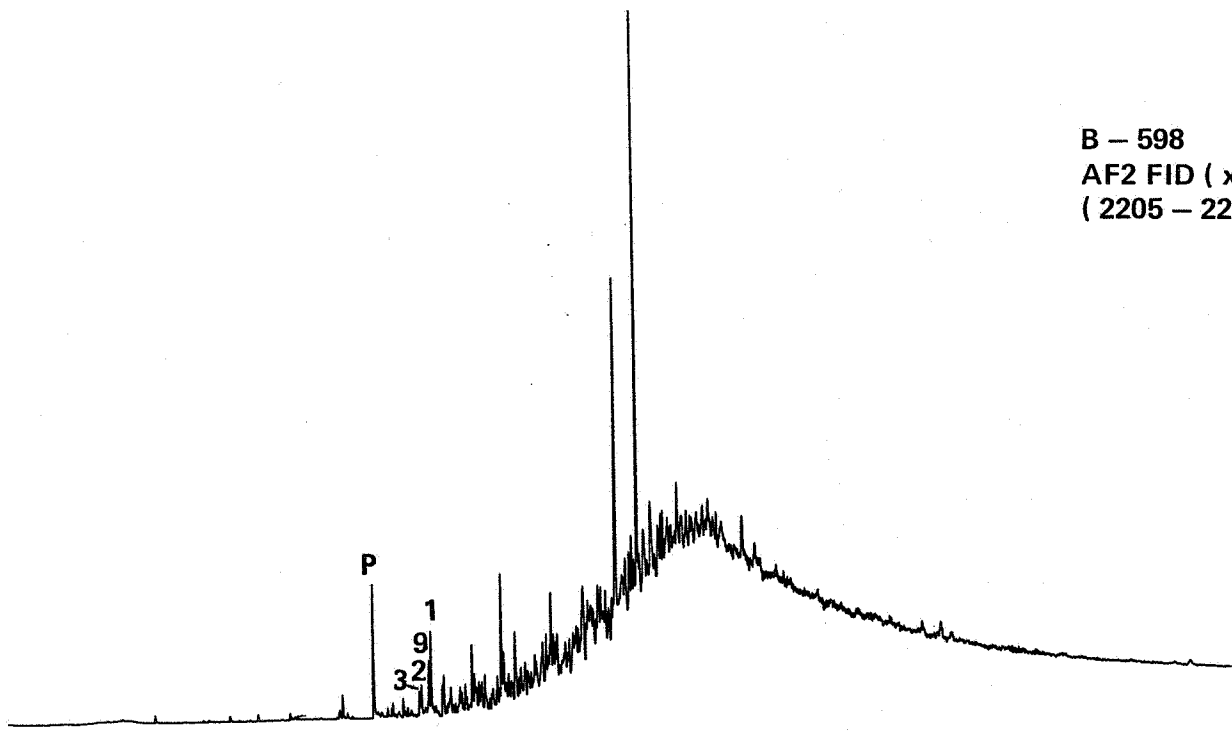
B - 582 - 83
AF2 FID (x 8)
(2061 - 2079m)



B - 582 - 83
AF2 FPD (x 8)
(2061 - 2079 m)



B - 598
AF2 FID (x 16)
(2205 - 2214m)



B - 598
AF2 FPD (x 8)
(2205 - 2214m)

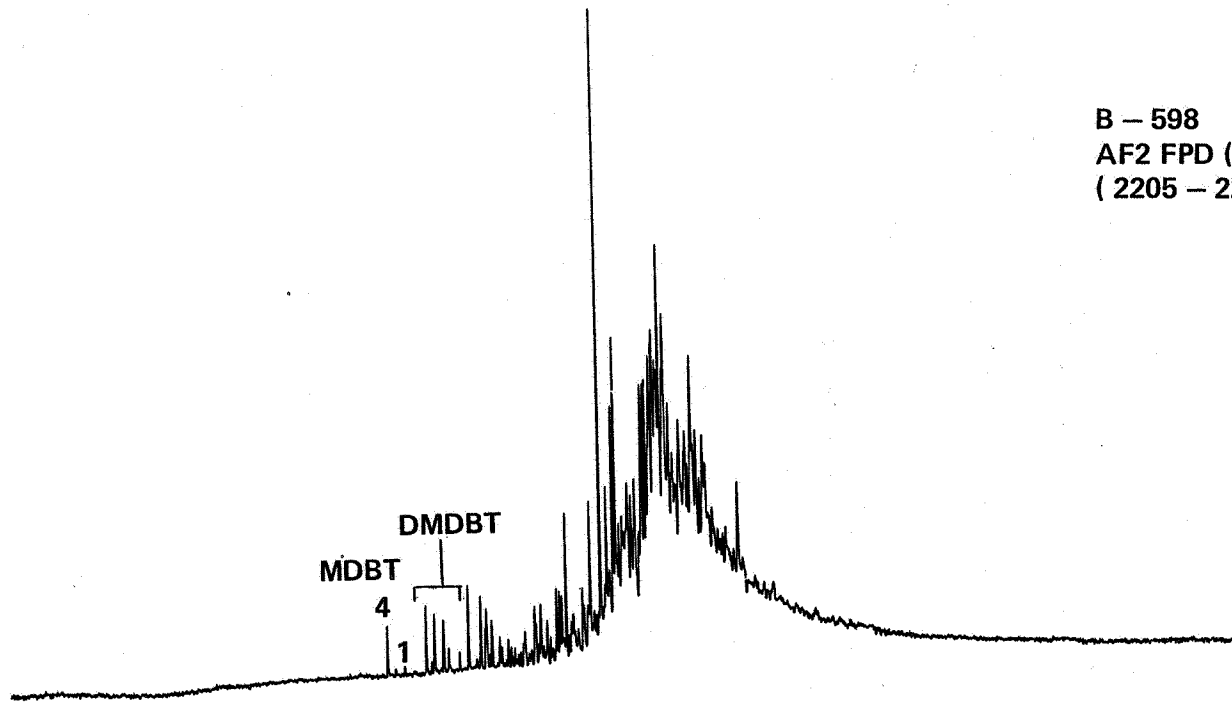
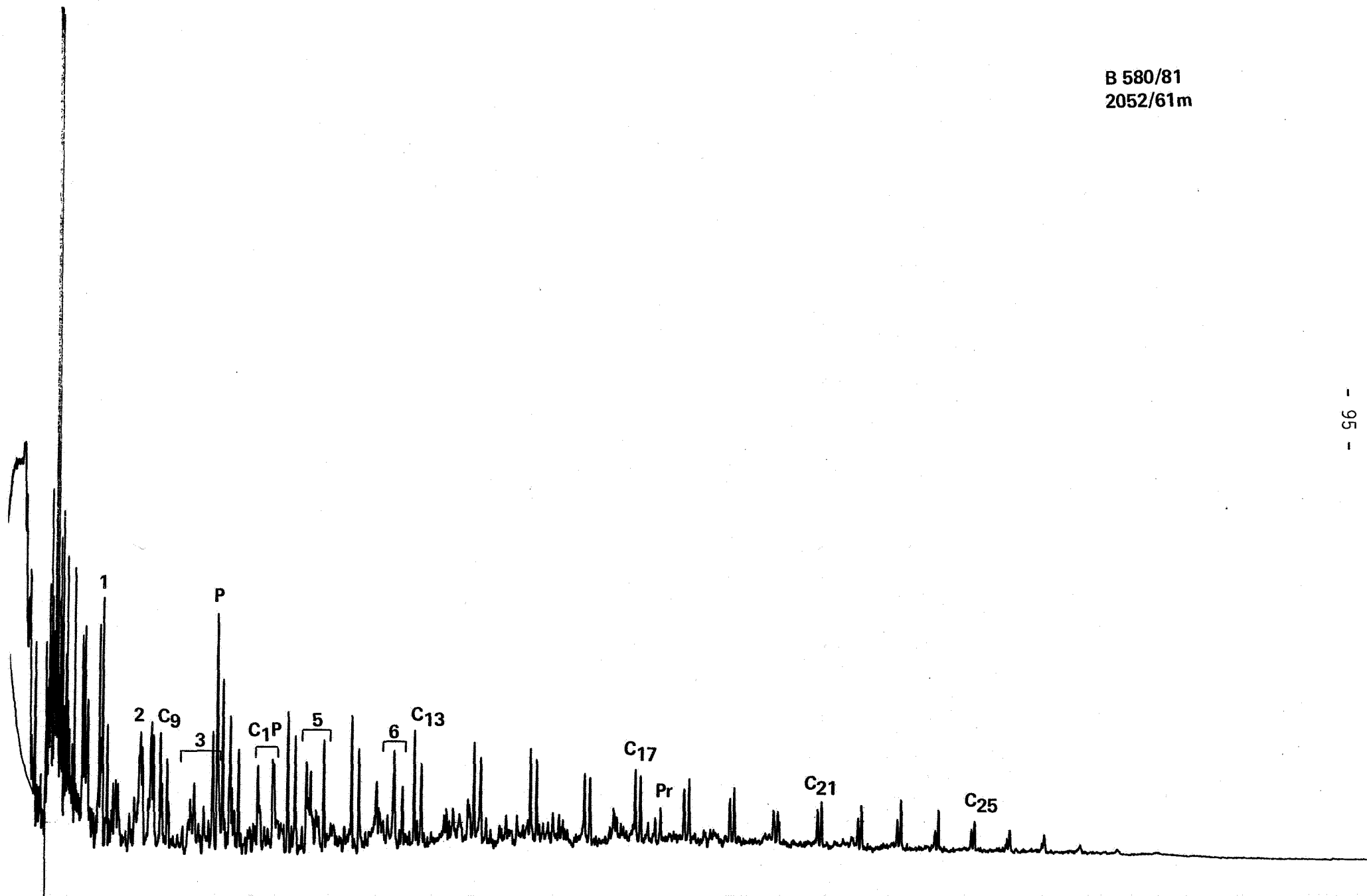


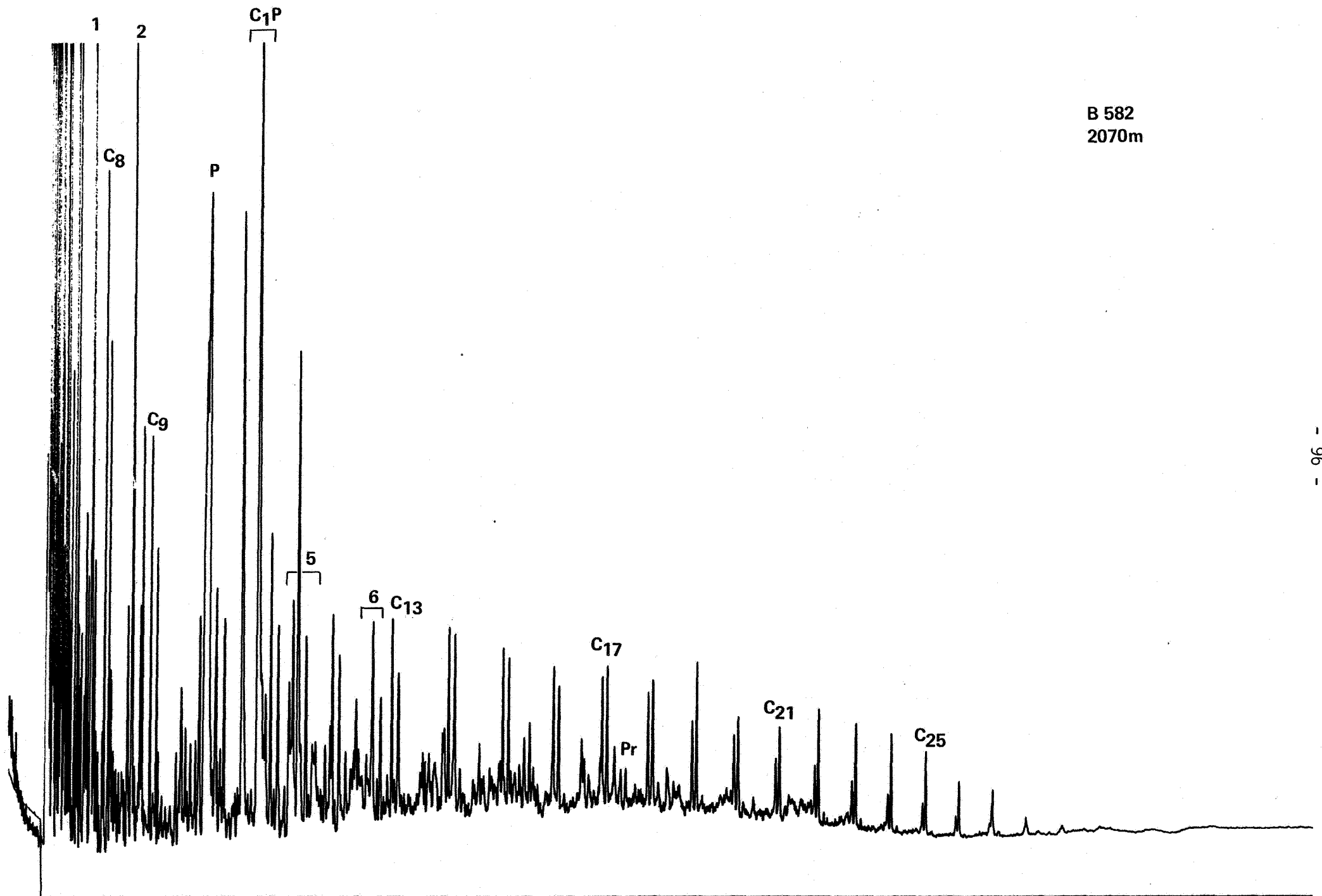
FIGURE 4

Pyrolysis-gc chromatograms

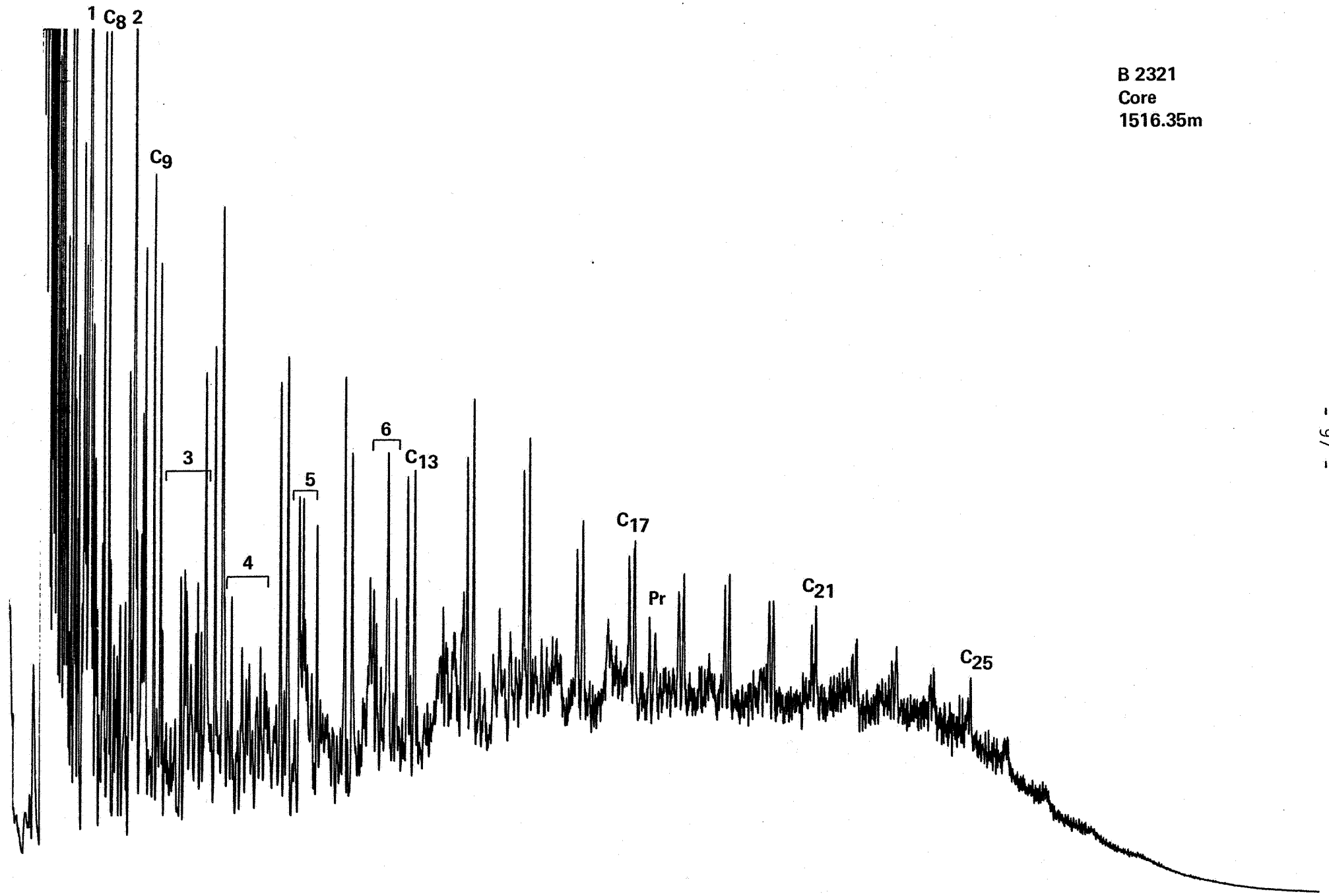
B 580/81
2052/61m



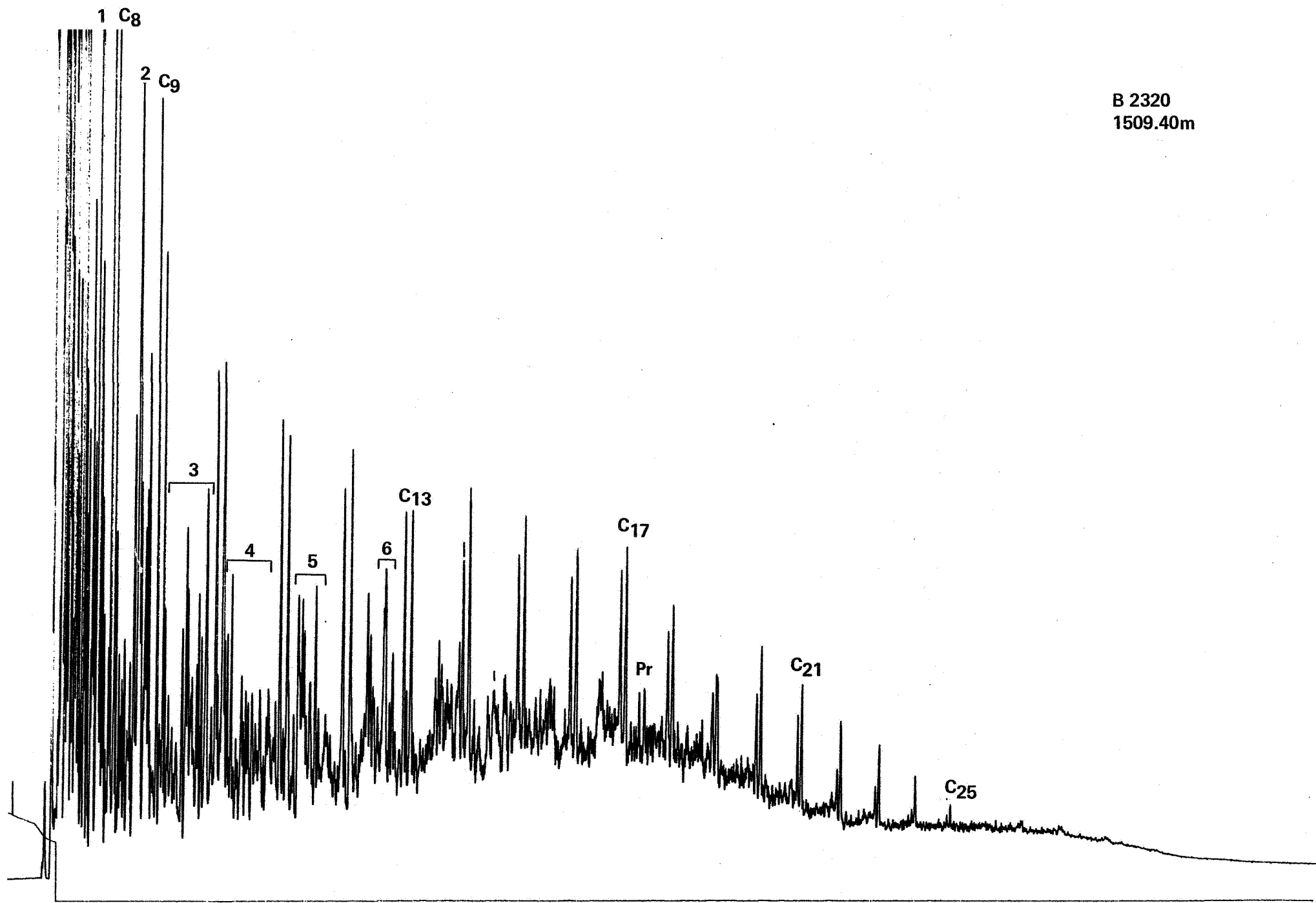
B 582
2070m



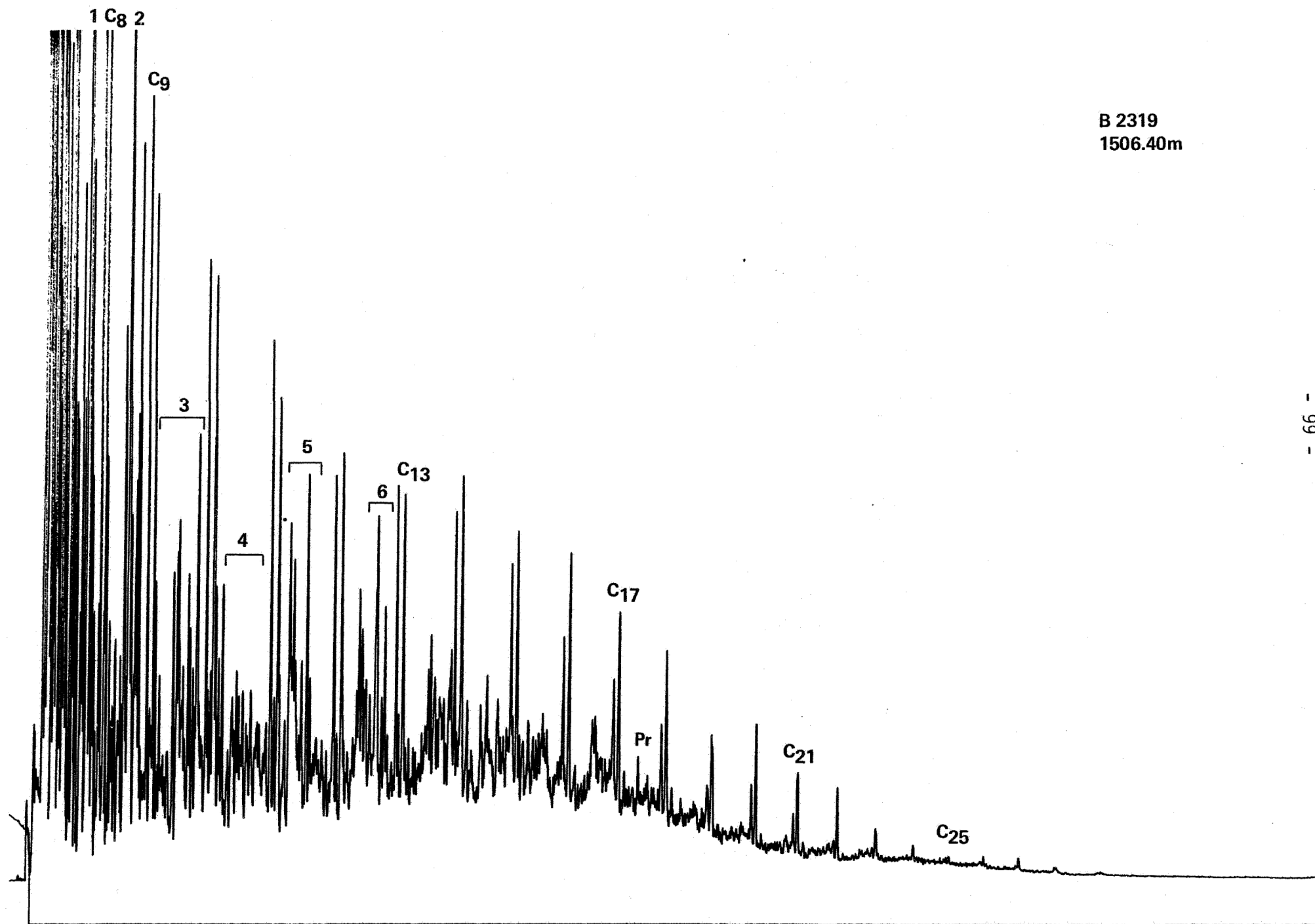
B 2321
Core
1516.35m



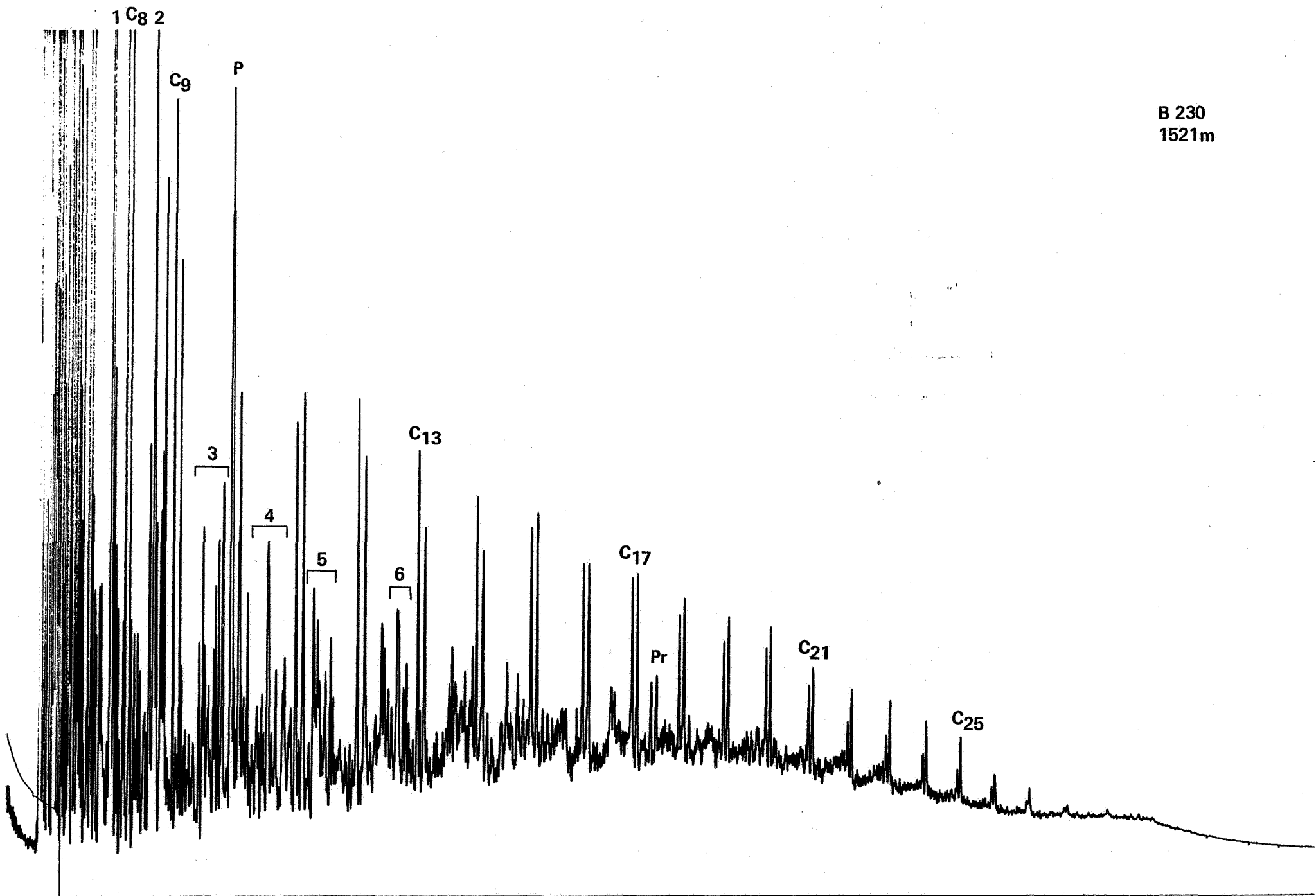
B 2320
1509.40m



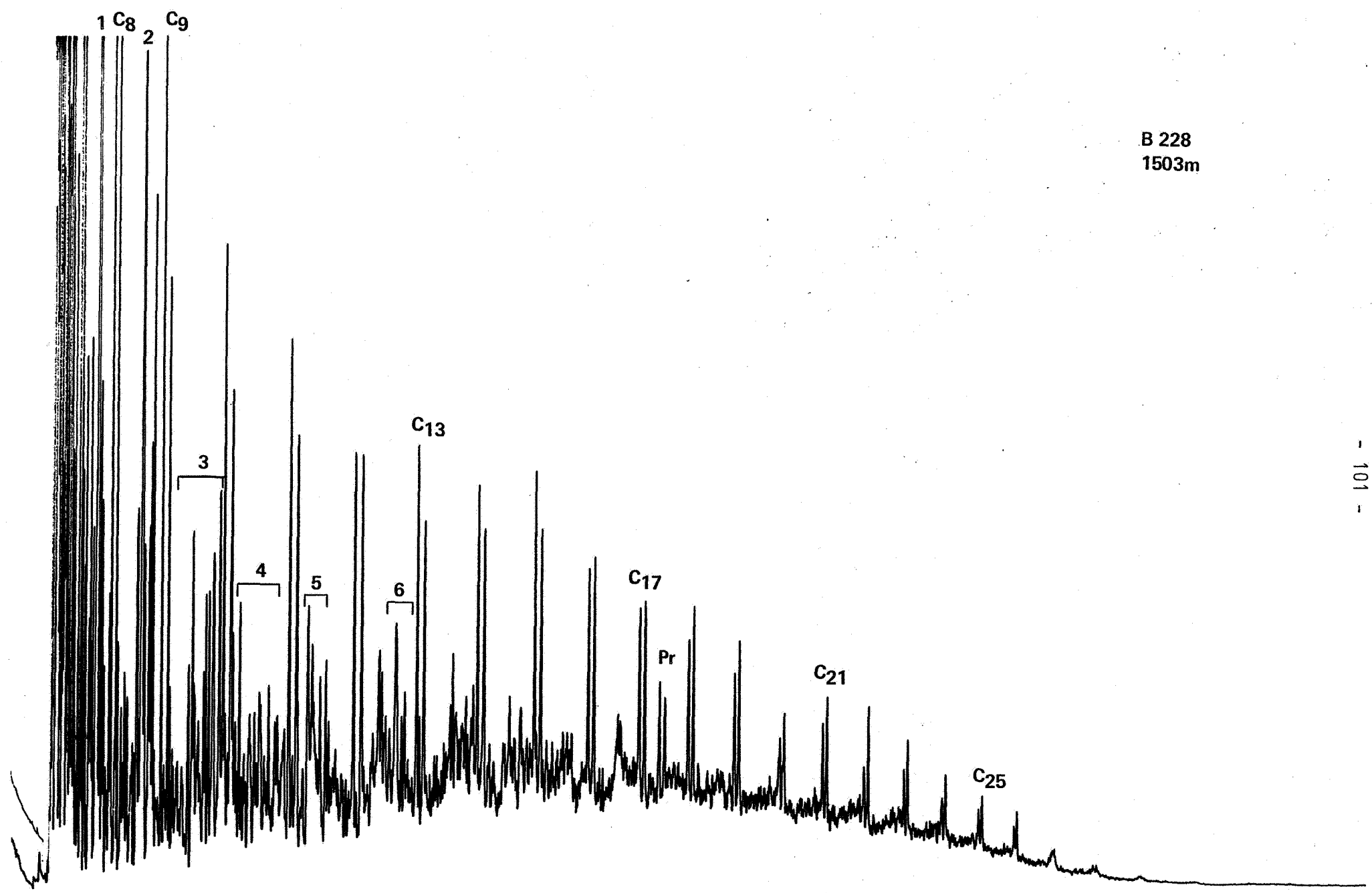
B 2319
1506.40m



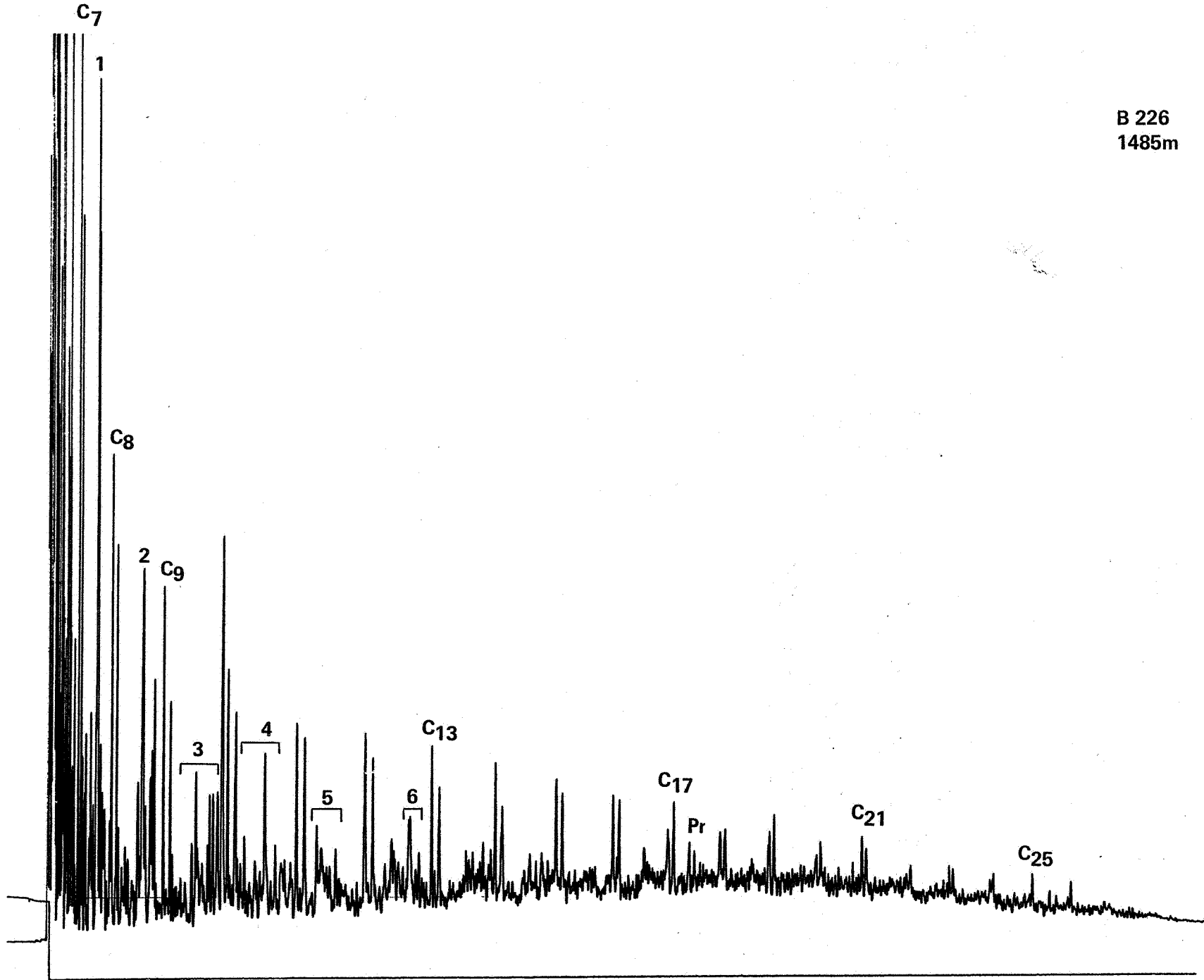
B 230
1521m



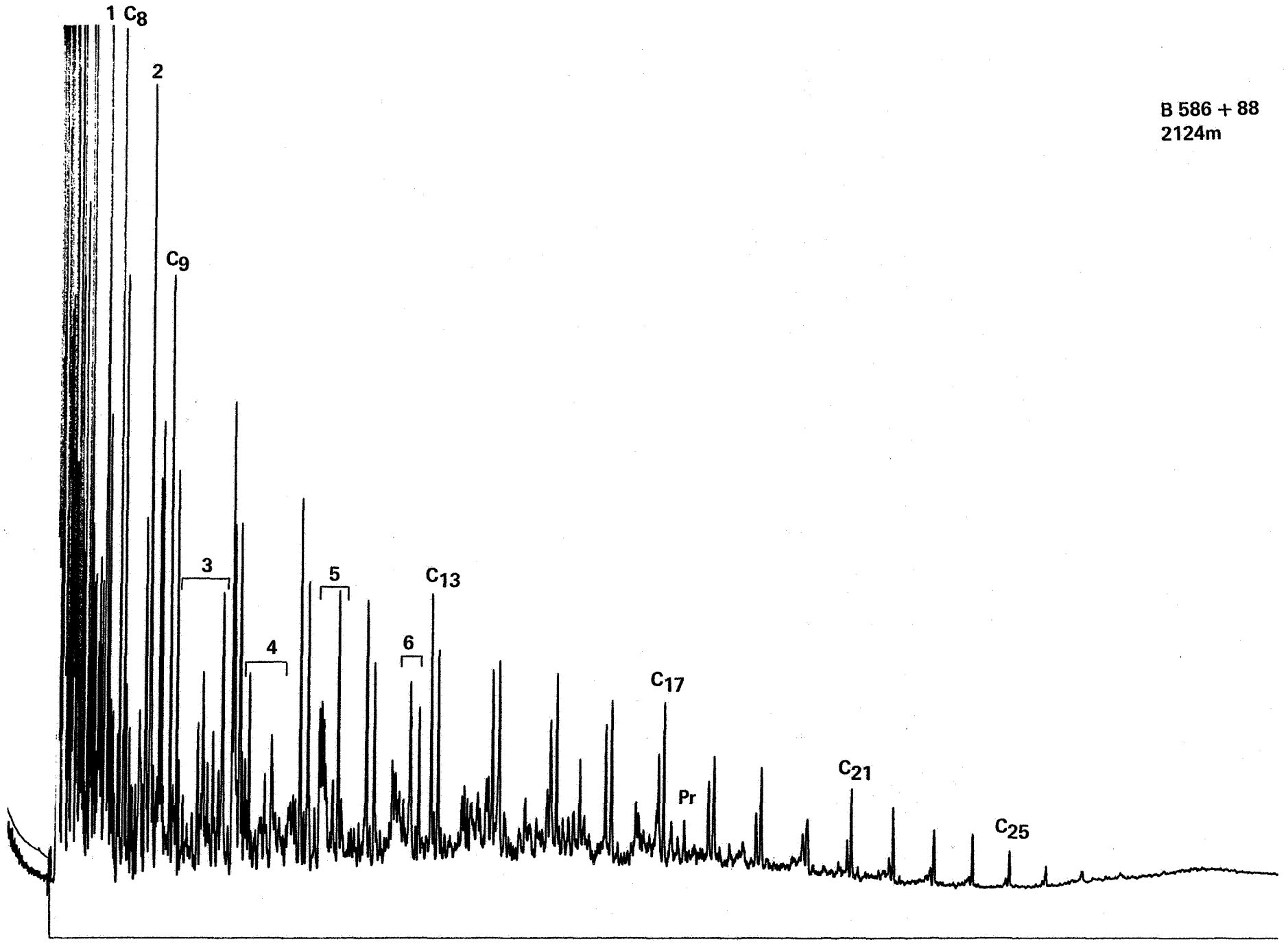
B 228
1503m



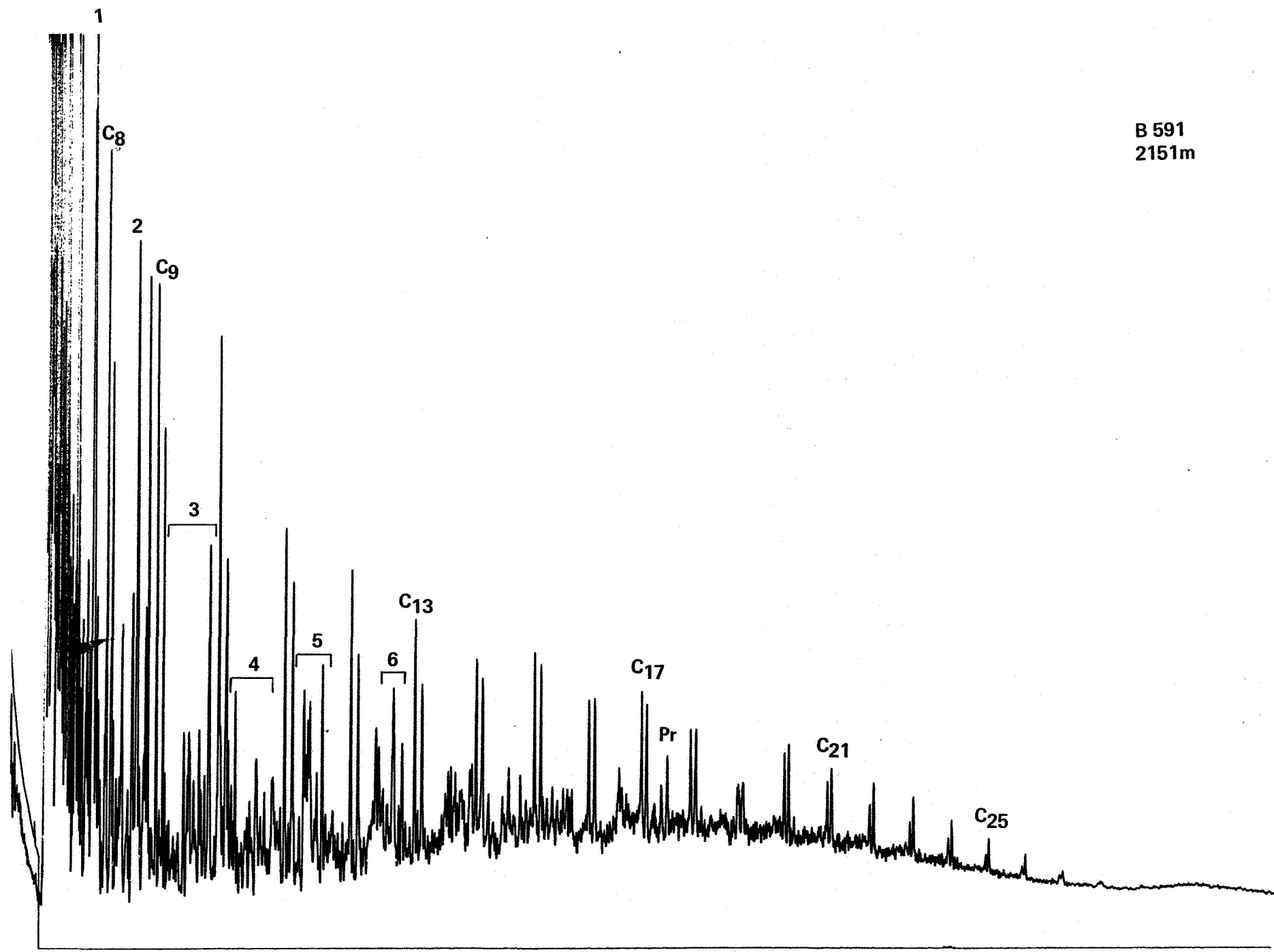
B 226
1485m



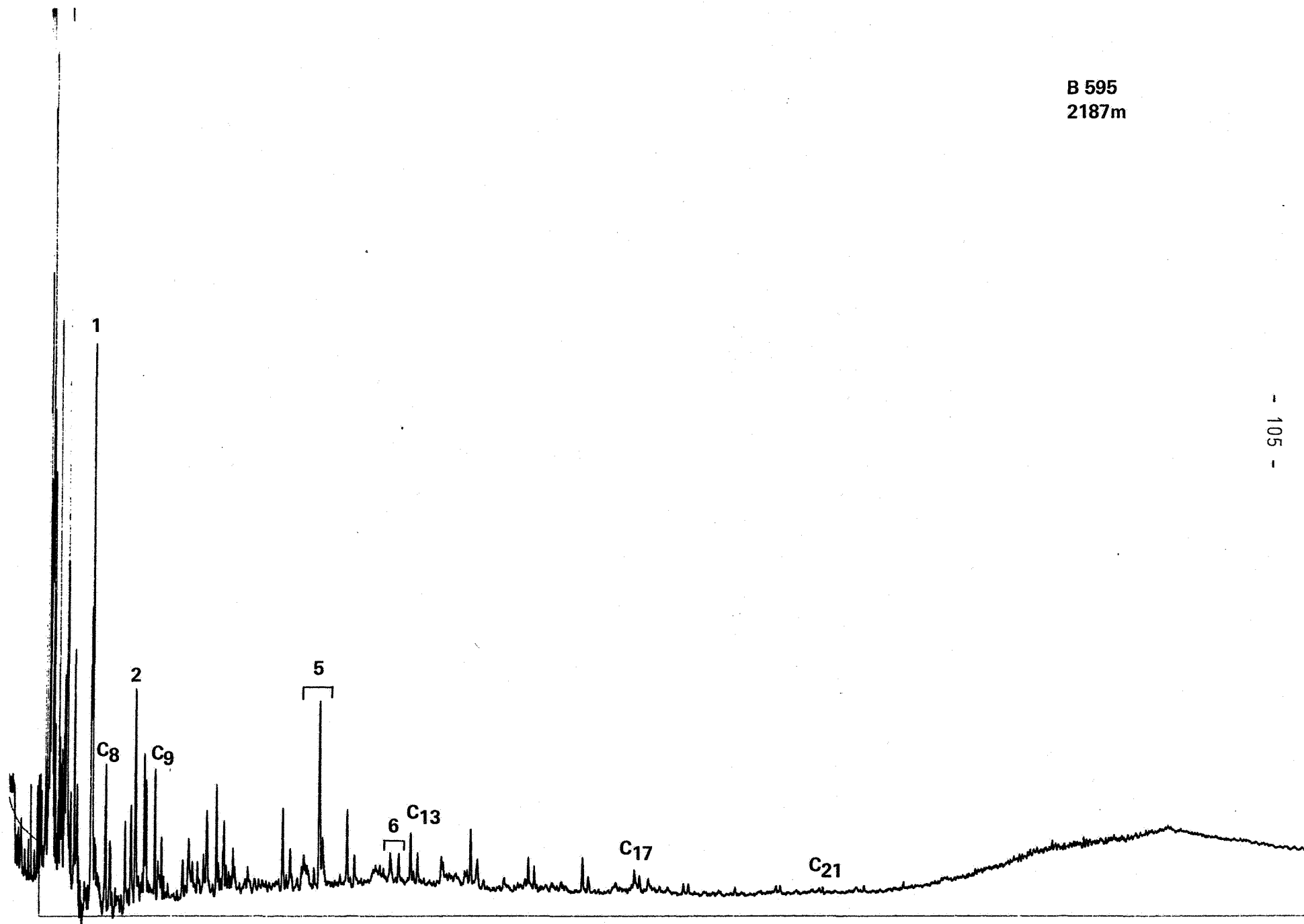
B 586 + 88
2124m



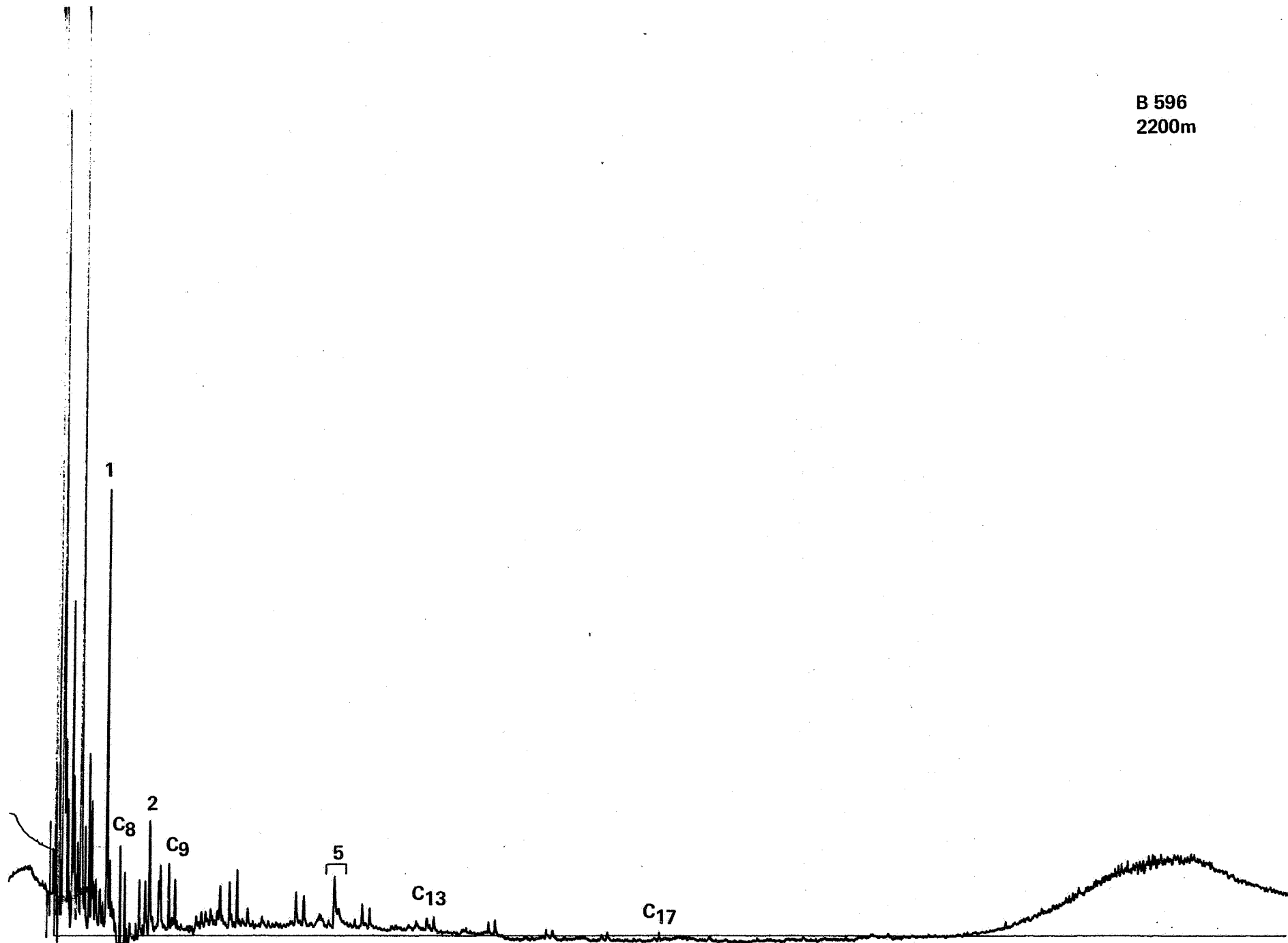
B 591
2151m



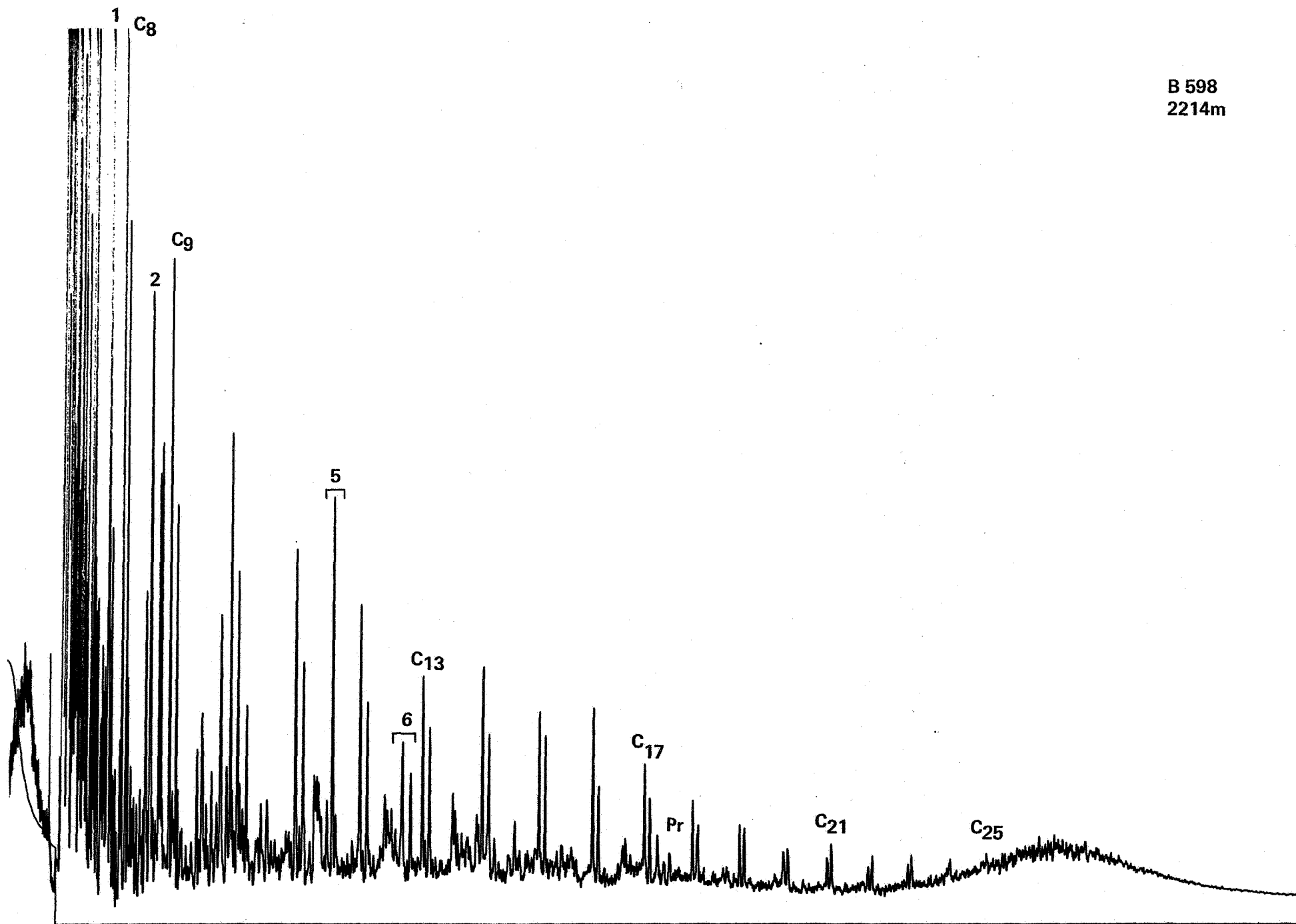
B 595
2187m



B 596
2200m

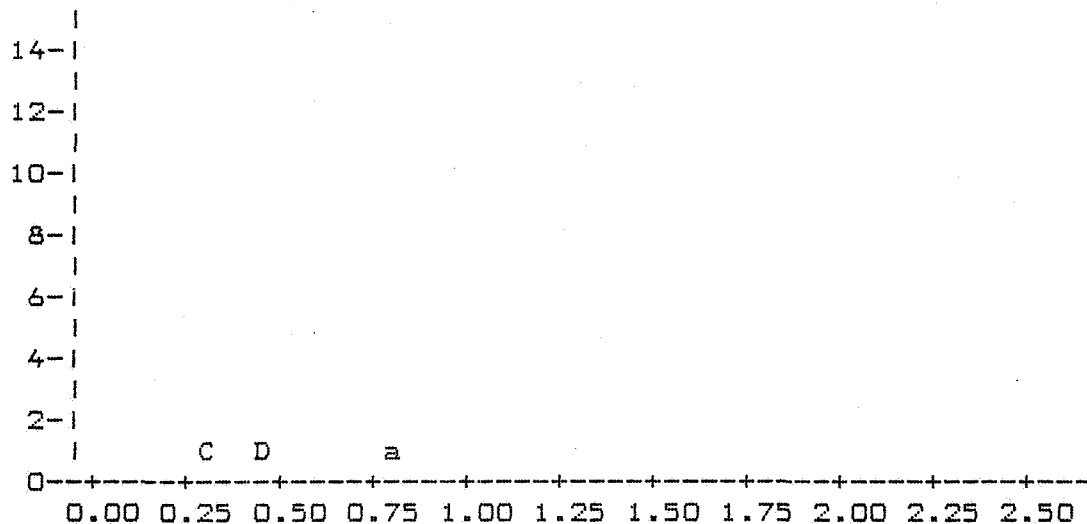


B 598
2214m



IKU# B 177 1000.OM 31/5-2

- 112 -



PP LOW HIGH LIT #VAL MEAN STDV
Y 0.32 0.47 ALL 2 0.39 0.10
OVERALL 3 0.53 0.25

ORDERED VALUES FOLLOW:

0.32C 0.46D 0.81a

14-|

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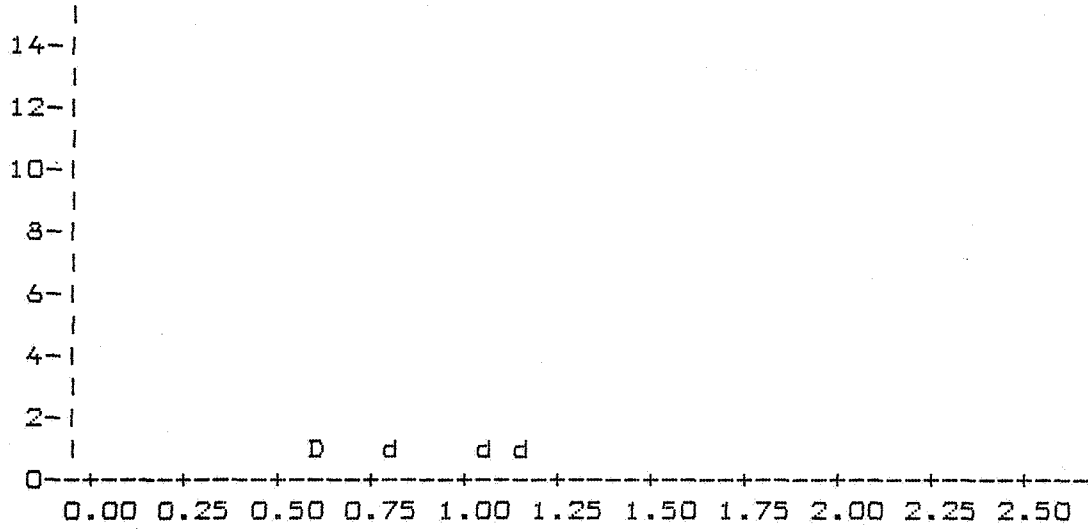
0.00 0.25 0.50 0.75 1.00 1.25 1.50 1.75 2.00 2.25 2.50

PP	LOW	HIGH	LIT	#VAL	MEAN	STDV
Y	0.32	0.40	ALL	4	0.36	0.03
			OVERALL	4	0.36	0.03

ORDERED VALUES FOLLOW:

0.32D 0.36D 0.37D 0.39D

IKU# B 214 1370.0M 31/5-2 - 115 -

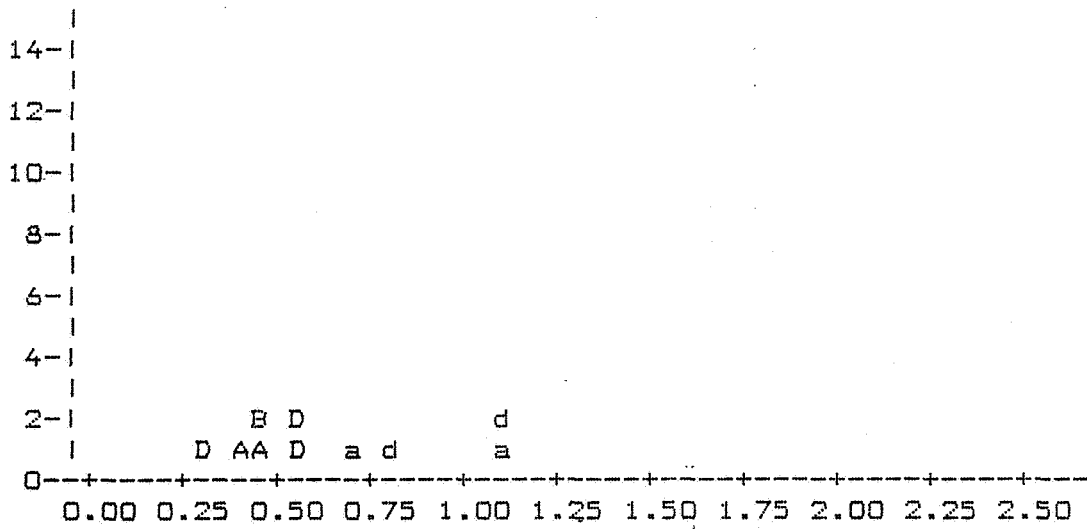


PP LOW HIGH LIT #VAL MEAN STDV
 Y 0.61 0.62 ALL 1 0.61 0.00
 OVERALL 4 0.92 0.25

ORDERED VALUES FOLLOW:

0.61D 0.82d 1.08d 1.15d

IKU# B228 1503.OM 31/5-2 - 116 -



PP LOW HIGH LIT #VAL MEAN STDV
Y 0.32 0.60 ALL 6 0.47 0.10
OVERALL 10 0.66 0.28

ORDERED VALUES FOLLOW:

0.32D 0.41A 0.45A 0.45B 0.58D 0.59D 0.72a 0.80d 1.11a 1.12d

IKU# B238

1593.OM 31/5-2

- 117 -

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0.00 0.25 0.50 0.75 1.00 1.25 1.50 1.75 2.00 2.25 2.50

PP LOW HIGH LIT #VAL MEAN STDV

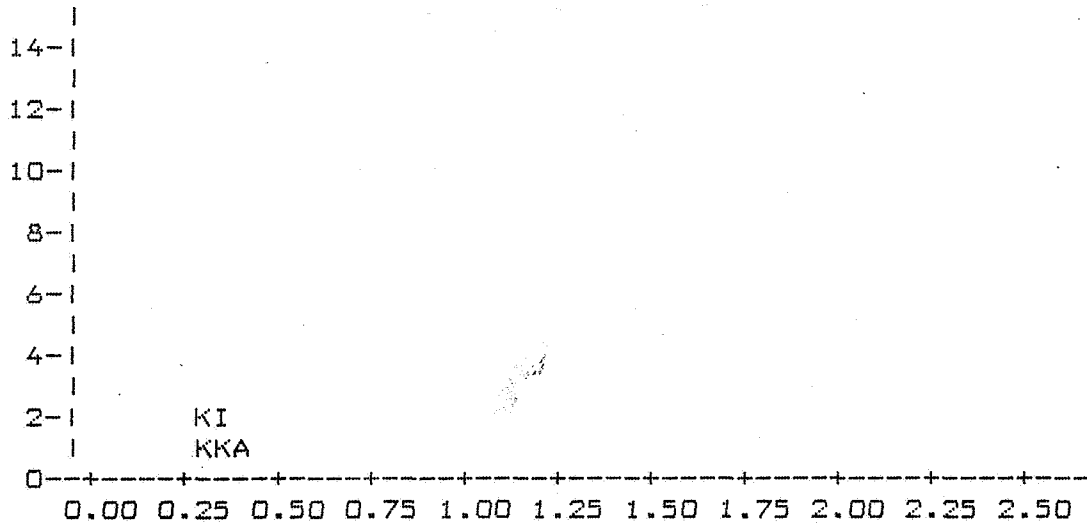
Y 0.48 0.49 ALL 1 0.48 0.00

OVERALL 6 0.63 0.21

ORDERED VALUES FOLLOW:

0.33d 0.48D 0.63b 0.63a 0.83d 0.89d

IKU# B 250 1701.0M 31/5-2 - 118 -



PP LOW HIGH LIT #VAL MEAN STDV
Y 0.31 0.41 ALL 5 0.35 0.04
OVERALL 5 0.35 0.04

ORDERED VALUES FOLLOW:

0.31K 0.32K 0.37I 0.37K 0.40A

IKU# B 551 1791.0M 31/5-2 - 119 -

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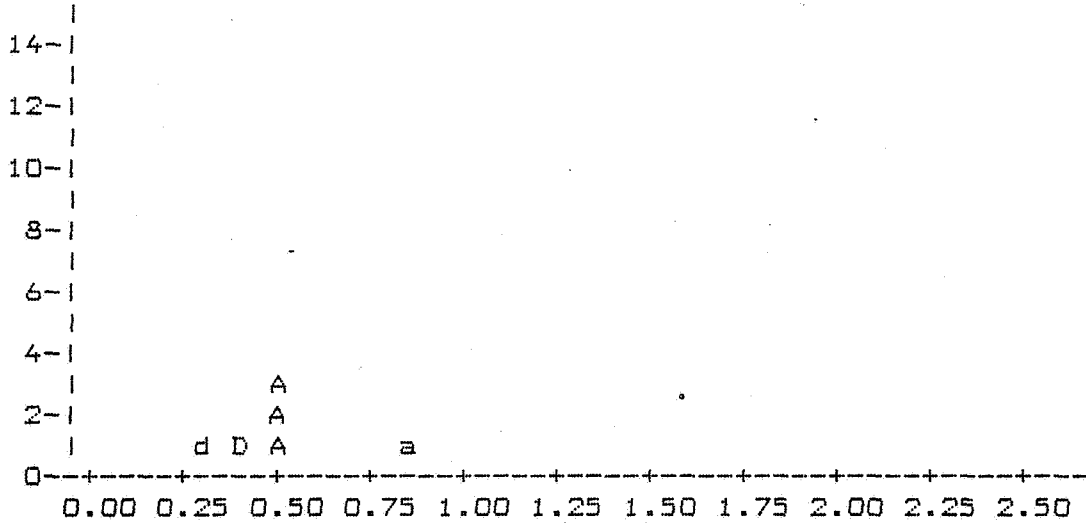
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ccDDC C c c

0.00 0.25 0.50 0.75 1.00 1.25 1.50 1.75 2.00 2.25 2.50

PP LOW HIGH LIT #VAL MEAN STDV
Y 0.38 0.57 ALL 6 0.46 0.06
OVERALL 10 0.51 0.22

ORDERED VALUES FOLLOW:

0.28c 0.32c 0.38D 0.41D 0.45D 0.46B 0.47C 0.56C 0.82c 0.97c

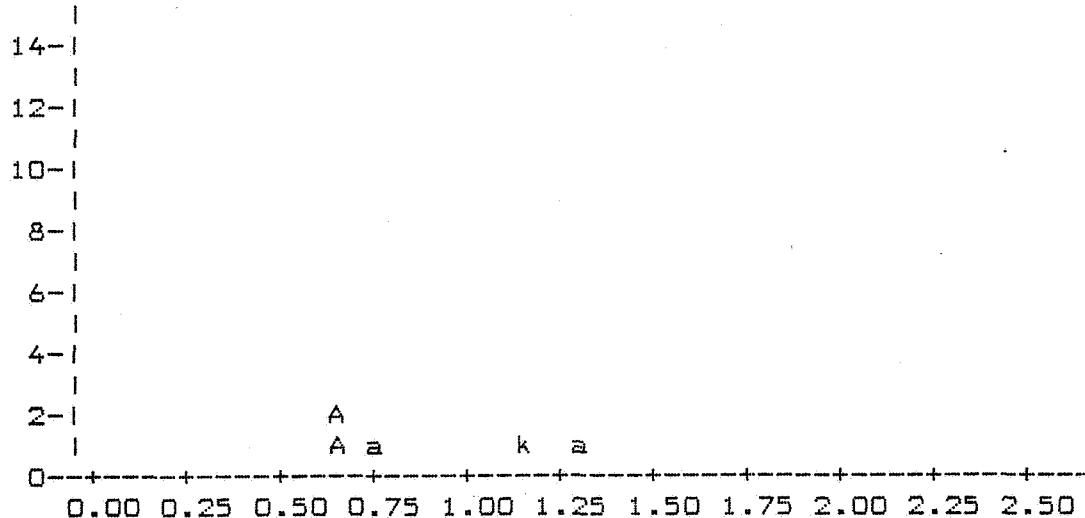


PP LOW HIGH LIT #VAL MEAN STDV
Y 0.40 0.54 ALL 4 0.48 0.06
OVERALL 6 0.52 0.18

ORDERED VALUES FOLLOW:

0.31d 0.40D 0.50A 0.50A 0.53A 0.85a

IKU# B 571 1971.OM 31/5-2 - 121 -



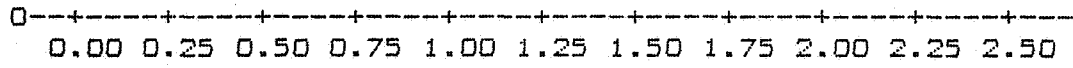
PP LOW HIGH LIT #VAL MEAN STDV
Y 0.65 0.68 ALL 2 0.66 0.01
OVERALL 5 0.92 0.30

ORDERED VALUES FOLLOW:

0.65A 0.67A 0.79a 1.16k 1.32a

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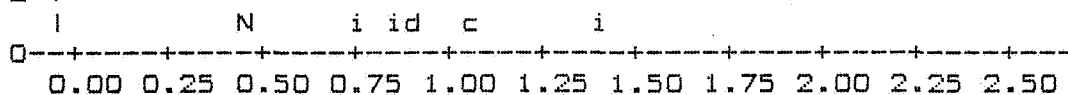


PP LOW HIGH LIT #VAL MEAN STDV
Y 0.36 0.49 ALL 5 0.41 0.05
OVERALL 10 0.61 0.29

ORDERED VALUES FOLLOW:

0.32d 0.36M 0.37A 0.41A 0.42D 0.48M 0.71m 0.92d 0.95a 1.11a

14-|
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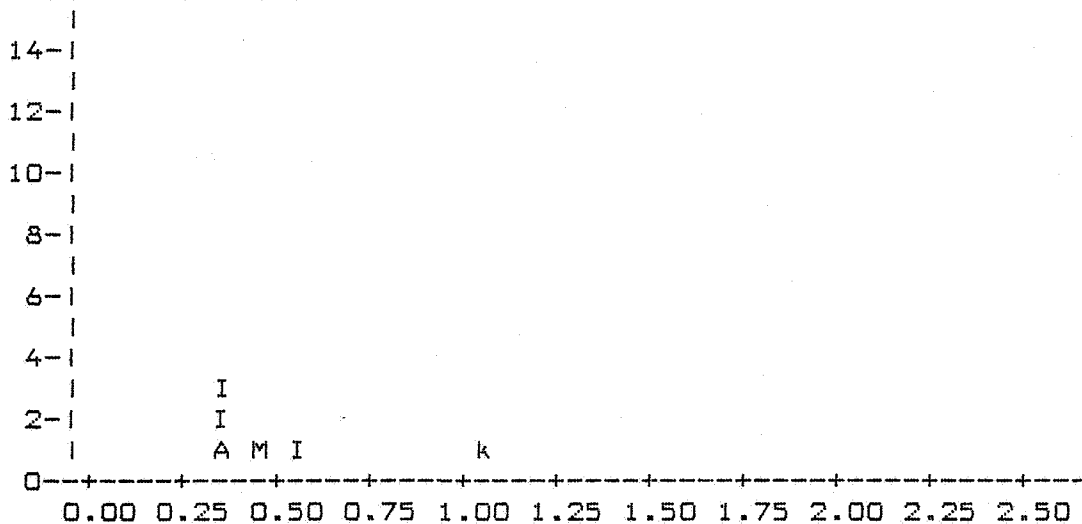
PP LOW HIGH LIT #VAL MEAN STDV
Y 0.49 0.50 ALL 1 0.49 0.00
OVERALL 6 0.92 0.31

ORDERED VALUES FOLLOW:

0.49N 0.77i 0.87i 0.90d 1.09c 1.41i

IKU# B 598 2214.OM 31/5-2

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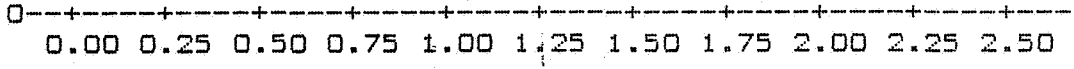
PP LOW HIGH LIT #VAL MEAN STDV
Y 0.36 0.59 ALL 5 0.44 0.09
OVERALL 6 0.54 0.27

ORDERED VALUES FOLLOW:

0.36I 0.39I 0.39A 0.48M 0.58I 1.06k

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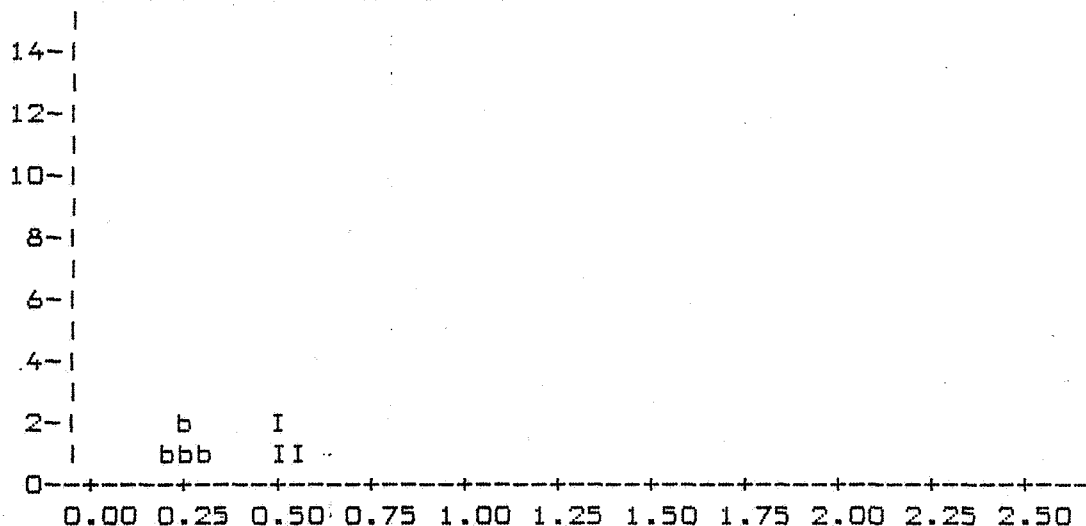
B B
b B B
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PP LOW HIGH LIT #VAL MEAN STDV
Y 0.45 0.64 ALL 7 0.54 0.07
OVERALL 9 0.50 0.11

ORDERED VALUES FOLLOW:

0.33b 0.34b 0.45D 0.48B 0.48B 0.55B 0.60D 0.60B 0.63B



PP LOW HIGH LIT #VAL MEAN STDV
 Y 0.50 0.59 ALL 3 0.53 0.04
 OVERALL 7 0.39 0.14

ORDERED VALUES FOLLOW:

0.24b 0.26b 0.29b 0.34b 0.50I 0.52I 0.58I

IKU# B 623 2439.0M 31/5-2

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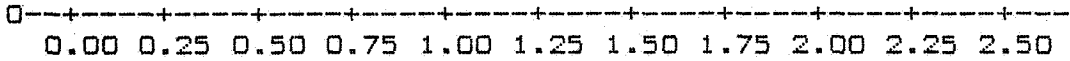
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PP LOW HIGH LIT #VAL MEAN STDV
OVERALL 2 1.05 0.08

ORDERED VALUES FOLLOW:

0.99a 1.11a

IKU# B 629 2493.0M 31/5-2

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0.00 0.25 0.50 0.75 1.00 1.25 1.50 1.75 2.00 2.25 2.50

PP	LOW	HIGH	LIT	#VAL	MEAN	STDV
Y	0.44	0.45	ALL	1	0.44	0.00
	OVERALL			1	0.44	0.00

ORDERED VALUES FOLLOW:

0.44I